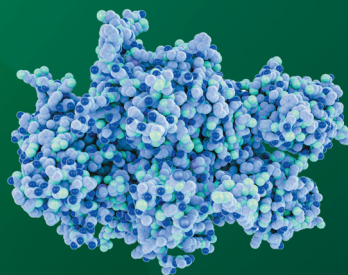
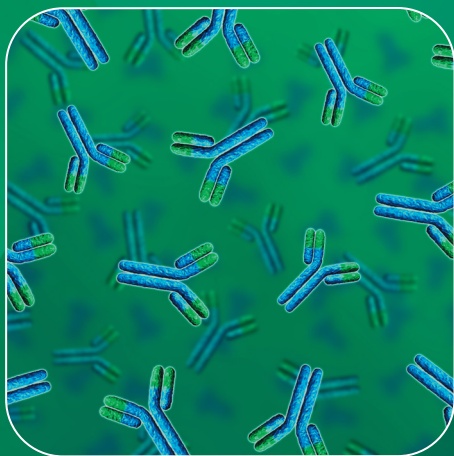


YMC

Biochromatography Columns



RP
SEC
IEX
HIC



HPLC Columns for Biochromatography

| | Reversed Phase (RP) | Size Exclusion (SEC) | Ion Exchange (IEX) | Hydrophobic Interaction (HIC) |
|----------------------|--|---|---|---|
| Separation principle | Hydrophobicity | Molecular weight | Electric charge | Hydrophobicity |
| Max. MW | Up to about 150,000 Da | Up to about 1,000,000 Da | Up to several millions Da | Up to about 1,000,000 Da |
| Resolution | +++ | ++ | +++ | +++ |
| Speed | +++ | + | ++ / +++ | +++ |
| Loading | ++ | ++ | +++ | +++ |
| Stability | + / ++ | +++ | +++ | +++ |
| Usage (e.g.) | <ul style="list-style-type: none"> • Peptide mapping • LC/MS • Nucleic acids and oligonucleotides | <ul style="list-style-type: none"> • Impurity analysis of antibody-drug conjugates • MAb separation | <ul style="list-style-type: none"> • Proteins/MAb • Charge variant analysis • Isoform analysis • Nucleic acids and oligonucleotides | <ul style="list-style-type: none"> • Drug-binding analysis of antibody-drug conjugates |

Application data mainly by courtesy of YMC Co., Ltd.

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XBridge Protein BEH, XBridge Peptide BEH are trademarks of Waters Corp.

Aeris is a trademark of Phenomenex, Inc.

Pro Pac WCX-10, MabPac HIC-10 are trademarks of Thermo Fisher Scientific.

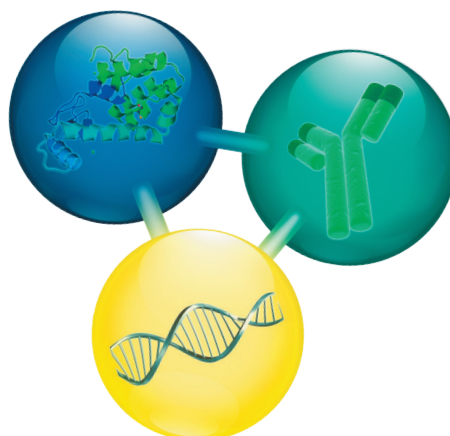
Advance Bio RP-mAb is a trademark of Agilent Technologies, Inc.

TSKgel SuperSW3000, TSKgel BioAssist Q/S, TSKgel SP-NPR, TSKgel Butyl-NPR are trademarks of Tosoh Corp.

GE Healthcare Mono Q/S is a trademark of GE Corp.

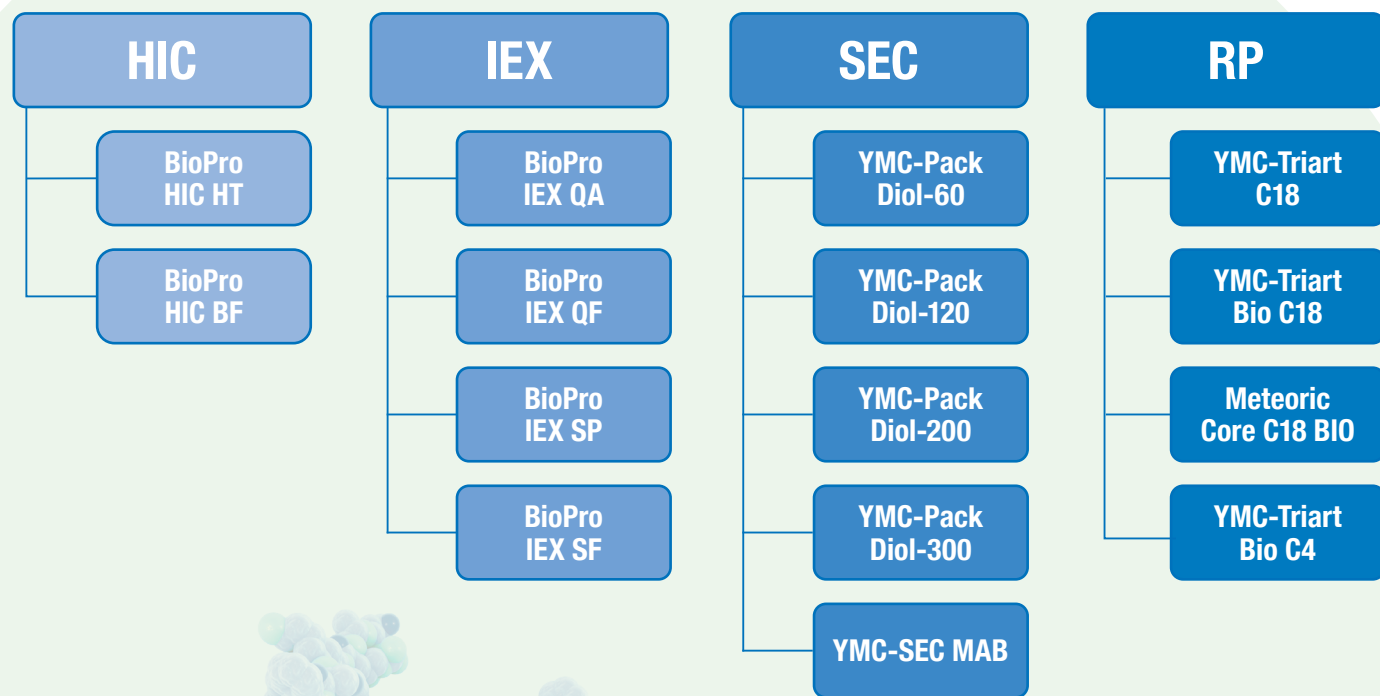
Every effort has been taken to ensure this list is accurate at the time of printing this brochure.

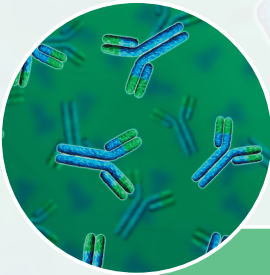
| | Page |
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| Phase selection guide..... | 04–05 |
| Method Validation Kits for BioLC..... | 06 |
| BioLC applications..... | 07–20 |
| Antibodies/ADCs..... | 08–14 |
| Proteins/Peptides..... | 15–17 |
| Oligonucleotides/Plasmids..... | 18–20 |
| Reversed Phase (RP)..... | 21–40 |
| Size Exclusion (SEC)..... | 41–50 |
| Ion Exchange (IEX)..... | 51–62 |
| Hydrophobic Interaction (HIC)..... | 63–72 |
| Substance index..... | 74–75 |



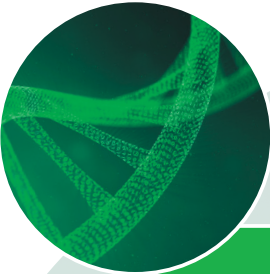
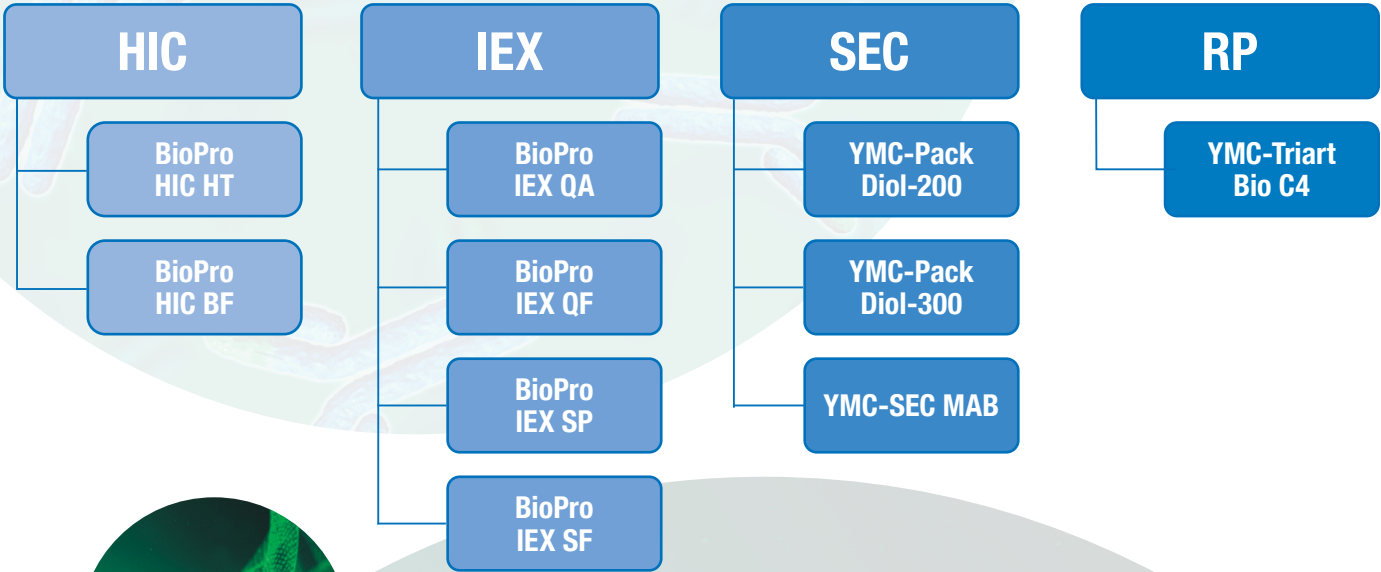


Proteins / Peptides

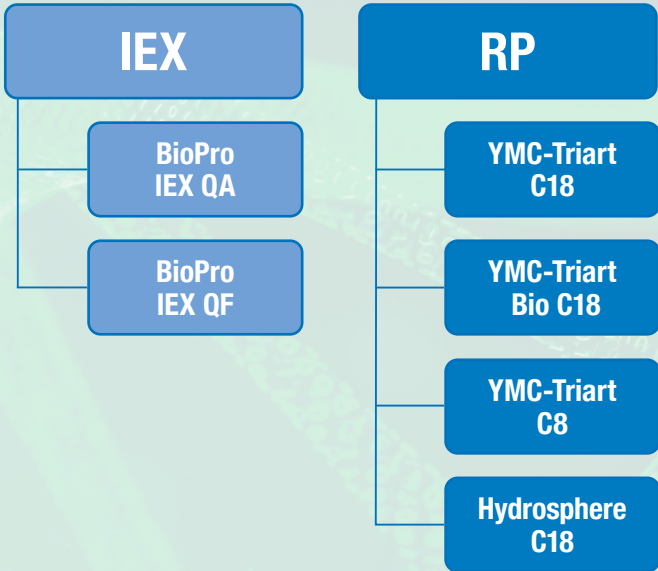




(Monoclonal) Antibodies



Oligonucleotides / Nucleic Acids



Bio QC – Validation kit

Method Validation Kits for BioLC

- for documentation of robustness and reproducibility
- three analytical columns from specified lots

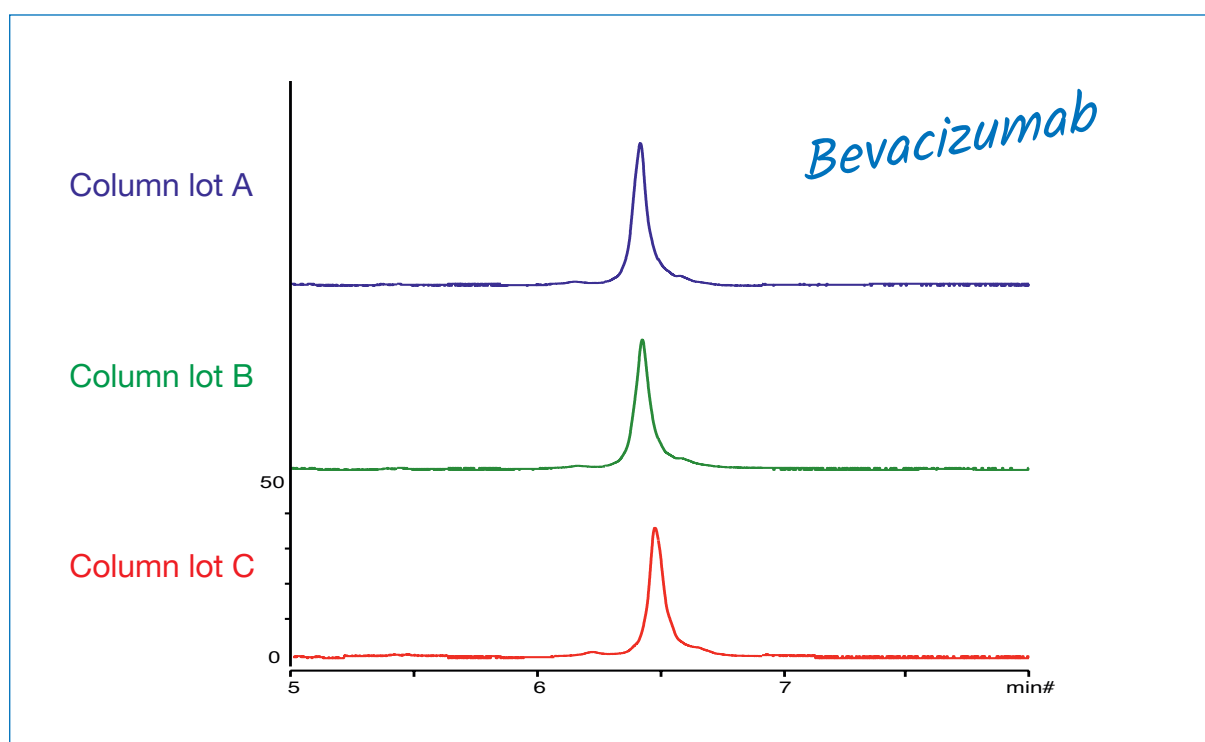
Validation kit:

contains three analytical columns packed with stationary phases from three different batches, in order to solely test the robustness of the particular method.

Available dimensions:

Length: 30 or 33, 50, 75, 100, 150, 250, 300 mm

ID: 2.0 or 2.1, 3.0, 4.0, 4.6, 8.0 mm



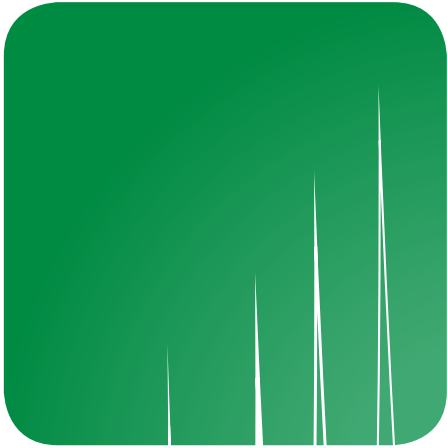
To order a validation kit simply use the ordering number for the column of interest, e.g. **TB30SP9-05Q1PT** and add V1: **TB30SP9-05Q1PTV1**.

For details on YMC selectivities and the International Product Code please refer to the specific product sections in this catalogue.

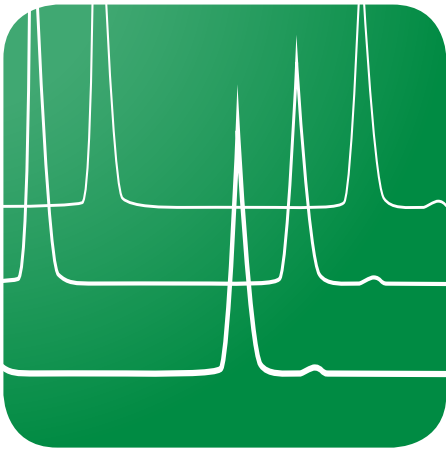
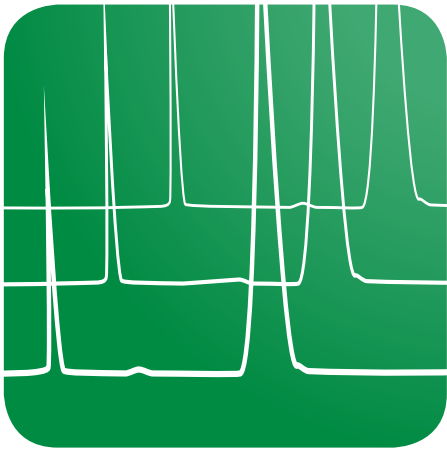
Batch Reservation Service

Occasionally, a critical analytical method may not prove as robust as you would prefer. Columns from a particular media batch may be the only way that you can, for example, isolate a critical process impurity. In such cases, YMC will reserve a specific batch of material for the use of an individual customer. YMC will also reserve prepacked columns for release according to a pre-arranged schedule.

Please call YMC or contact your YMC representative for details.



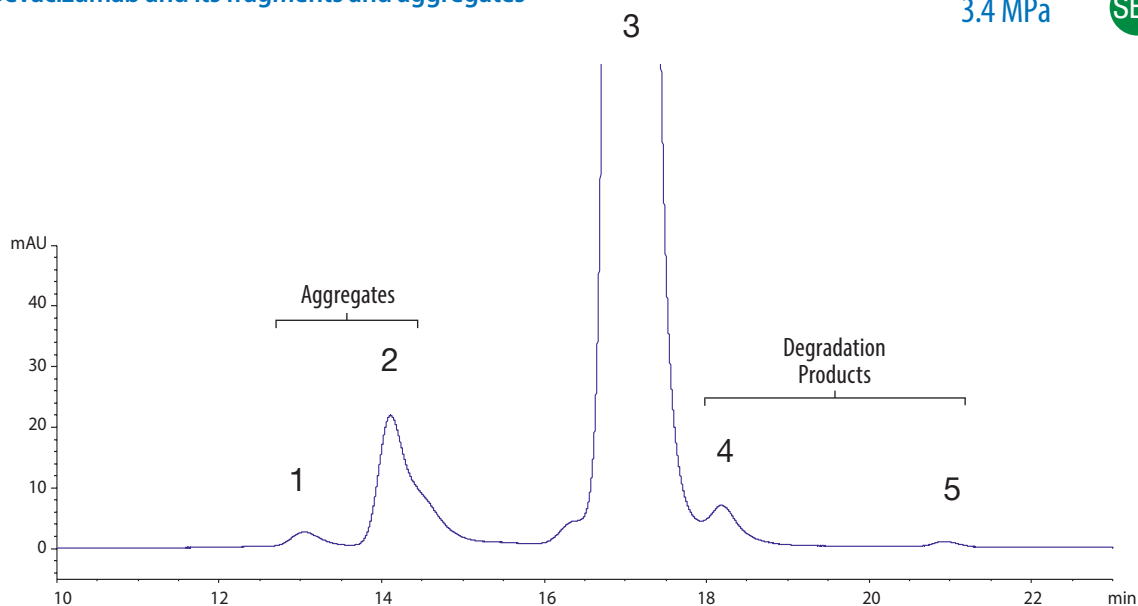
BioLC
Applications



BioLC applications – Antibodies

Bevacizumab and its fragments and aggregates

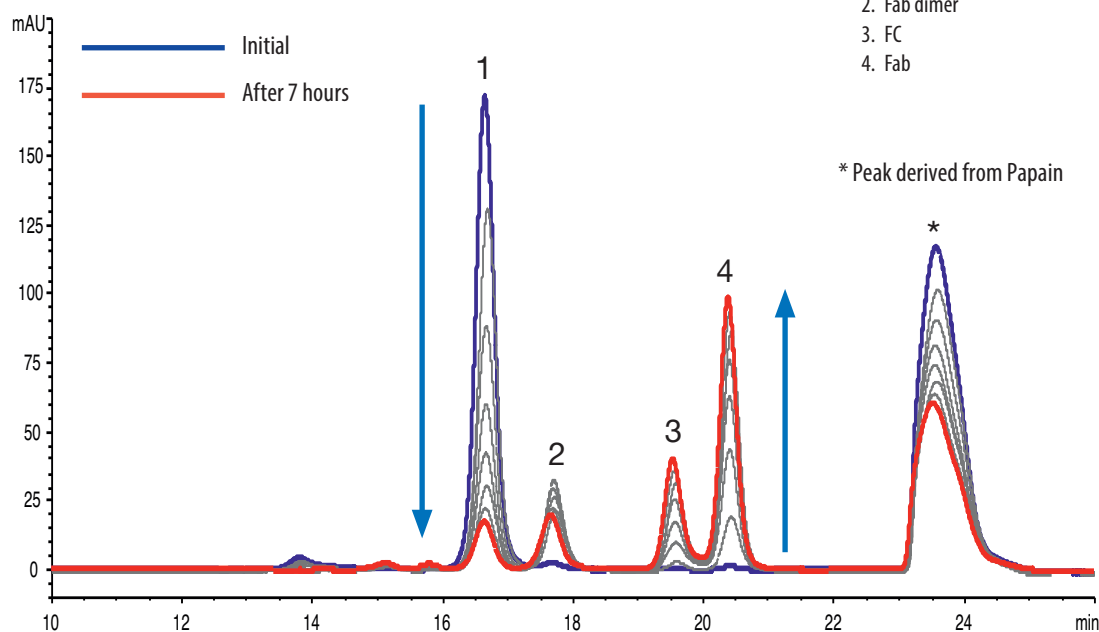
3.4 MPa



Column: YMC-SEC MAB (3 μ m, 25 nm) 300 x 4.6 mm ID
 Part No.: DLM25S03-3046WT
 Eluent: 0.1 M phosphate buffer (pH = 7) cont. 0.2 M NaCl
 Flow rate: 0.165 mL/min
 Temperature: 25 °C
 Detection: UV at 280 nm
 Cell path: 10 mm
 Injection: 10 μ L (5 mg/mL)
 Sample: Bevacizumab (Avastin®)

Analysis of digested antibody

- 1. Antibody monomer
- 2. Fab dimer
- 3. FC
- 4. Fab

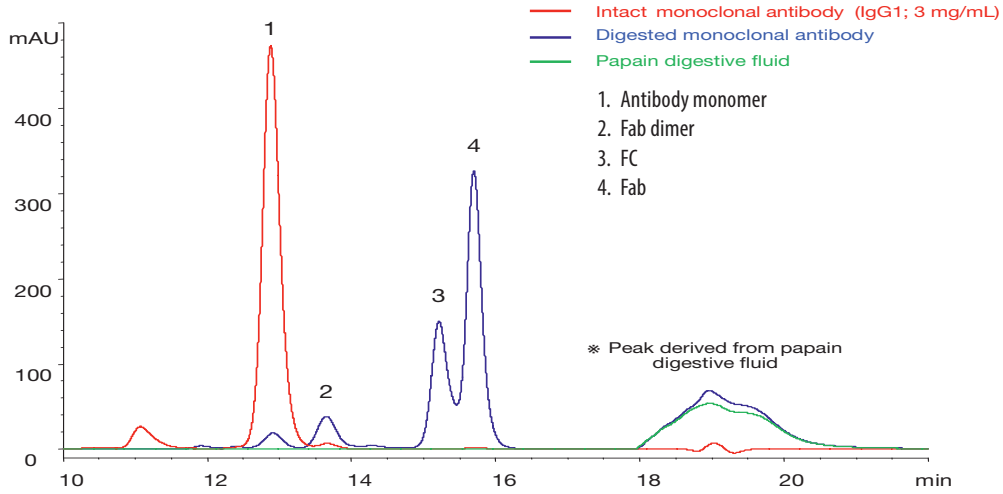


Column: YMC-SEC MAB (3 μ m, 25 nm) 300 x 4.6 mm ID
 Part No.: DLM25S03-3046WT
 Eluent: 0.1 M phosphate buffer (pH = 7) cont. 0.2 M NaCl
 Flow rate: 0.165 mL/min

Temperature: 25 °C
 Detection: UV at 280 nm
 Injection: 2 μ L (3 mg/mL)
 Sample: Humanised monoclonal IgG1 + Papain

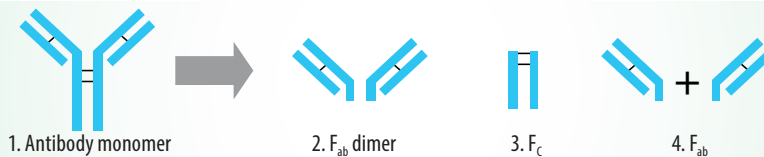
Immunglobulin digest

SEC



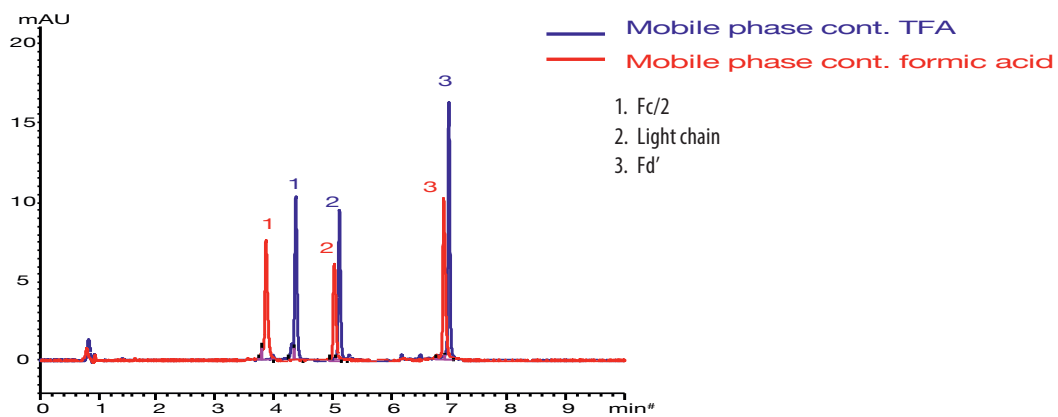
Column: YMC-Pack Diol-200 (2 μ m, 20 nm) 300 x 4.6 mm ID
 Part No.: DL20S02-3046PTH
 Eluent: 0.1 M KH_2PO_4 - K_2HPO_4 (pH 7.0) containing 0.2 M NaCl
 Flow rate: 0.2 mL/min

Temperature: ambient
 Detection: UV at 280 nm
 Sample: IgG1 (3 mg/mL)



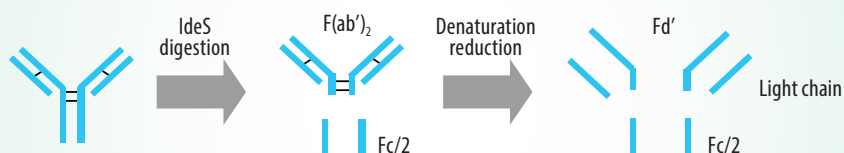
LC/MS compatible analysis of monoclonal antibody fragments

RP



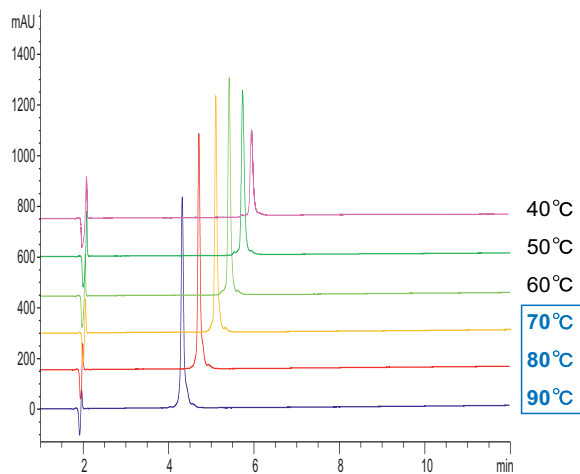
Column: YMC-Triart Bio C4 (1.9 μ m, 30 nm) 150 x 2.1 mm ID
 Part No.: TB30SP9-15Q1PT
 Eluent <TFA>: A) water/TFA (100/0.1)
 B) acetonitrile/TFA (100/0.1)
 Gradient <TFA>: 25-50%B (0-10 min), 90%B (10-12.5 min)
 Eluent <Formic acid>: A) water/formic acid (100/0.1)
 B) acetonitrile/formic acid (100/0.1)

Gradient <Formic acid>: 20-45%B (0-10 min), 90%B (10-12.5 min)
 Flow rate: 0.4 mL/min
 Temperature: 80°C
 Injection: 4 μ L (0.25 mg/mL)
 Detection: UV at 280 nm
 Sample: mAb Subunit Standard (Waters Corp.)



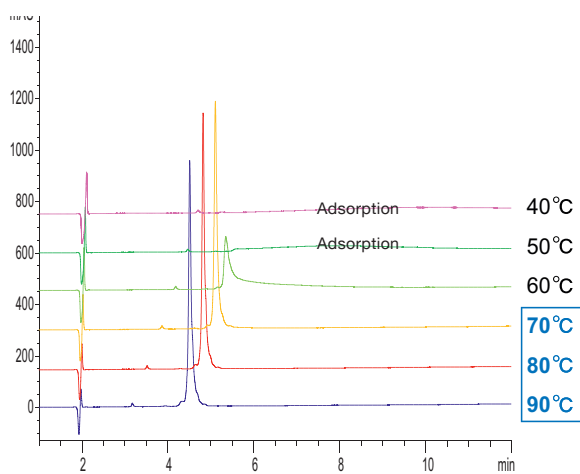
BioLC applications – Antibodies

Adalimumab (Humira®, MW: ca. 148 kDa)



Bevacizumab (Avastin®, MW: ca. 148 kDa)

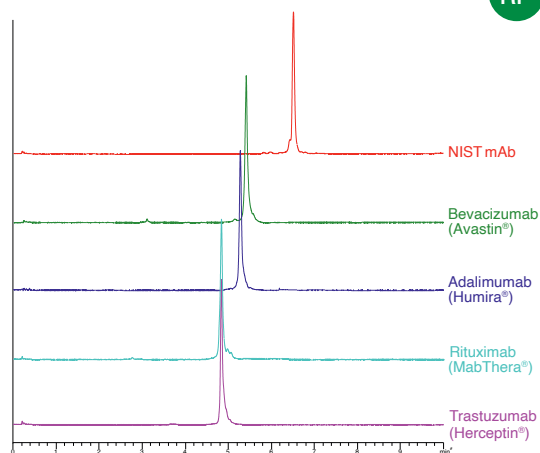
RP



Column: YMC-Triart Bio C4 (3 µm, 30 nm) 150 x 3.0 mm ID
 Part No.: TB30S03-1503PTH
 Eluent: A) water/TFA (100/0.1)
 B) acetonitrile/TFA (100/0.1)
 Gradient: 30–60%B (0–15 min), 90%B (15–30min)
 Flow rate: 0.4 mL/min
 Detection: UV at 220 nm
 Injection: 4 µL
 Sample: Adalimumab (0.5 mg/mL) or Bevacizumab (0.5 mg/mL)

Analysis of different monoclonal antibodies

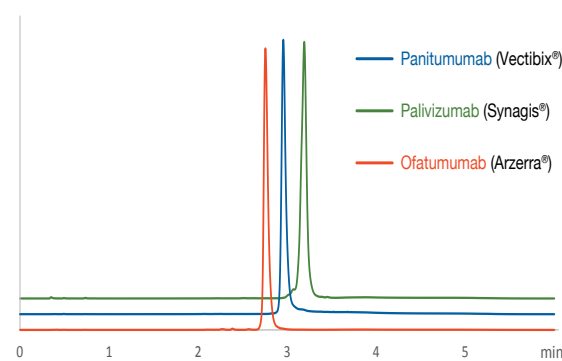
RP



Column: YMC-Triart Bio C4 (1.9 µm, 30 nm) 50 x 2.1 mm ID
 Part No.: TB30SP9-05Q1PT
 Eluent: A) water/TFA (100/0.1)
 B) acetonitrile/TFA (100/0.1)
 Gradient: 25–45%B (0–10 min)
 Flow rate: 0.4 mL/min
 Temperature: 80 °C
 Detection: UV at 280 nm (0.13s, 40Hz)
 Injection: 2 µL (0.5 mg/mL)

Analysis of challenging monoclonal antibodies

RP



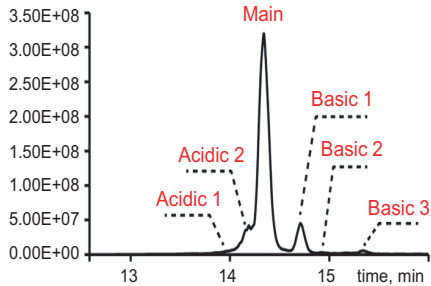
Column: YMC-Triart Bio C4 (1.9 µm, 30 nm) 50 x 2.1 mm ID
 Part No.: TB30SP9-05Q1PT
 Eluent: A) 0.1 % TFA in water
 B) 0.1 % TFA in acetonitrile
 Gradient: 25–50%B (0–4 min)
 Flow rate: 0.4 mL/min
 Temperature: 90 °C
 Detection: Fluorescence: ex 280 nm, em 350 nm
 Injection: 0.5 µL

By courtesy of University of Geneva, Institute of Pharmaceutical Sciences of Western Switzerland (ISPSO)

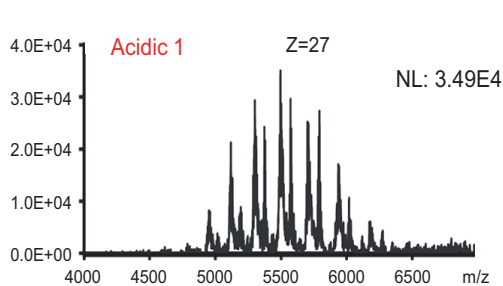
Native online SCX-MS analysis of monoclonal antibodies



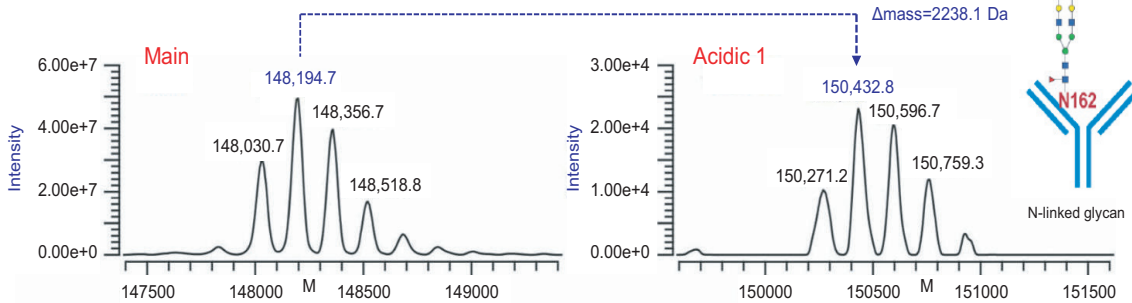
[TIC, native NISTmAb]



[Raw mass spectrum]



[Deconvoluted mass spectra]



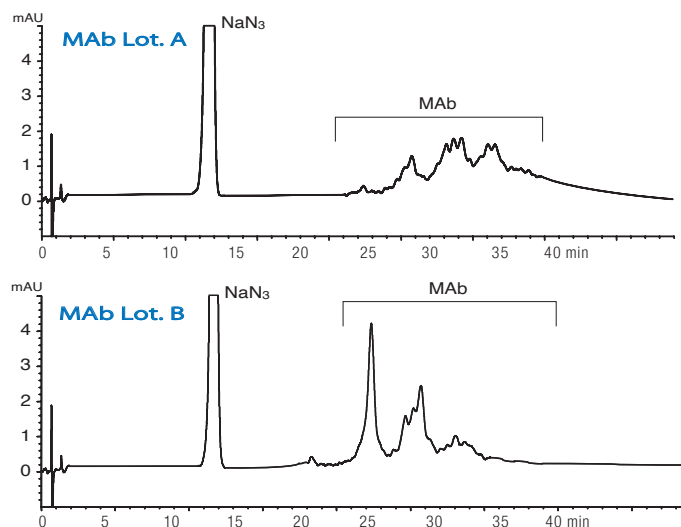
Column: BioPro IEX SF (5 μm) 100 x 4.6 mm ID
 Part No.: SF00S05-1046WP
 Eluent: A) 20 mM CH₃COONH₄-CH₃COOH (pH 5.6)
 B) 140 mM CH₃COONH₄-10 mM NH₄HCO₃ (pH 7.4)
 Gradient: 0%B (0–2 min), 0–100%B (2–18 min), 100%B (18–22 min)
 Flow rate: 0.4 mL/min
 (To enable online simultaneous UV and MS detection,
 a post-column analytical splitter (~400:1 ratio) was connected)

Temperature: 45 °C
 Detection: nanospray ionization-mass spectrometry (NSI-MS)
 Load: 50 μg
 System: LC) ACQUITY UPLC I-Class system (Waters)
 MS) Exactive™ Plus EMR mass spectrometer
 (Thermo Fisher Scientific)

Courtesy of S. Wang, Regeneron Pharmaceuticals Inc.

Reference: Y. Yan, A. P. Liu, S. Wang, T. J. Daly and N. Li, Ultrasensitive Characterization of Charge Heterogeneity of Therapeutic Monoclonal Antibodies, Anal. Chem., 2018, 90, 13013-20.

Different production batches of IgG1

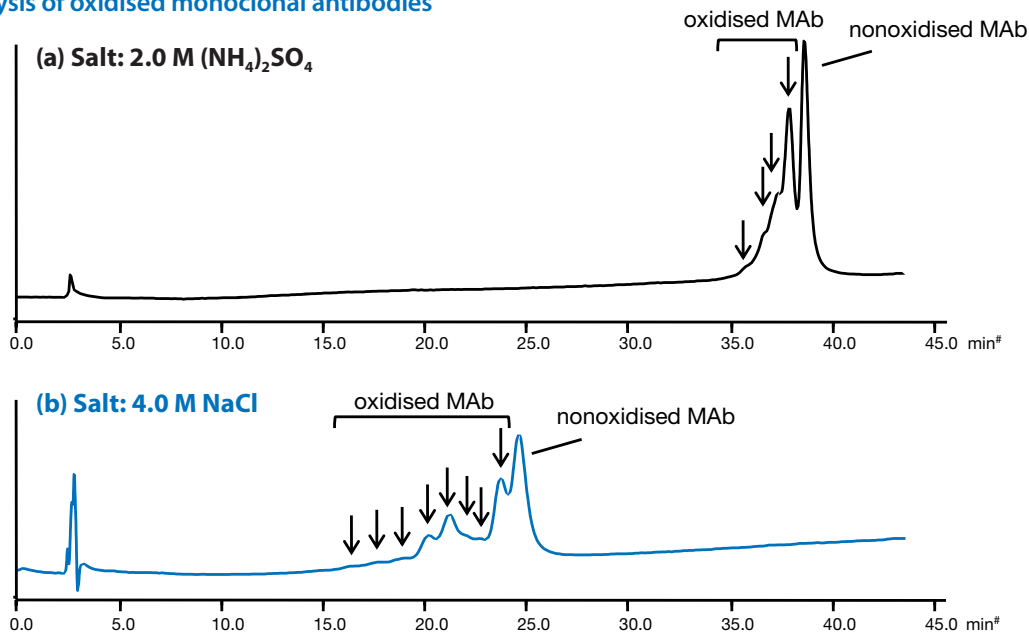


Column: BioPro IEX QF (5 μm) 100 x 4.6 mm ID
 Part No.: QF00S05-1046WP
 Eluent: A) 20 mM Tris-HCl (pH 8.1)
 B) 20 mM Tris-HCl (pH 8.1) containing 0.5 M NaCl
 Gradient: 10–25%B (0–60 min)
 Flow rate: 1.0 mL/min (360 cm/hr)

Temperature: 25 °C
 Detection: UV at 220 nm
 Injection: 14 μL (0.1 mg/mL)
 Sample: Mouse monoclonal IgG1 anti-human IgG4
 (Purified by DEAE chromatography, containing NaN₃)

BioLC applications – Antibodies

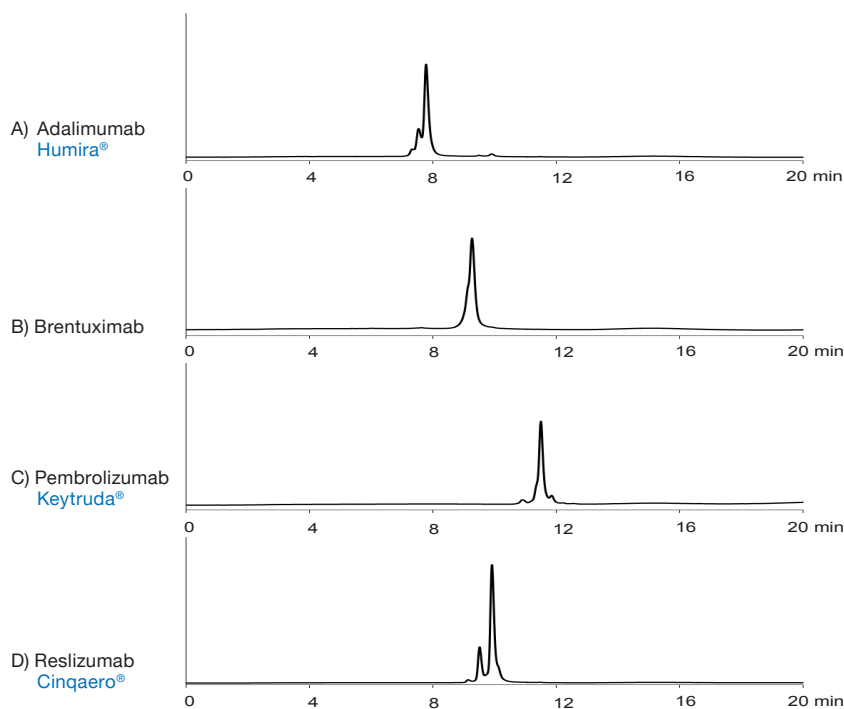
Analysis of oxidised monoclonal antibodies



HIC

| | | | |
|-----------|---|--------------|------------------|
| Column: | BioPro HIC BF (4 μm) 100 x 4.6 mm ID | Flow rate: | 0.3 mL/min |
| Part No.: | BHB00S04-1046WT | Temperature: | 25 °C |
| Eluent: | A) 100 mM NaH ₂ PO ₄ -Na ₂ HPO ₄ (pH 7.0) containing salt | Detection: | UV at 280 nm |
| Gradient: | B) 100mM NaH ₂ PO ₄ -Na ₂ HPO ₄ (pH 7.0) | Injection: | 5 μL (1.0 mg/mL) |
| | 40–80%B (0–40 min), 80%B (40–45 min) | Sample: | oxidised NISTmAb |

HIC analysis of different monoclonal antibodies using isopropanol as modifier



HIC

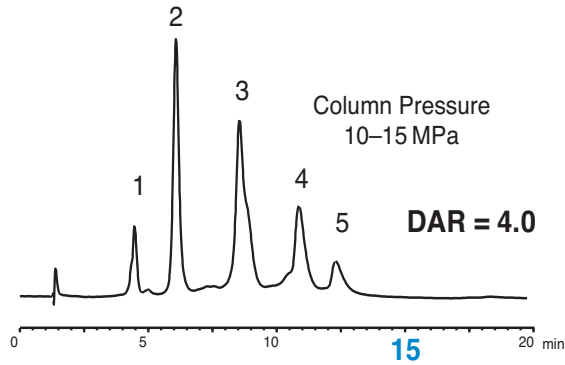
| | | | |
|------------|---|--------------|----------------------------------|
| Column: | BioPro HIC BF (4 μm) 100 x 4.6 mm ID | Temperature: | 20 °C |
| Part No.: | BHB00S04-1046WT | Detection: | Fluorescence: ex 280nm, em 360nm |
| Eluent: | A) 20 mM NaH ₂ PO ₄ -Na ₂ HPO ₄ (pH 7.4) containing 1.5 M (NH ₄) ₂ SO ₄ | Injection: | 3 μL (2 mg/mL) |
| Gradient: | B) 20 mM NaH ₂ PO ₄ -Na ₂ HPO ₄ (pH 7.4) / 2-propanol (85/15) | | |
| Flow rate: | 0–100%B (0–20 min) | | |
| | 1.0 mL/min | | |

By courtesy of University of Geneva.
Institute of Pharmaceutical Sciences of Western Switzerland (ISPSO)

BioLC applications – Antibody-Drug-Conjugates

High throughput DAR determination by shortening analysis time

HIC

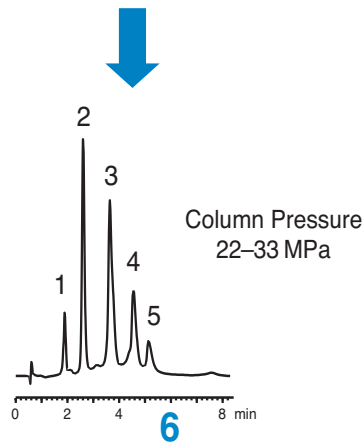


1. DAR 0
2. DAR 2
3. DAR 4
4. DAR 6
5. DAR 8

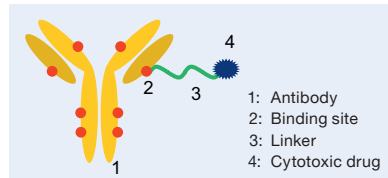
Flow rate
0.5 mL/min

2.5 x faster

Flow rate
1.2 mL/min



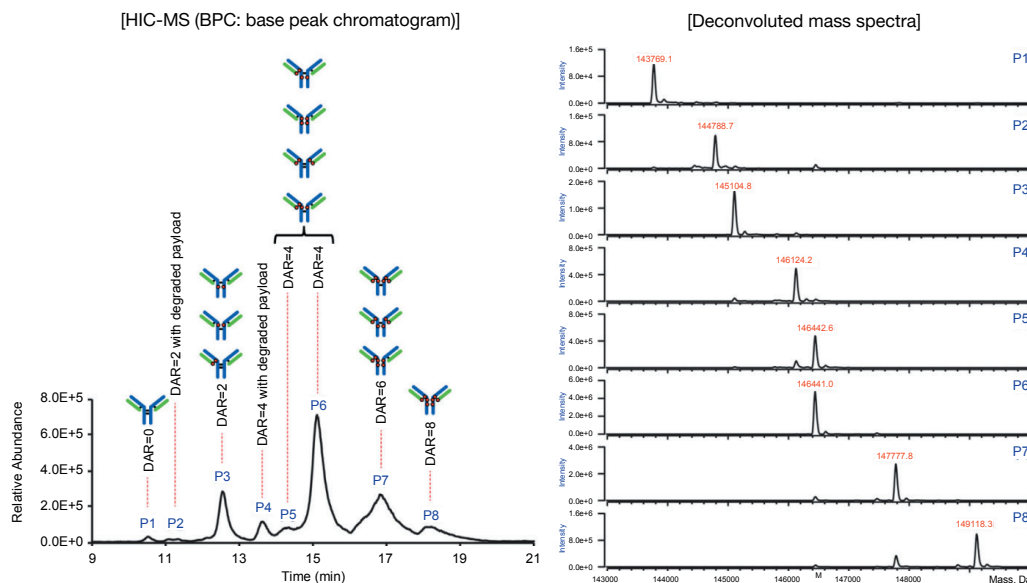
Column: BioPro HIC HT (2.3 μ m) 100 x 4.6 mm ID
 Part No.: BHH00SQ3-1046PTH
 Eluent: A) 20 mM NaH_2PO_4 - Na_2HPO_4 (pH 7.0) containing 1.0 M $(\text{NH}_4)_2\text{SO}_4$
 B) 20 mM NaH_2PO_4 - Na_2HPO_4 (pH 7.0)/2-propanol (85/15)
 Gradient: 0–100%B (0–15 min), 100%B (15–20 min)
 0–100%B (0–6.25 min), 100%B (6.25–8.3 min)
 Temperature: 25 $^\circ\text{C}$
 Detection: UV at 280 nm
 Injection: 10 μL
 Sample: Brentuximab vedotin (Adcetris[®]) (2.5 mg/mL)



BioLC applications – Antibody-Drug-Conjugates

Native online HIC-MS analysis of cys-linked ADCs

HIC



Column: BioPro HIC BF (4 μ m) 100 x 4.6 mm ID
 Part number: BHB00S04-1046WT
 Eluent: A) 3 M ammonium acetate in water
 B) 2-propanol/water (30/70)
 Gradient: 10%B (0–2 min), 10–97%B (2–18 min), 97%B (18–22 min)
 Flow rate: 0.3 mL/min
 Temperature: ambient
 Detection: UV at 280 nm, NSI-MS

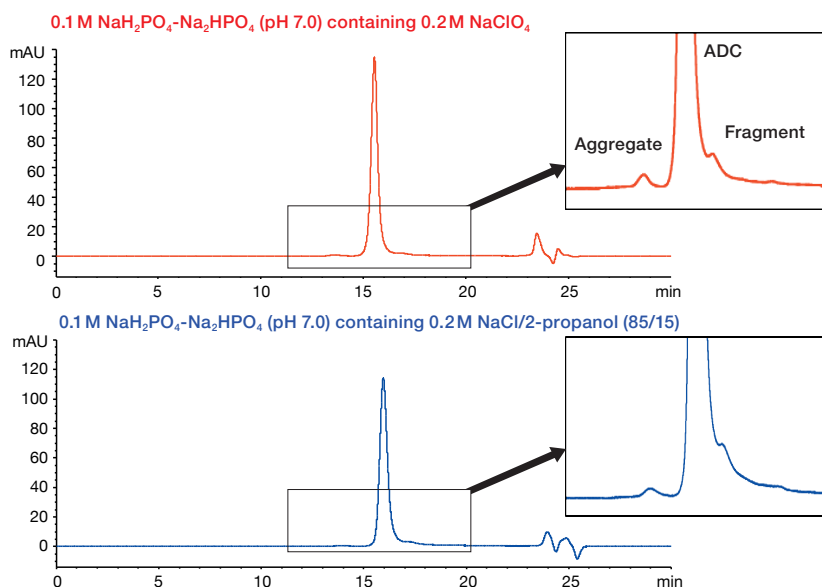
Injection: 10 μ g
 Sample: SigmaMAB ADC-mimic
 Setup: Post-column makeup flow:
 100% water at 1.5 mL/min (reducing salt conc. 6-fold)
 Splitter to reduce the flow rate to 1–5 μ L/min

Courtesy by S. Wang, Regeneron Pharmaceuticals Inc.

Reference: Y. Yan, T. Xing, S. Wang, T. J. Daly, N. Li, Online coupling of analytical hydrophobic interaction chromatography with native mass spectrometry for the characterization of monoclonal antibodies and related products, *J. Pharm. Biomed. Anal.* 186 (2020) 113313.

Separation of Brentuximab vedotin from its aggregates and fragments

SEC



Column: YMC-SEC MAB (3 μ m, 25 nm) 300 x 4.6 mm ID
 Part No.: DLM25S03-3046WT
 Eluent: 0.1 M phosphate buffer (pH = 7) cont. 0.2 M NaClO₄
 0.1 M phosphate buffer (pH = 7) cont. 0.2 M NaCl/2-propanol (85/15)
 Flow rate: 0.165 mL/min

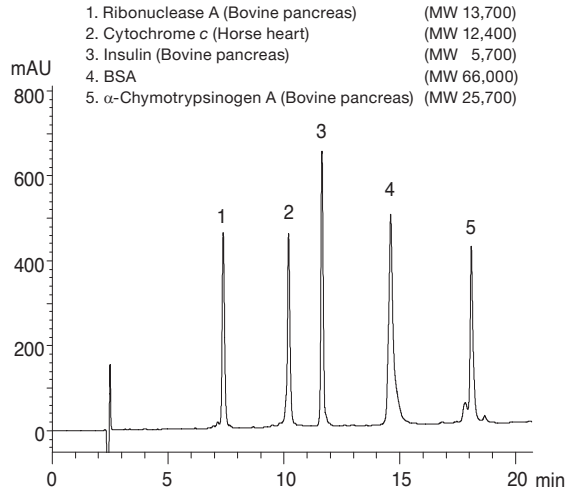
Temperature: 25 °C
 Detection: UV at 280 nm
 Injection: 4 μ L (2.5 mg/mL)
 Sample: Brentuximab vedotin (Adcetris®) for injection

Courtesy of Prof. S. Manabe, Hoshi University, Tokyo/Tohoku University, Sendai Japan.

Reference: H. Wang, M. S. Levi, A. V. Del Grosso, W. M. McCormick, L. Bhattacharyya, An improved size exclusion-HPLC method for molecular size distribution analysis of immunoglobulin G using sodium perchlorate in the eluent, *J. Pharm. Biomed. Anal.*, 138 (2017) 330-343.

Proteins (MW 5,700 ~ 66,000)

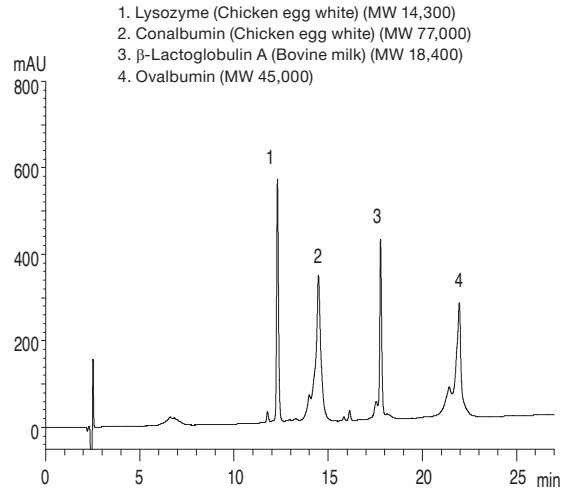
RP



Column: YMC-Triart Bio C4 (5 μ m, 30 nm) 150 x 3.0 mm ID
 Part No.: TB30S03-1503PTH
 Eluent: A) water/TFA (100/0.1)
 B) acetonitrile/TFA (100/0.1)
 Gradient: 20-60%B (0-27 min), 90%B (27-35 min)
 Flow rate: 0.4 mL/min
 Temperature: 70 °C
 Detection: UV at 220 nm
 Injection: 10 μ L (0.25-0.50 mg/mL)

Proteins (MW 14,300 ~ 77,000)

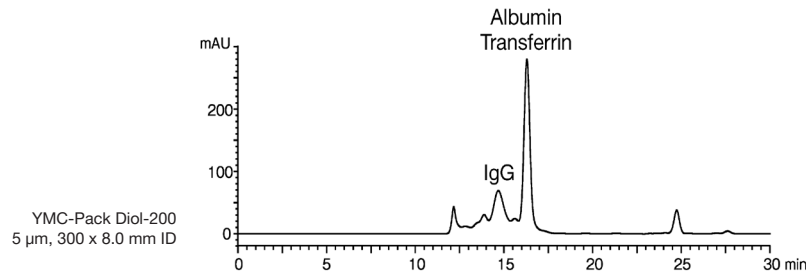
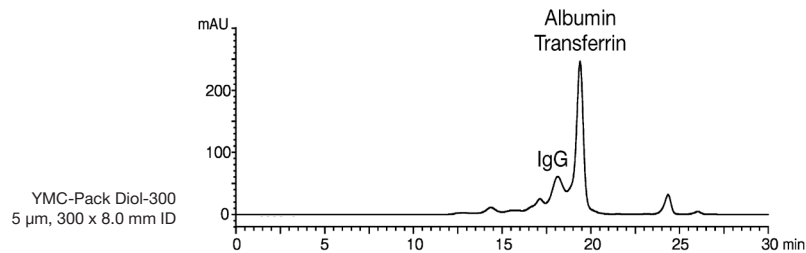
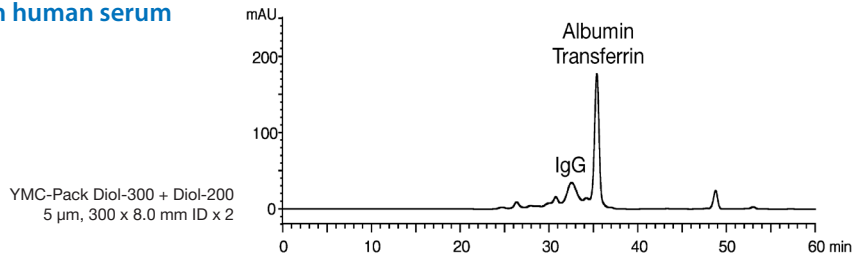
RP



Column: YMC-Triart Bio C4 (5 μ m, 30 nm) 150 x 3.0 mm ID
 Part No.: TB30S03-1503PTH
 Eluent: A) water/TFA (100/0.1)
 B) acetonitrile/TFA (100/0.1)
 Gradient: 20-60%B (0-27 min), 90%B (27-35 min)
 Flow rate: 0.4 mL/min
 Temperature: 70 °C
 Detection: UV at 220 nm
 Injection: 10 μ L (0.25 - 0.50 mg/mL)

Proteins in human serum

SEC



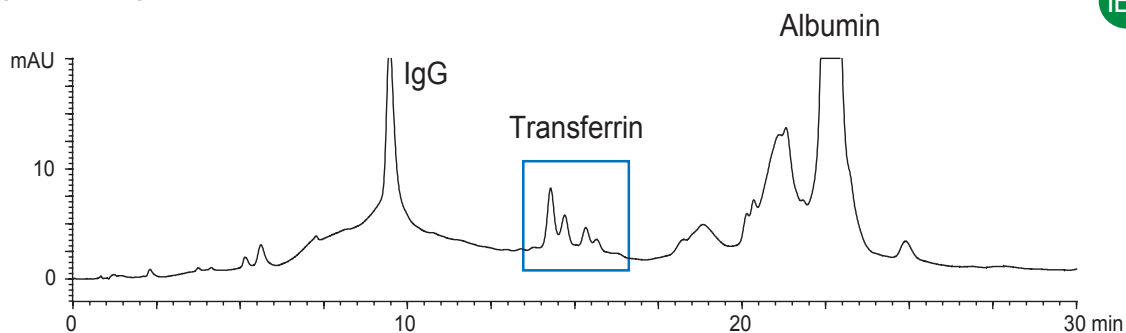
Eluent: 0.1 M KH_2PO_4 - K_2HPO_4 (pH 7.0) containing 0.2 M NaCl
 Part Nos.: DL30S05-3008WT + DL20S05-3008WT
 Flow rate: 0.5 mL/min
 Temperature: ambient (25 °C)

Detection: UV at 280 nm
 Injection: 20 μ L
 Sample: Human serum (100 μ L/mL)

BioLC applications – Proteins

Separation of proteins in human serum

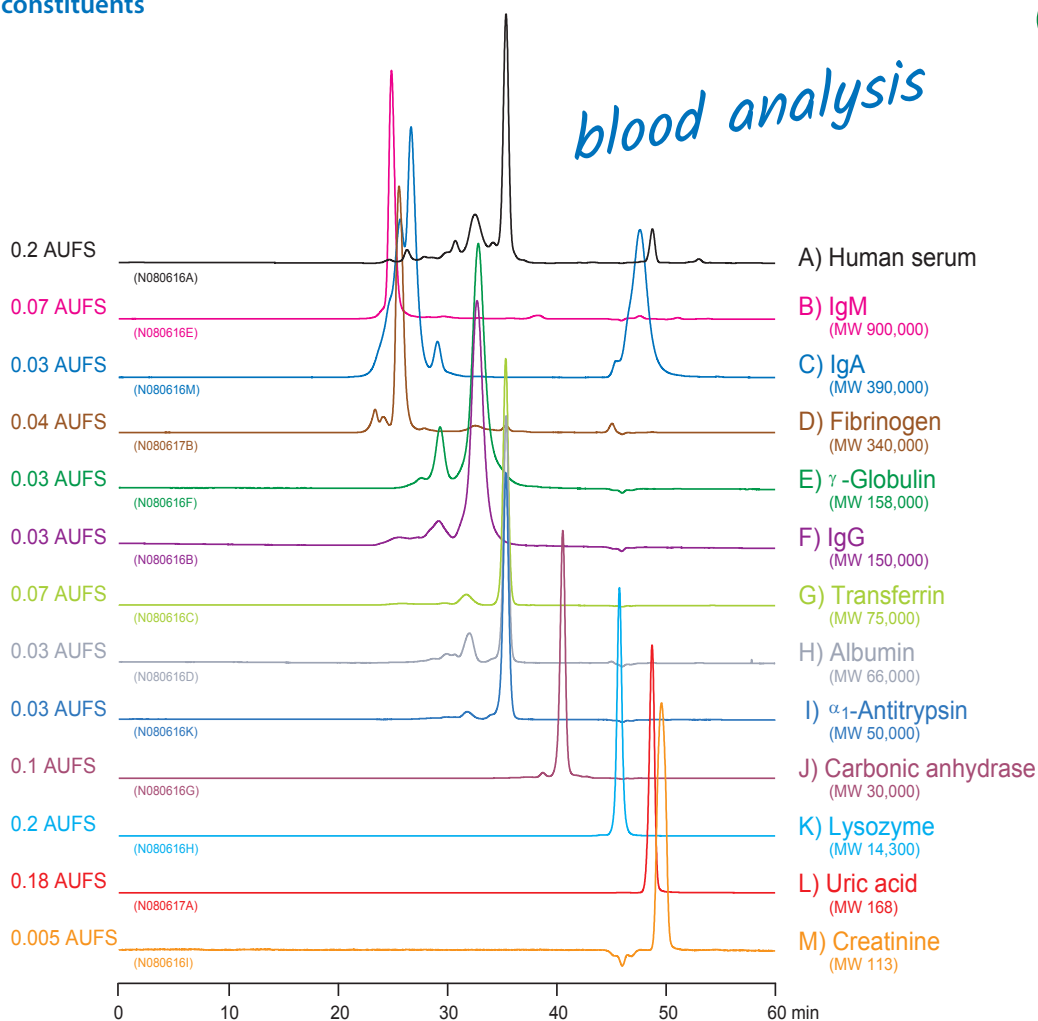
IEX



| | | | |
|-----------|--|--------------|-------------------------|
| Column: | BioPro IEX QA (5 µm) 50 x 4.6 mm ID | Flow rate: | 0.5 mL/min |
| Part No.: | QAA0S05-0546WP | Temperature: | 25 °C |
| Eluent: | A) 20 mM Tris-HCl (pH 8.6) | Detection: | UV at 280 nm |
| | B) 20 mM Tris-HCl (pH 8.6) containing 0.5 M NaCl | Injection: | 20 µL |
| Gradient: | 0–30%B (0–15 min), 30–100%B (15–30 min) | Sample: | Human serum (100 µL/mL) |

Plasma constituents

SEC

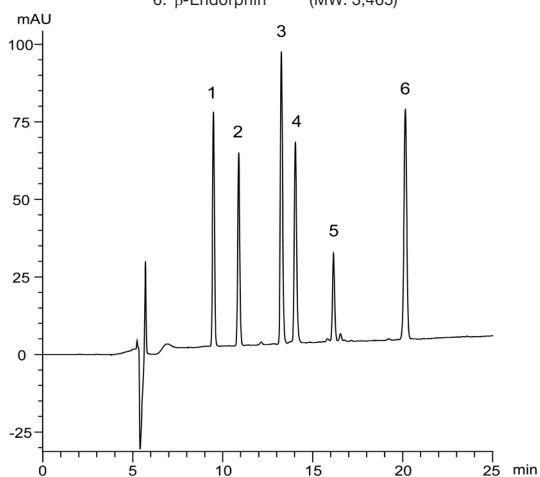


| | | | |
|------------|---|--------------|------------------------------|
| Columns: | YMC-Pack Diol-300 + Diol-200 (5 µm) 300 x 8.0 mm ID x 2 | Temperature: | ambient (25 °C) |
| Part Nos.: | DL30S05-3008WT + DL20S05-3008WT | Detection: | UV at 280 nm |
| Eluent: | 0.1 M KH ₂ PO ₄ -K ₂ HPO ₄ (pH 7.0) containing 0.2 M NaCl | Injection: | 20 µL (L: 1 µL) |
| Flow rate: | 0.5 mL/min | Sample: | A) 100 µL/mL; B-M) 1.0 mg/mL |

Peptides covering different MW

RP

1. Oxytocin (MW: 1,007)
2. Met-Enkephalin (MW: 574)
3. Leu-Enkephalin (MW: 556)
4. Neurotensin (MW: 1,673)
5. γ -Endorphin (MW: 1,859)
6. β -Endorphin (MW: 3,465)

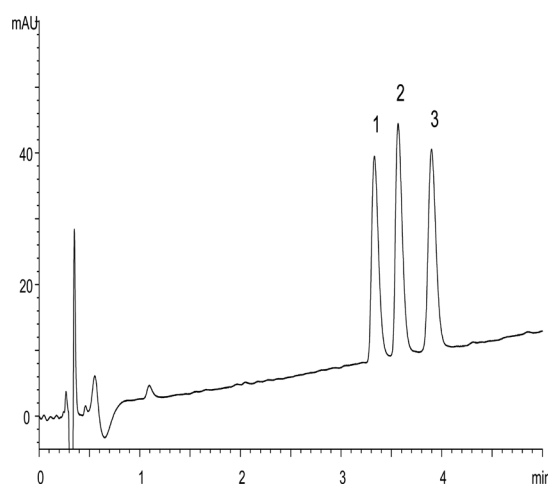


Column: YMC-Triart C18 (5 μ m, 12 nm) 150 x 2.0 mm ID
 Part No.: TA12S05-1502WT
 Eluent: A) water/TFA (100/0.1)
 B) acetonitrile / TFA (100/0.1)
 Gradient: 20%–45%B (0–25 min)
 Flow rate: 0.2 mL/min
 Temperature: 37 °C
 Detection: UV at 220 nm
 Injection: 2 μ L (0.075 \approx 0.25 mg/mL)

Antimicrobial peptides

RP

1. α -Defensin-1 (human) (MW: 3,442)
2. α -Defensin-2 (human) (MW: 3,371)
3. α -Defensin-3 (human) (MW: 3,486)

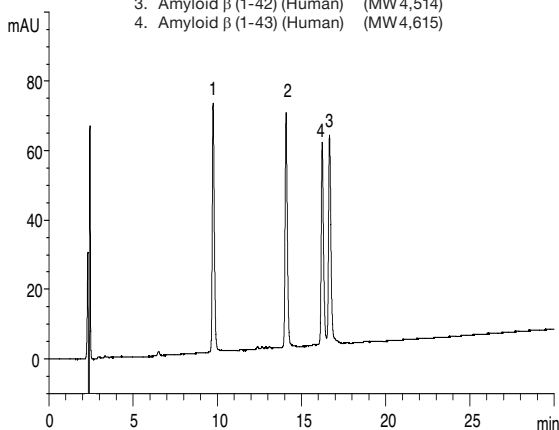


Column: YMC-Triart C18 (1.9 μ m, 12 nm) 50 x 2.0 mm ID
 Part No.: TA12SP9-0502PT
 Eluent: A) water/formic acid (100/0.1)
 B) 2-propanol/acetonitrile/formic acid (50/50/0.08)
 Gradient: 10%–25%B (0–10 min)
 Flow rate: 0.4 mL/min
 Temperature: 70 °C
 Detection: UV at 220 nm
 Injection: 1 μ L (50 μ g/mL)

Amyloid β -peptides

RP

1. Amyloid β (1-38) (Human) (MW 4,132)
2. Amyloid β (1-40) (Human) (MW 4,330)
3. Amyloid β (1-42) (Human) (MW 4,514)
4. Amyloid β (1-43) (Human) (MW 4,615)



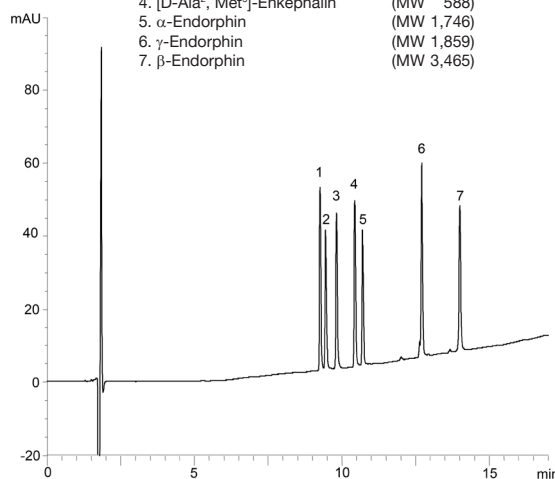
Amyloid β (1-43): Asp-Ala-Glu-Phe-Arg-His-Asp-Ser-Gly-Tyr-Glu-Val-His-His-Gln-Lys-Leu-Val-Phe-Phe-Ala-Glu-Asp-Val-Gly-Ser-Asn-Lys-Gly-Ala-Ile-Ile-Gly-Leu-Met-Val-Gly-Gly-Val-Val-Ile-Ala-Thr

Column: YMC-Triart Bio C4 (3 μ m, 30 nm) 150 x 3.0 mm ID
 Part No.: TB30S03-1503PTH
 Eluent: A) water/TFA (100/0.1)
 B) acetonitrile/TFA (100/0.1)
 Gradient: 25–40%B (0–30 min), 90%B (30–40 min)
 Flow rate: 0.4 mL/min
 Temperature: 70 °C
 Detection: UV at 220 nm
 Injection: 4 μ L (each 0.1 mg/mL)

Peptides

RP

1. BAM-12P (MW 1,425)
2. [D-Ala², Met⁵]-Enkephalinamide (MW 587)
3. Met-Enkephalin (MW 574)
4. [D-Ala², Met⁵]-Enkephalin (MW 588)
5. α -Endorphin (MW 1,746)
6. γ -Endorphin (MW 1,859)
7. β -Endorphin (MW 3,465)

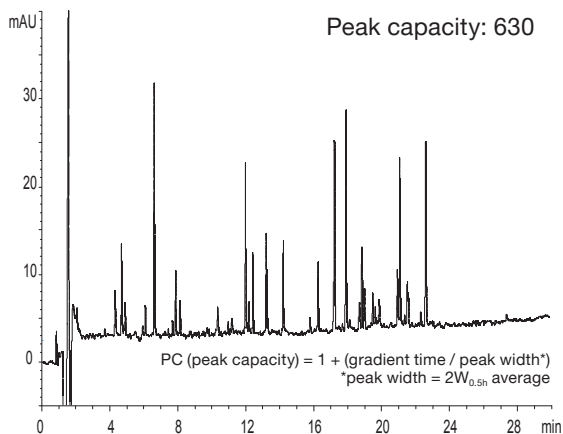


Column: Meteoric Core C18 BIO (2.7 μ m, 16 nm) 150 x 2.1 mm ID
 Part No.: CAW16SQ7-15Q1PT
 Eluent: A) water/TFA (100/0.1)
 B) acetonitrile/TFA (100/0.1)
 Gradient: 15–55%B (0–15 min), 55%B (15–17 min)
 Flow rate: 0.2 mL/min
 Temperature: 40 °C
 Detection: UV at 220 nm
 Injection: 2 μ L (0.02–0.5 mg/mL)
 Pressure: 14.9–16.1 MPa (2,160–2,330 psi)

BioLC applications – Peptide Mapping / Oligonucleotides

Tryptic digest of Hemoglobin

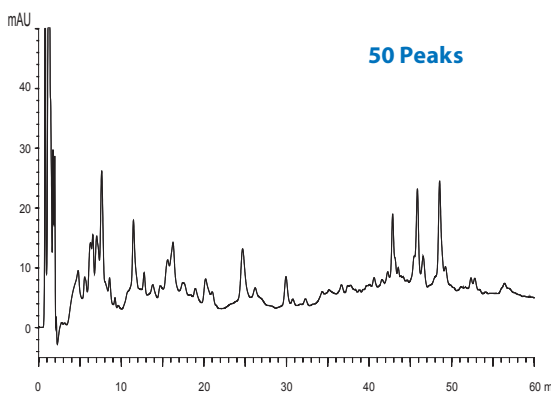
RP



Column: YMC-Triart C18 (1.9 μ m, 12 nm) 200 x 2.0 mm ID
(Two coupled 100 x 2.0 mm ID)
Part No.: TA12SP9-1002PT (2x)
Eluent: A) water/TFA (100/0.1)
B) acetonitrile/TFA (100/0.08)
Gradient: 5–40%B (0–30 min)
Flow rate: 0.4 mL/min
Temperature: 70 °C
Detection: UV at 220 nm
Injection: 20 μ L
Sample: Tryptic digest of Bovine Hemoglobin (2.5 nmol/mL)
Pressure: 58.1–61.6 MPa (8,430–8,930 psi)

Peptide mapping of tryptic digests of BSA with highest sensitivity

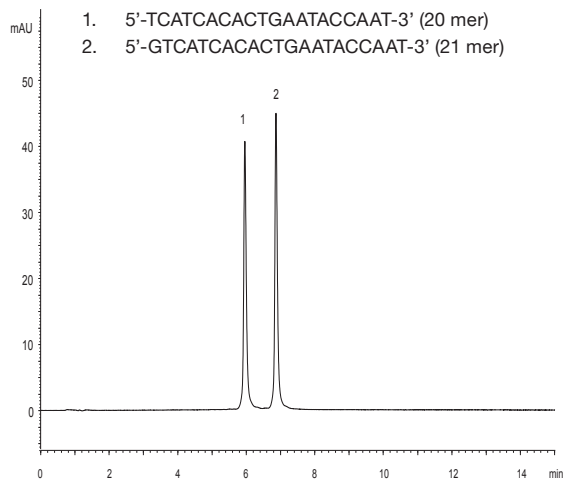
IEX



Column: BioPro IEX QA (5 μ m) 50 x 4.6 mm ID
Part No.: QAA0S05-0546WP
Eluent: A) 20 mM Tris-HCl (pH 8.6)
B) 20 mM Tris-HCl (pH 8.6) containing 0.5 M NaCl
Gradient: 0–15%B (0–30 min), 15–60%B (30–60 min)
Flow rate: 0.5 mL/min
Temperature: 25 °C
Detection: UV at 220 nm
Injection: 20 μ L
Sample: Tryptic digest of BSA

Separation of synthetic oligonucleotides (single-strand DNA)

IEX

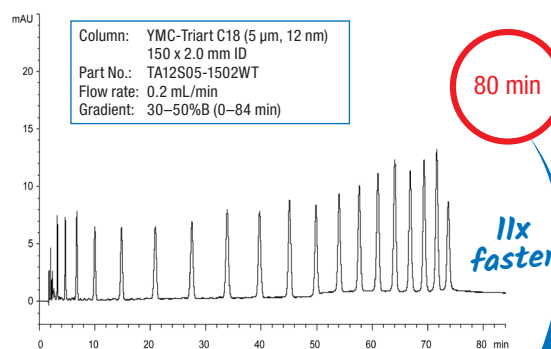


Column: BioPro IEX QF (5 μ m) 100 x 4.6 mm ID
Part No.: QF00S05-1046WP
Eluent: A) 10 mM NaOH
B) 10 mM NaOH containing 1.0 M NaClO₄
Gradient: 25–55%B (0–15 min), 100%B (15–20 min)
Flow rate: 1.0 mL/min
Temperature: 25 °C
Detection: UV at 260 nm
Injection: 4 μ L (5 nmol/L)

Oligonucleotides d(T)2-20 method transfer from HPLC to UHPLC

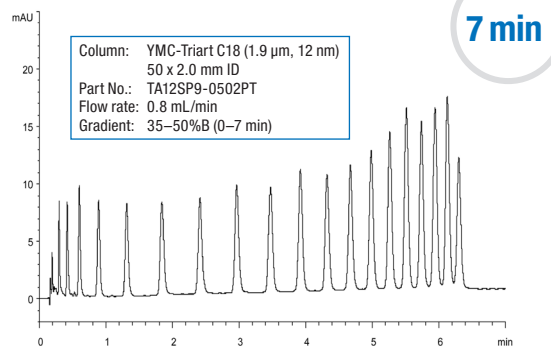
RP

Conventional LC method



Column: YMC-Triart C18 (5 μ m, 12 nm)
150 x 2.0 mm ID
Part No.: TA12S05-1502WT
Flow rate: 0.2 mL/min
Gradient: 30–50%B (0–84 min)

UHPLC method



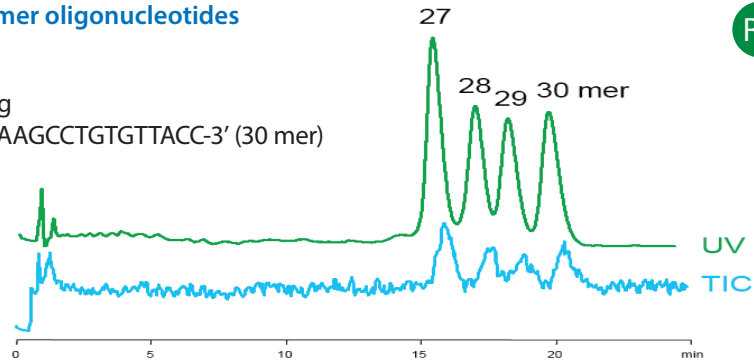
Column: YMC-Triart C18 (1.9 μ m, 12 nm)
50 x 2.0 mm ID
Part No.: TA12SP9-0502PT
Flow rate: 0.8 mL/min
Gradient: 35–50%B (0–7 min)

Eluent: A) 10 mM di-n-butylamine-acetic acid (pH 6.0)
B) methanol
Temperature: 37 °C
Detection: UV at 269 nm
Injection: 1 μ L (5 nmol/mL)

LC-MS analysis of synthetic 27-30 mer oligonucleotides

RP

Sample: Primer of DNA sequencing
5'-CCGCTCGAGCTAAAAAAGCCTGTGTTACC-3' (30 mer)

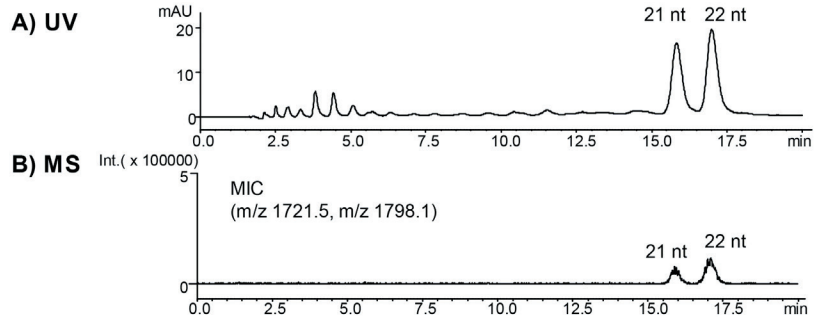


Column: Hydrosphere C18 (3 μ m) 50 x 2.0 mm ID
Part No.: HS12S03-0502WT
Eluent: A) 10 mM DBAA (pH 6.0)
B) Mobile phase A / acetonitrile (50/50)
Gradient: 58%–62%B (0–20 min), 62%B (20–25 min)

Flow rate: 0.2 mL/min
Temperature: 35 °C
Detection: UV at 269 nm and ESI negative-mode
Injection: 1 μ L (10 pmol/component)

LC/MS analysis of miRNA

RP



Courtesy of M. Yamada, SHIMADZU CORPORATION

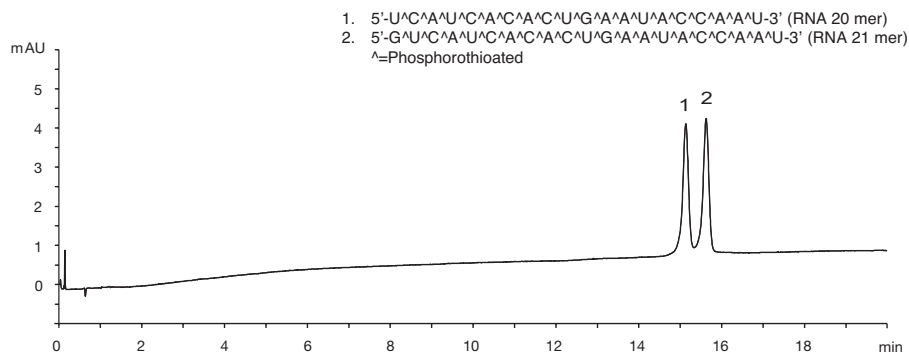
5'-pUGG AGU GUG ACA AUG GUG UUG-3' (21 nt, MW 6,890.1)
5'-pUGG AGU GUG ACA AUG GUG UUG U-3' (22 nt, MW 7,196.3)

Column: YMC-Triart C18 (3 μ m, 12 nm) 150 x 2.0 mm ID
Part No.: TA12S03-1502WT
Eluent: A) 10 mM di-n-butylamine-acetic acid (pH 7.5)
B) 10 mM di-n-butylamine-acetic acid (pH 7.5)/acetonitrile (50/50)
Gradient: 62–72%B (0–20 min)
Flow rate: 0.2 mL/min

Temperature: 30 °C
Detection: A) UV at 260 nm
B) ESI-negative mode
Injection: 4 μ L (5 nmol/mL)
System: LC) Shimadzu Prominence
MS) Shimadzu LCMS2020

Challenging phosphorothioate oligonucleotides

RP



- 5'-U[^]A[^]C[^]A[^]A[^]U[^]C[^]A[^]A[^]C[^]A[^]A[^]C[^]A[^]U[^]G[^]A[^]A[^]U[^]A[^]A[^]C[^]A[^]A[^]U[^]-3' (RNA 20 mer)
 - 5'-G[^]U[^]C[^]A[^]A[^]U[^]C[^]A[^]A[^]C[^]A[^]A[^]C[^]A[^]U[^]G[^]A[^]A[^]U[^]A[^]A[^]C[^]A[^]A[^]U[^]-3' (RNA 21 mer)
- [^]=Phosphorothioated

Column: YMC-Triart C8 **metal-free** (1.9 μ m, 12 nm) 100 x 2.1 mm ID
Part No.: TO12SP9-10Q1PTP
Eluent: A) 15mM triethylamine-400 mM HFIP*
B) methanol
Gradient: 10–20%B (0–20 min)

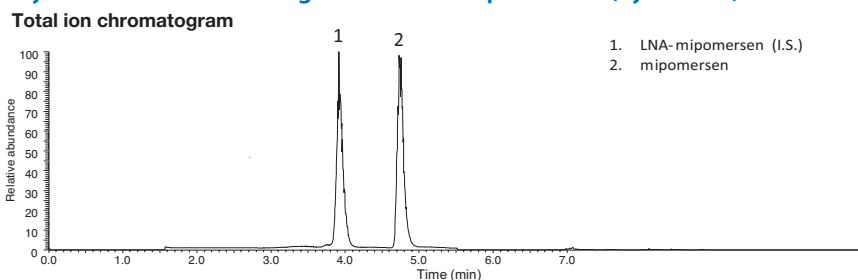
Flow rate: 0.42 mL/min
Temperature: 70 °C
Detection: UV at 260 nm
Injection: 1 μ L (each 1.25 nmol/mL)

*1,1,1,3,3,3-hexafluoro-2-propanol

BioLC applications – Oligonucleotides / Plasmids

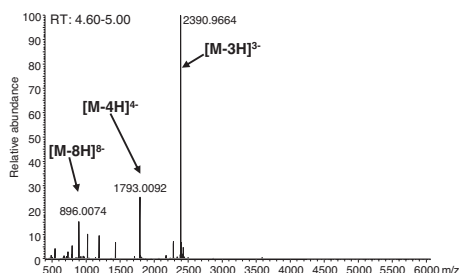
LC-HRMS analysis of the antisense oligonucleotide Mipomersen (Kynamro®)

RP

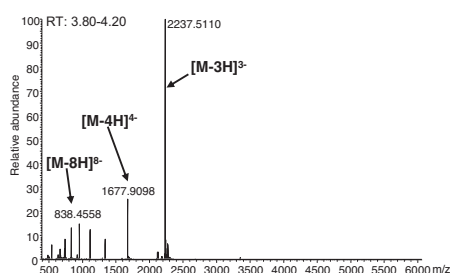


1. LNA- mipomersen (I.S.)
2. mipomersen

Mass spectrum of mipomersen



Mass spectrum of LNA-mipomersen



Column: YMC-Triart C8 **metal-free** (1.9 μm, 12nm)¹ 100 x 2.1 mm ID
 Part No.: TO12SP9-10Q1PTP
 Eluent: A) water/triethylamine/HFIP² (100/0.4/2; triethylamine 28.0mM, HFIP 135.8mM)
 B) methanol/triethylamine/HFIP (100/0.4/2)
 Gradient: [Sample separation step] 10–40%B (0–5.0 min)

[Column wash steps]
 40–70%B (5.0–5.1 min), 70%B (5.1–7.0 min), 70–10%B (7.0–7.1 min),
 10%B (7.1–8.0 min), 10–90%B (8.0–8.1 min), 90%B (8.1–9.0 min),
 90–10%B (9.0–9.1 min), 10%B (9.1–10.0 min),
 10–90%B (10.0–10.1 min), 90%B (10.1–11.0 min),
 90–10%B (11.0–11.1 min)
 Flow rate: 0.3mL/min
 Temperature: 50°C
 Injection: 10 μL (1000ng/mL)
 System : LC) Vanquish Binary Pump H system
 HRMS) Orbitrap HRMS Q Exactive Plus

*1 Prewash the column prior to the first use with water/methanol/phosphoric acid (70/30/0.1) for 1 hour

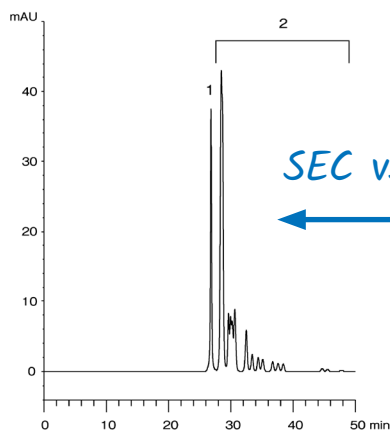
*2 1,1,1,3,3,3-hexafluoro-2-propanol

Reference: Y. Sun et al, Development of a bioanalytical method for an antisense therapeutic using high-resolution mass spectrometry, Bioanalysis, 2020 NOV 26, doi: 10.4155/bio-2020-0225.

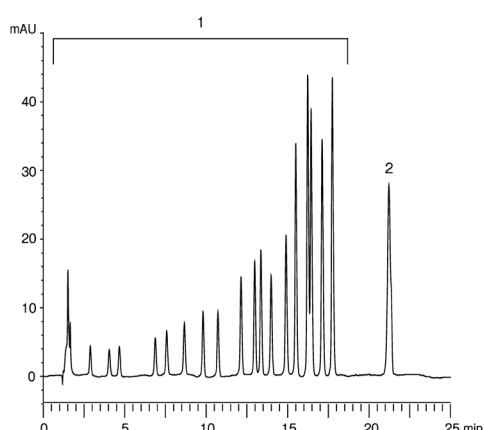
Plasmid pBR322 restriction and pBR322 Hae III restriction fragment

SEC IEX

1. Plasmid pBR322 (4,361 bp)
2. Plasmid pBR322 Hae III digest (8-587 bp)



1. Plasmid pBR322 Hae III digest (8-587 bp)
2. Plasmid pBR322 (4,361 bp)

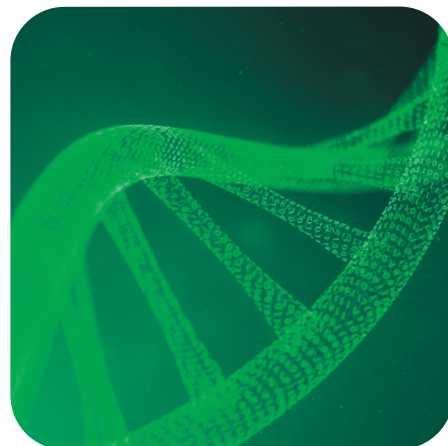
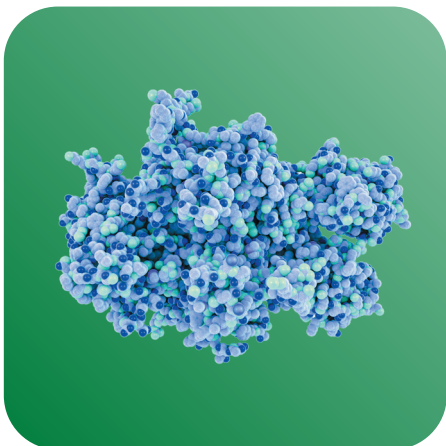


Columns: YMC-Pack Diol-300 + Diol-200 (5 μm) 500 x 8.0 mm ID
 Part Nos.: DL30S05-5008WT + DL20S05-5008WT
 Eluent: 0.1 M KH₂PO₄-K₂HPO₄ (pH 7.0) containing 0.2 M NaCl
 Flow rate: 0.7 mL/min
 Temperature: ambient (25 °C)
 Detection: UV at 260 nm
 Injection: 10 μL

Column: BioPro IEX QF (5 μm) 100 x 4.6 mm ID
 Part No.: QF00S05-1046WP
 Eluent: A) 20 mM Tris-HCl (pH 8.1)
 B) 20 mM Tris-HCl (pH 8.1) containing 0.1 M NaCl
 Gradient: 70–85%B (0–20 min), 85%B (20–25 min)
 Flow rate: 0.5 mL/min
 Temperature: 35 °C
 Detection: UV at 260 nm
 Injection: 10 μL



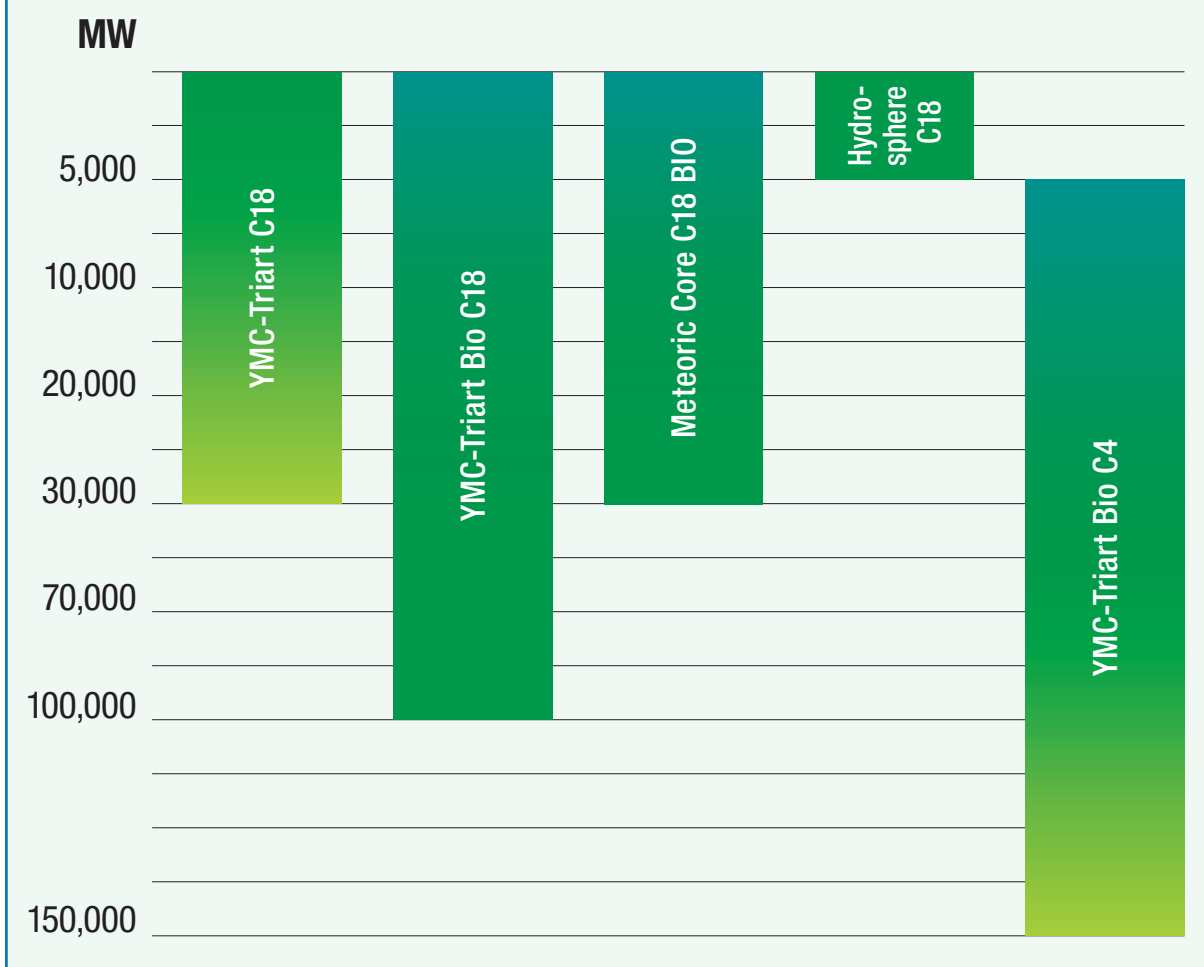
RP



RP – Bioseparation Columns

- Applicable to proteins, antibodies, peptides and oligonucleotides
- Selection of C18, C8 and C4 columns
- For UHPLC and HPLC
- pH- and temperature stable phases
- Superior reproducibility

Column Selection Tool according to molecular weight



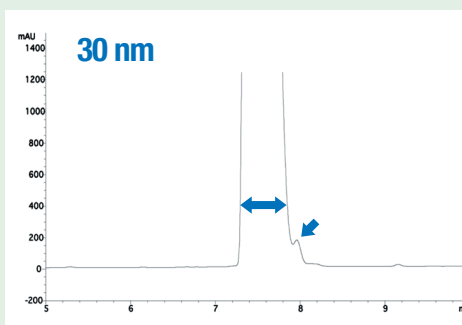
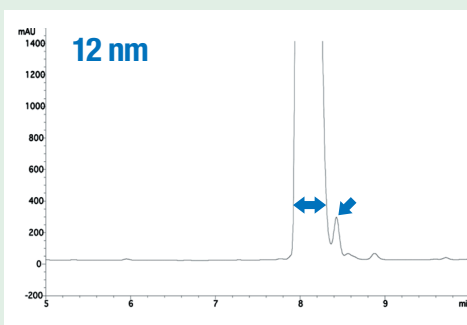
- most appropriate MW range
- extended MW range by elevated temperature
- appropriate MW range

Influence of pore size

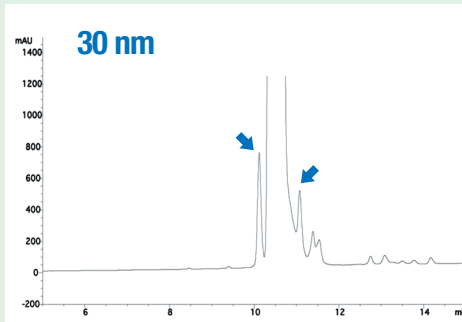
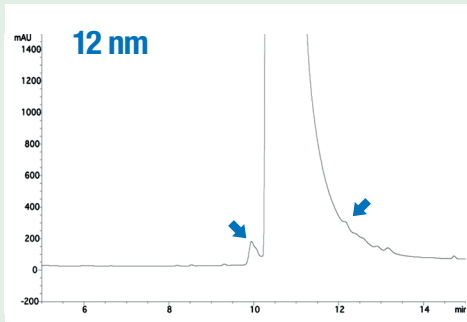
As shown in the table (on the left), the C18 column with 12 nm pore size is suitable for small peptides up to a MW of 5,000 Da. The highest efficiency for large peptides or small proteins can be obtained by using a wide pore C8 phase with 20 nm porosity. Most proteins can be eluted efficiently with a wide pore C4 column with 30 nm porosity.

However, the separation may also be influenced by the hydrophobicity of the peptide/protein and the nature of the column's bonded phase. Therefore, for initial method development, it can be useful, in the first instance, to follow the arrow shown in the *Column Selection Tool* for method optimisation.

Angiotensin II
(MW 1,046)



BSA
(MW 67,000)



For smaller peptides a small pore size is more successful. Larger molecules are separated much better with larger pore sizes!

RP – UHPLC / HPLC Selectivities

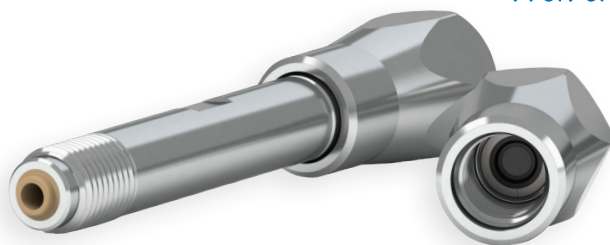
Selectivities for proteins/peptides and antibodies

| | YMC-Triart Bio C4 | YMC-Triart C18 | YMC-Triart Bio C18 | Meteoric Core Bio C18 |
|---|--------------------------------|--------------------------------|--------------------------------|--------------------------------|
| Modification | C4 (USP L26) | C18 (USP L1) | C18 (USP L1) | C18 (USP L1) |
| Particle Size / μm | 1.9, 3, 5 | 1.9, 3, 5 | 1.9, 3, 5 | 2.7 |
| Pore Size / nm | 30 | 12 | 30 | 8 (16) |
| pH range | 1.0 – 10.0 | 1.0 – 12.0 | 1.0 – 12.0 | 1.5 – 10.0 |
| Temperature range | pH < 7: 90 °C pH > 7: 50 °C | pH < 7: 90 °C pH > 7: 50 °C | pH < 7: 90 °C pH > 7: 50 °C | pH < 7: 70 °C pH > 7: 50 °C |

Selectivities for oligonucleotides

| | YMC-Triart C18 | YMC-Triart Bio C18 | Hydrosphere C18 | YMC-Triart C8 |
|---|--------------------------------|--------------------------------|-----------------|--------------------------------|
| Modification | C18 (USP L1) | C18 (USP L1) | C18 (USP L1) | C8 (USP L7) |
| Particle Size / μm | 1.9, 3, 5 | 1.9, 3, 5 | 2, 3, 5 | 1.9, 3, 5 |
| Pore Size / nm | 12 | 30 | 12 | 12 |
| pH range | 1.0 – 12.0 | 1.0 – 12.0 | 2.0 – 8.0 | 1.0 – 12.0 |
| Temperature range | pH < 7: 90 °C pH > 7: 50 °C | pH < 7: 90 °C pH > 7: 50 °C | 50 °C | pH < 7: 90 °C pH > 7: 50 °C |

*Biocompatible
hardware available!*



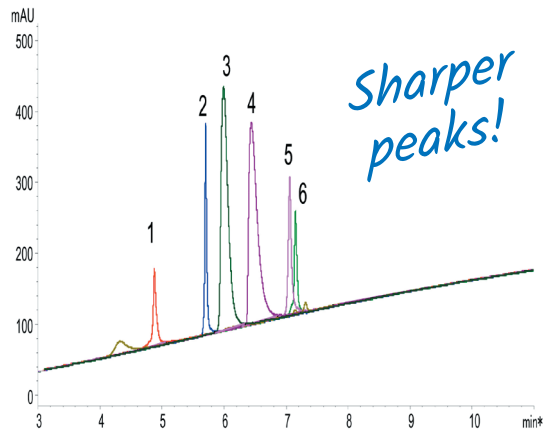
YMC-Triart **metal-free** columns are available for improved sensitivity and peak shape of coordinating compounds such as nucleotides or oligonucleotides, see page 38.

RP – YMC-Triart Bio C4: Sharper peaks

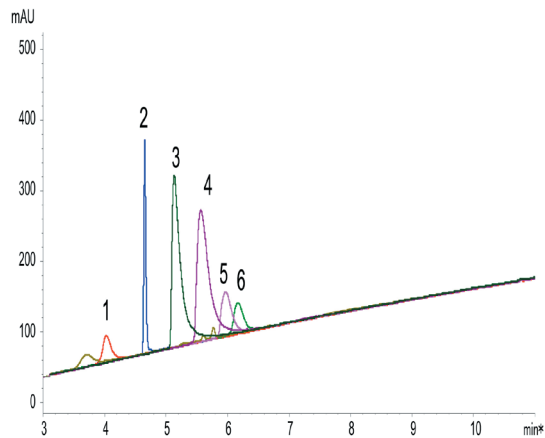
Better performance using YMC-Triart Bio C4

High sensitivity and sharp peaks under LC/MS compatible conditions

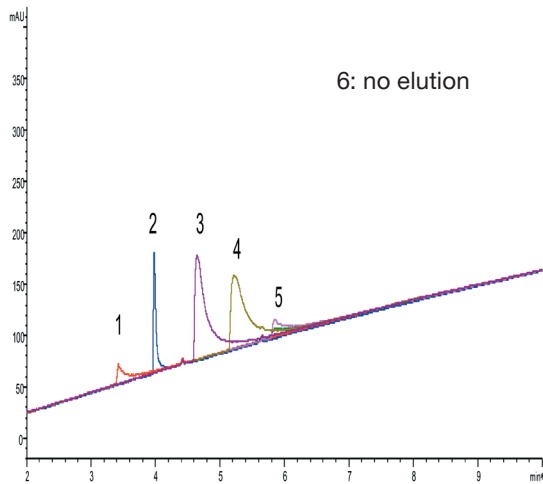
YMC-Triart Bio C4 (3 μ m, 30 nm)



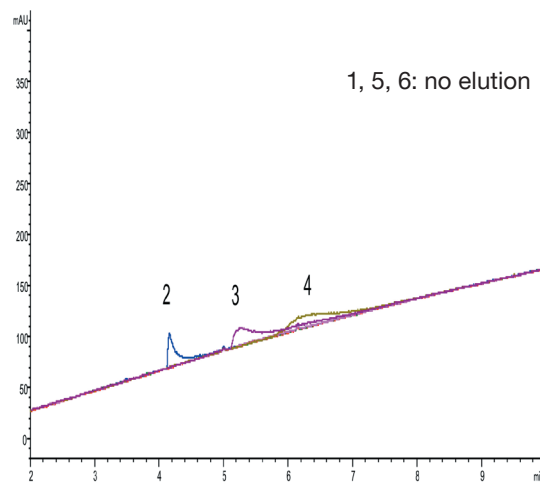
XBridge Protein BEH C4 (3.5 μ m, 30 nm)



AdvanceBio RP-mAb C4 (3.5 μ m, 45 nm)



Aeris widepore C4 (3.6 μ m, 20 nm)



Column: 150 x 3.0 mm ID
Part No.: TB30S03-1503PTH
Eluent: A) water/formic acid (100/0.1)
B) acetonitrile/formic acid (100/0.1)
Gradient: 10–95%B (0–15 min)
Flow rate: 0.4 mL/min (for 3.0 mm ID)
1.0 mL/min (for 4.6 mm ID)
Temperature: 40 °C

Detection: UV at 220 nm
Sample:
1. Cytochrome c (Horse heart)
2. Insulin (Bovine pancreas)
3. Transferrin (Human)
4. BSA
5. β -Lactoglobulin (Bovine)
6. α -Chymotrypsinogen A (Bovine pancreas)

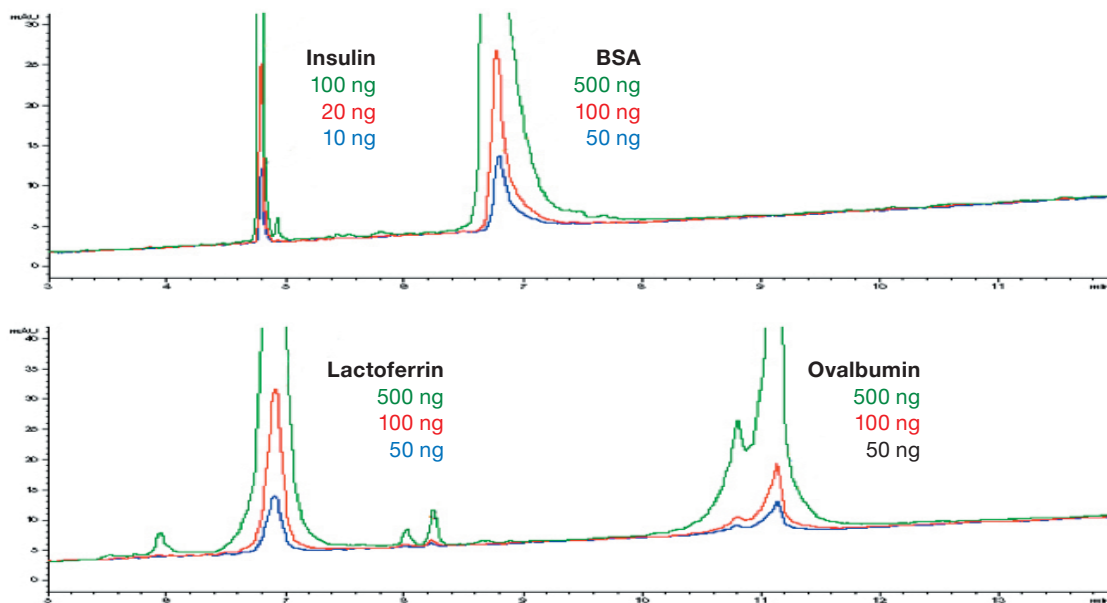
YMC-Triart Bio C4 shows better peak shape and recovery with a mobile phase containing formic acid, which is commonly used for LC/MS analysis. Therefore, YMC-Triart Bio C4 is ideal for high sensitivity analysis of proteins.

RP – YMC-Triart Bio C4: No column adsorption

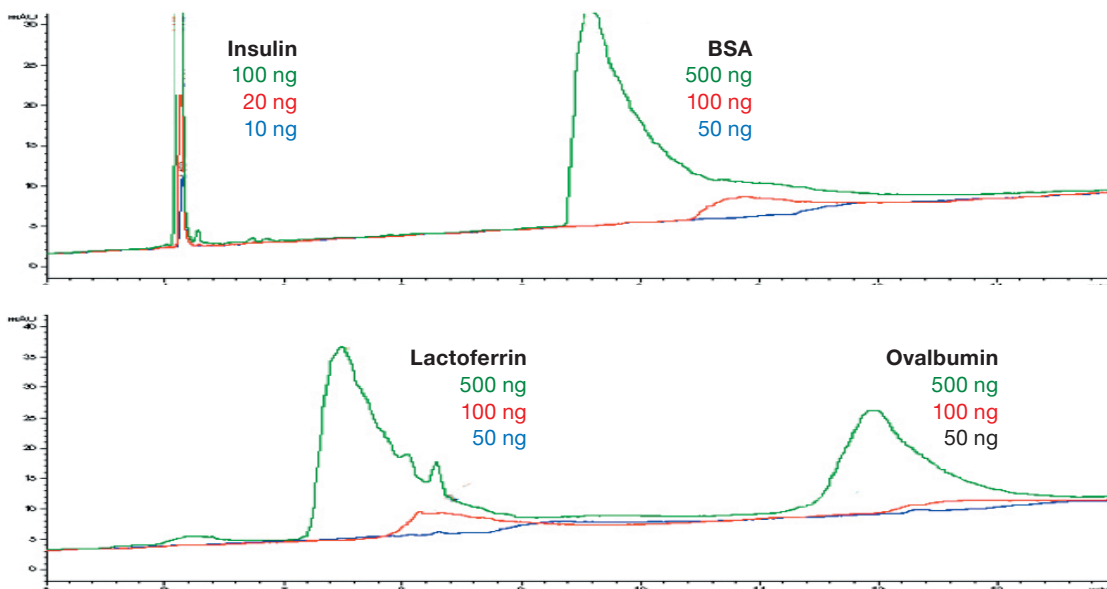
No sample adsorption by YMC-Triart Bio C4 columns

Ideal for Microanalysis

YMC-Triart Bio C4 (1.9 µm, 30 nm)



Aeris widepore C4 (3.6 µm, 20 nm)

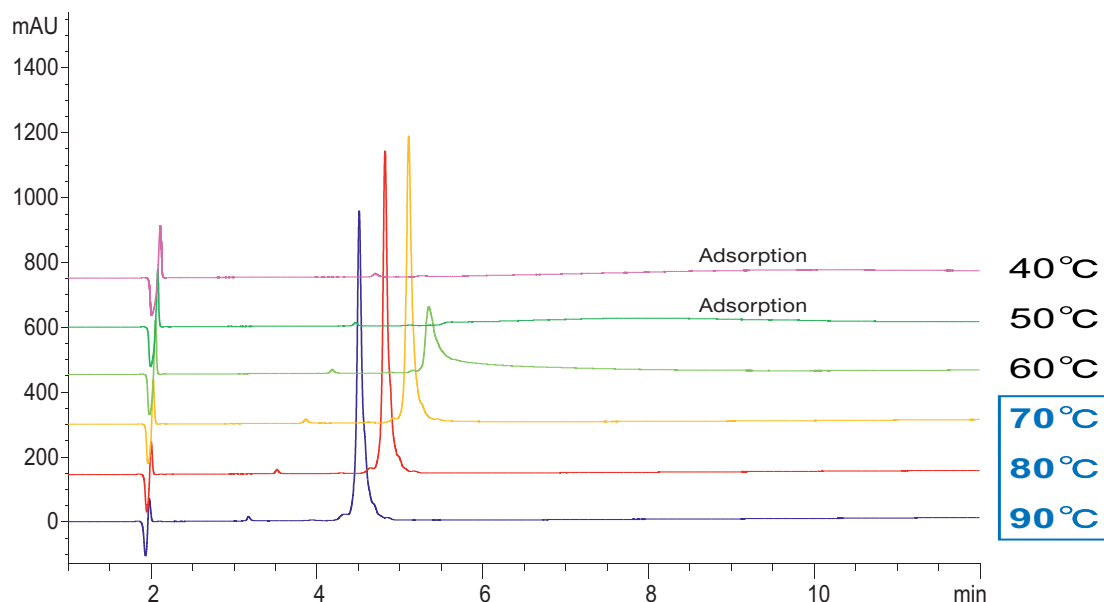


Column: 150 x 2.1 mm ID
 Part No.: TB30SP9-15Q1PT
 Eluent: A) water/TFA (100/0.05)
 B) acetonitrile/TFA (100/0.05)
 Gradient: 25-60%B (0-15 min), 90%B (15-20 min), 25%B (20-35 min)
 Flow rate: 0.2 mL/min
 Temperature: 40 °C
 Detection: UV at 220 nm

No sample adsorption was observed on YMC-Triart Bio C4 even at a low sample loading. This makes YMC-Triart Bio C4 ideal for microanalysis of proteins.

High temperature tolerance allows antibody analysis

Bevacizumab (Avastin®, MW: ca. 148 kDa)



Column: YMC-Triart Bio C4 (3 µm, 30 nm) 150 x 3.0 mm ID
Part No.: TB30S03-1503PTH
Eluent: A) water/TFA (100/0.1)
B) acetonitrile/TFA (100/0.1)
Gradient: 30–60%B (0–15 min), 90%B (15–30min)
Flow rate: 0.4 mL/min
Detection: UV at 220 nm
Injection: 4 µL
Sample: Bevacizumab (0.5 mg/mL)

“

“The possibility to use temperatures up to 90 °C with YMC-Triart Bio C4 simplifies the development of analytical methods. Furthermore, a good peak shape can be obtained without the addition of TFA, which means that I have fewer problems when using it for MS.”

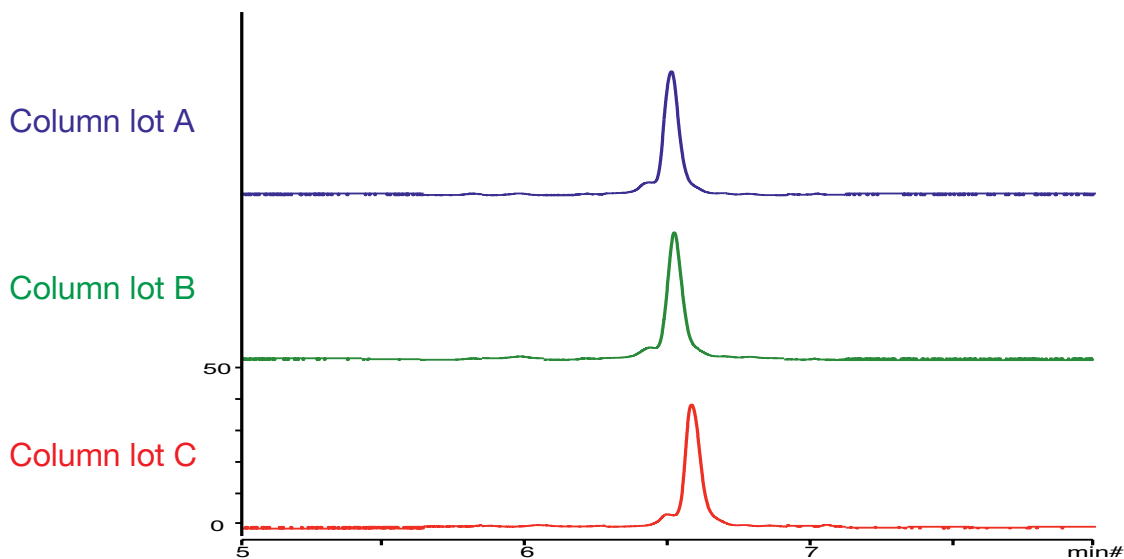
Lars M. H. Reinders, Institute for Energy and Environmental Technology e. V. (IUTA, DE)

”

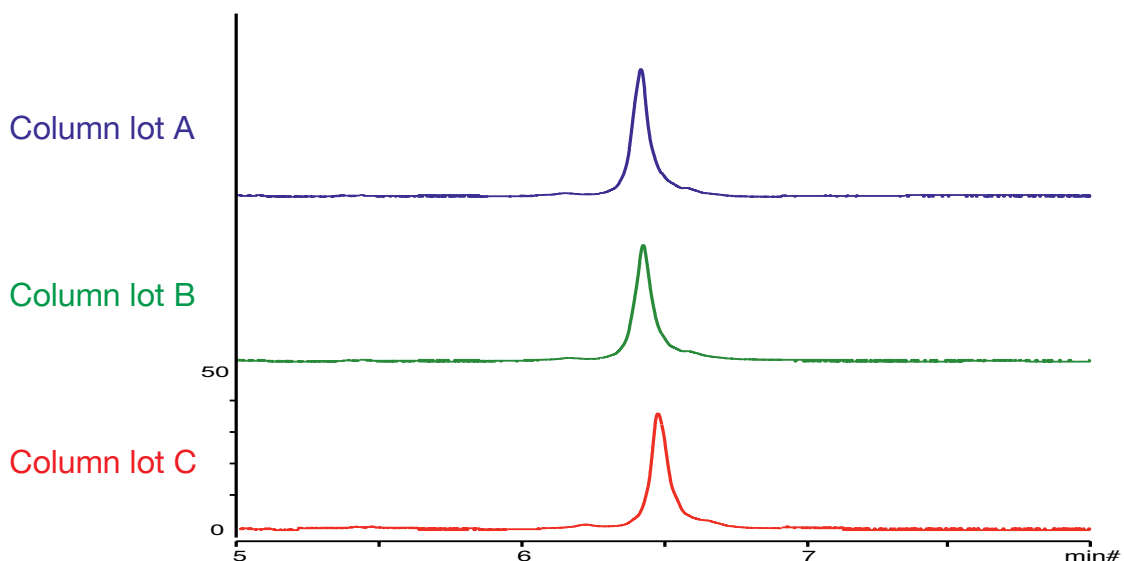
RP – YMC-Triart Bio C4: Reproducibility

Excellent Batch-to-batch reproducibility for antibody analysis

NIST mAb, 8671



Bevacizumab (Avastin®)



Column: YMC-Triart Bio C4 (1.9 μ m, 30 nm) 50 x 2.1 mm ID
 Part No.: TB30SP9-05Q1PT
 Eluent: A) water/TFA (100/0.1), B) acetonitrile/TFA (100/0.1)
 Gradient: 25–45%B (0–10 min)
 Flow rate: 0.4 mL/min
 Temperature: 80 °C
 Detection: UV at 280 nm
 Injection: 2 μ L (0.5 mg/mL)

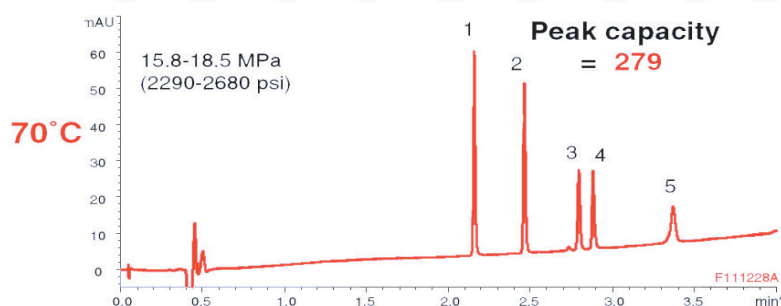
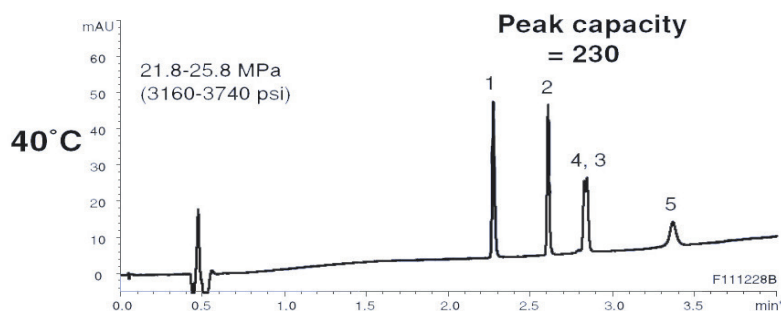
YMC-Triart Bio C4 shows excellent lot-to-lot reproducibility for antibodies. Not only is retention time highly reproducible, but also the resolution of minor impurity peaks. This makes YMC-Triart Bio C4 ideal for quality control of biopharmaceuticals.

RP – YMC-Triart C18: Temperature stability

More temperature flexibility using YMC-Triart

Highly efficient RP-HPLC separation of proteins

Mixture A (MW 500–18,400)



| Analytes | MW | Peak width ½h (min) | |
|----------|----|---------------------|-------|
| | | 40 °C | 70 °C |

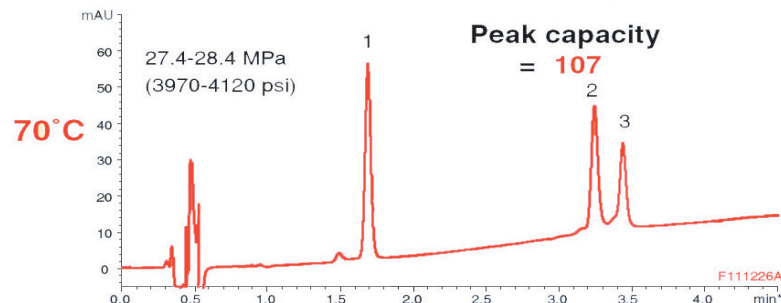
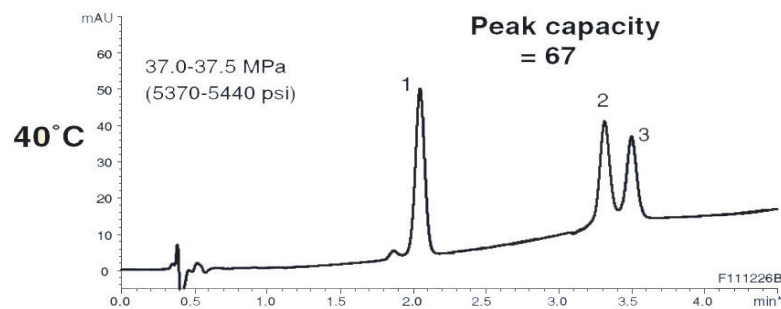
Mixture A

| | | | |
|----------------------|--------|-------|-------|
| 1. Oxytocin | 1,007 | 0.017 | 0.014 |
| 2. Leu-Enkephalin | 556 | 0.015 | 0.015 |
| 3. β-Endorphin | 3,465 | — | 0.016 |
| 4. Insulin | 5,733 | — | 0.015 |
| 5. β-Lactoglobulin A | 18,400 | 0.043 | 0.030 |

Mixture B

| | | | |
|-----------------------|--------|-------|-------|
| 1. Lysozyme | 14,300 | 0.069 | 0.044 |
| 2. α-Chymotrypsinogen | 25,700 | 0.080 | 0.049 |
| 3. β-Lactoglobulin A | 18,400 | 0.080 | 0.048 |

Mixture B (MW 14,300–25,700)



High temperatures only possible with YMC-Triart

Column: YMC-Triart C18 (1.9 μm, 12 nm) 50 x 2.0 mm ID
 Part-No.: TA12SP9-0502WT
 Eluent: A) water / TFA (100/0.1)
 B) acetonitrile / TFA (100/0.1) - mixture A
 B) acetonitrile / 2-propanol / TFA (50/50/0.1) - mixture B
 Gradient: 10–80%B (0–5 min) - mixture A
 30–60%B (0–5 min) - mixture B

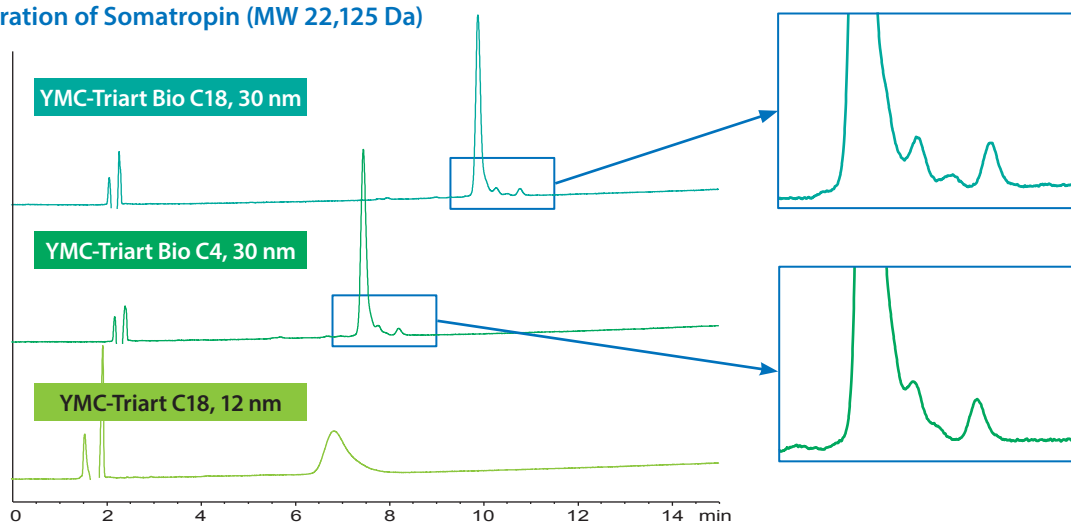
Flow rate: 0.4 mL/min
 Detection: UV at 220 nm
 Injection: 1 μL (50 μg/mL) - condition A
 1 μL (250 μg/mL) - condition B
 System: Agilent 1200SL

PC (peak capacity) = 1 + (gradient time / peak width*)
 *peak width = 2W_{0.5h} average

RP – YMC-Triart Bio C18: Great peak shapes

Ideal solutions for any kind of biomolecule

Separation of Somatropin (MW 22,125 Da)



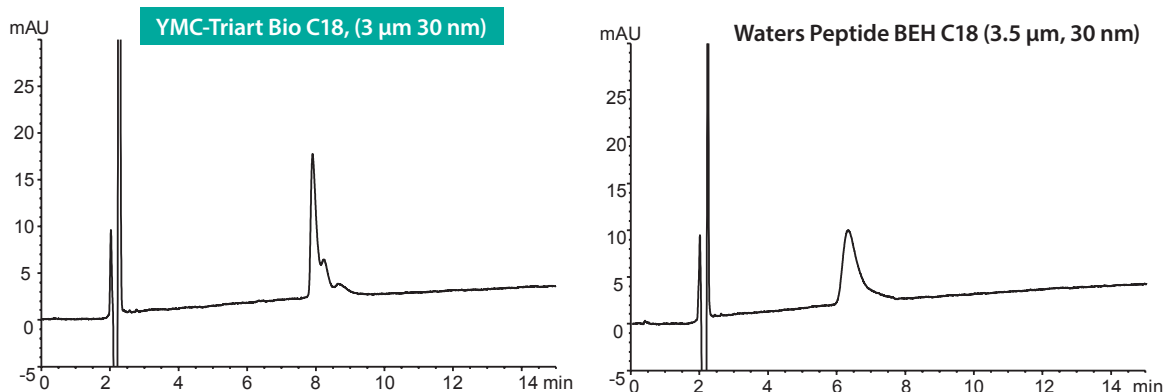
Columns: 150 x 3.0 mm ID (3 μ m)
 Part Nos.: TA30S03-1503PTH
 TB30S03-1503PTH
 TA12S03-1503PTH
 Eluent: A) water/TFA (100/0.1)
 B) acetonitrile/TFA (100/0.08)

Gradient: 50–70%B (0–15 min)
 Flow rate: 0.425 mL/min
 Temperature: 40 °C
 Detection: UV at 220 nm
 Injection: 4 μ L
 Sample: Somatropin (0.1 mg/mL)

In this example of somatropin, a peptide of 22,125 Da, good peak shape can be obtained with the widepore columns YMC-Triart Bio C18 and YMC-Triart Bio C4. Excellent separation was achieved using YMC-Triart Bio C18 with longer alkyl chains in its bonded phase.

Ideal for MS conditions

Good peak shape with mobile phase containing formic acid



Column: 150 x 3.0 mm ID; 150 x 4.6 mm ID
 Part No.: TA30S03-1503PTH
 Eluent: A) water/formic acid (100/0.1)
 B) acetonitrile/formic acid (100/0.08)
 Gradient: 45–65%B (0–15 min)

Flow rate: 0.425 mL/min for 3.0 mm ID; 1.0 mL/min for 4.6 mm ID
 Temperature: 40 °C
 Detection: UV at 220 nm
 Sample: Somatropin (0.1 mg/mL)

YMC-Triart Bio C18 is suitable for highly sensitive analysis and structural analysis of proteins using LC/MS since good peak shapes in mobile phase containing formic acid can be achieved.

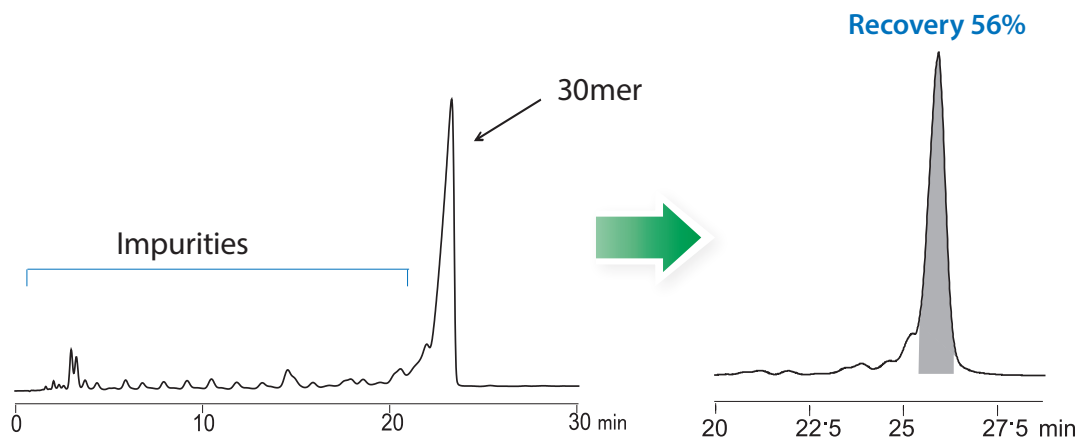
RP – Hydrosphere C18: Oligonucleotide purification

Easy purification of Oligonucleotides with YMC-Actus semi prep columns

Purification of synthetic 30mer oligonucleotide

Analysis 1.0 mL/min, 5 μ L injection
Hydrosphere C18
 50 x 4.6 mm ID, 5 μ m

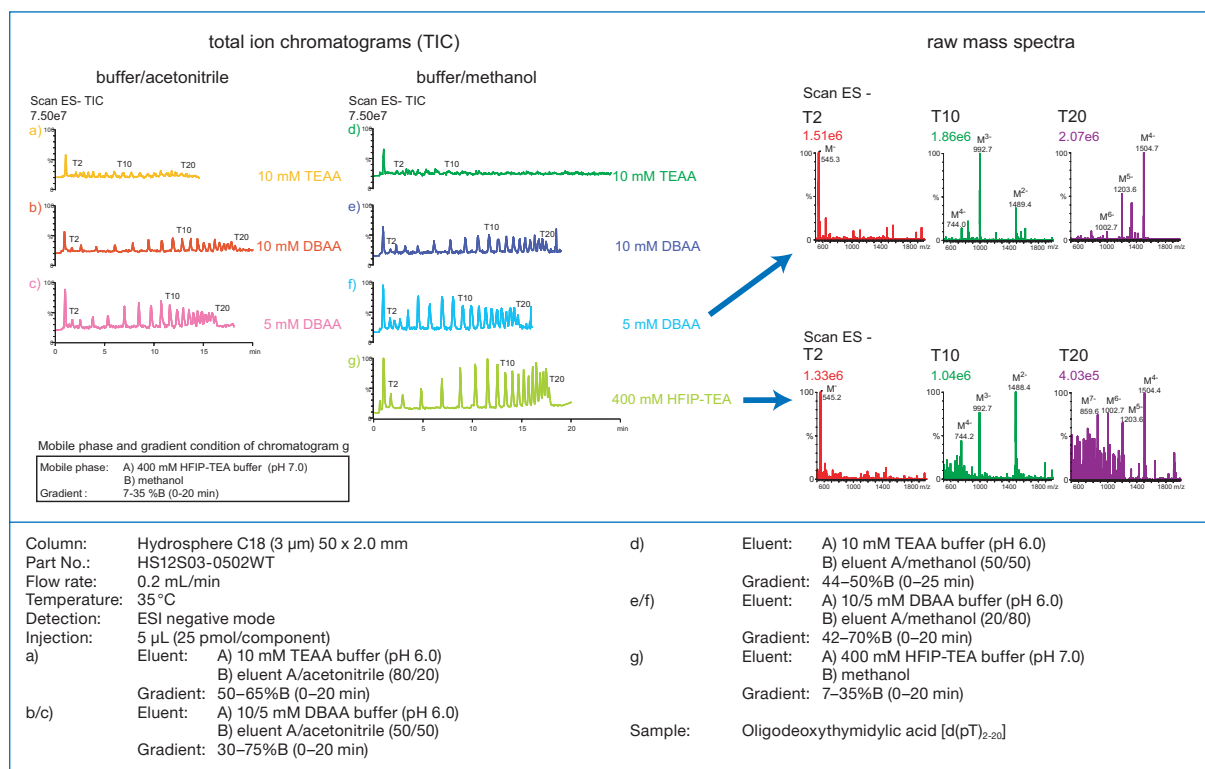
Purification 19 mL/min, 100 μ L injection
YMC-Actus Hydrosphere C18
 50 x 20 mm ID, 5 μ m



Part Nos.: HS12S05-0546WT
 HS12S05-0520WX
 Eluent: A) 10 mM DBA-acetic acid (pH 6.0) / methanol (60/40)
 B) 10 mM DBA-acetic acid (pH 6.0) / methanol (20/80)
 Gradient: 10%–35%B (0–30 min.)
 Temperature: ambient
 Detection: UV at 269 nm
 Sample: synthetic oligonucleotide (100 μ M)

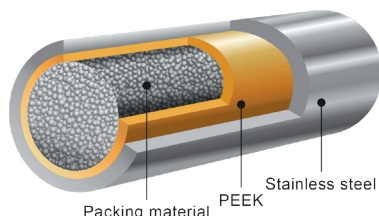
purity > 99%

Influences of mobile phase conditions on intensity of ESI-MS



RP – YMC-Triart: Biocompatible hardware

Metal-free column hardware ideal for oligonucleotide analysis

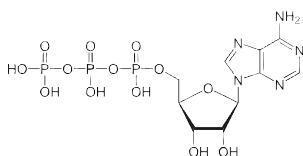


| Specifications | |
|-------------------|--|
| YMC-Triart Phases | C18, Bio C18, C8, Bio C4 |
| Particle Size | 1.9, 3, 5 μm |
| Inner layer | PEEK |
| Outer layer | Stainless steel |
| Frit | PEEK |
| Pressure limit | 1.9 μm : 100 MPa (15,000 psi) 3/5 μm : 45 MPa (6,525 psi) |

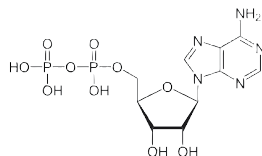
Special column connectors required.
See ordering information recommendations.

Improved sensitivity for coordination compounds

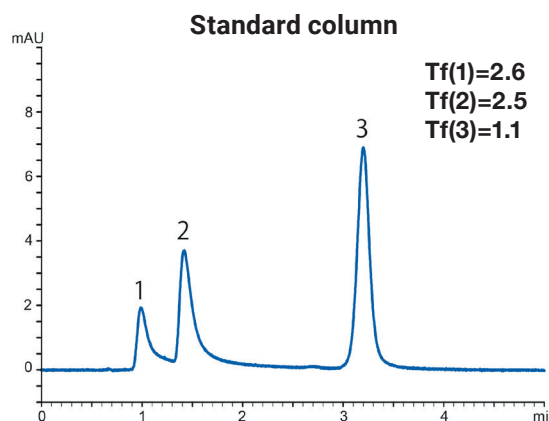
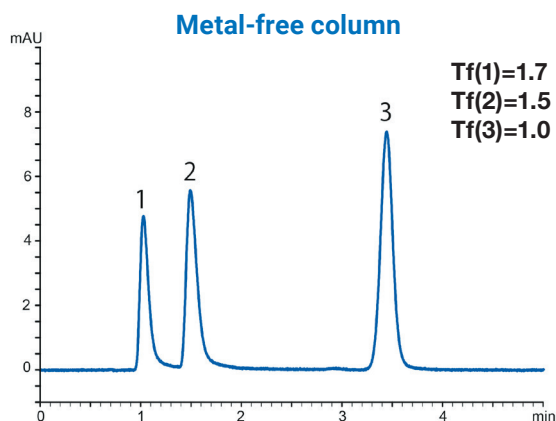
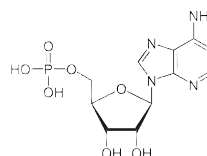
1. ATP



2. ADP



3. AMP

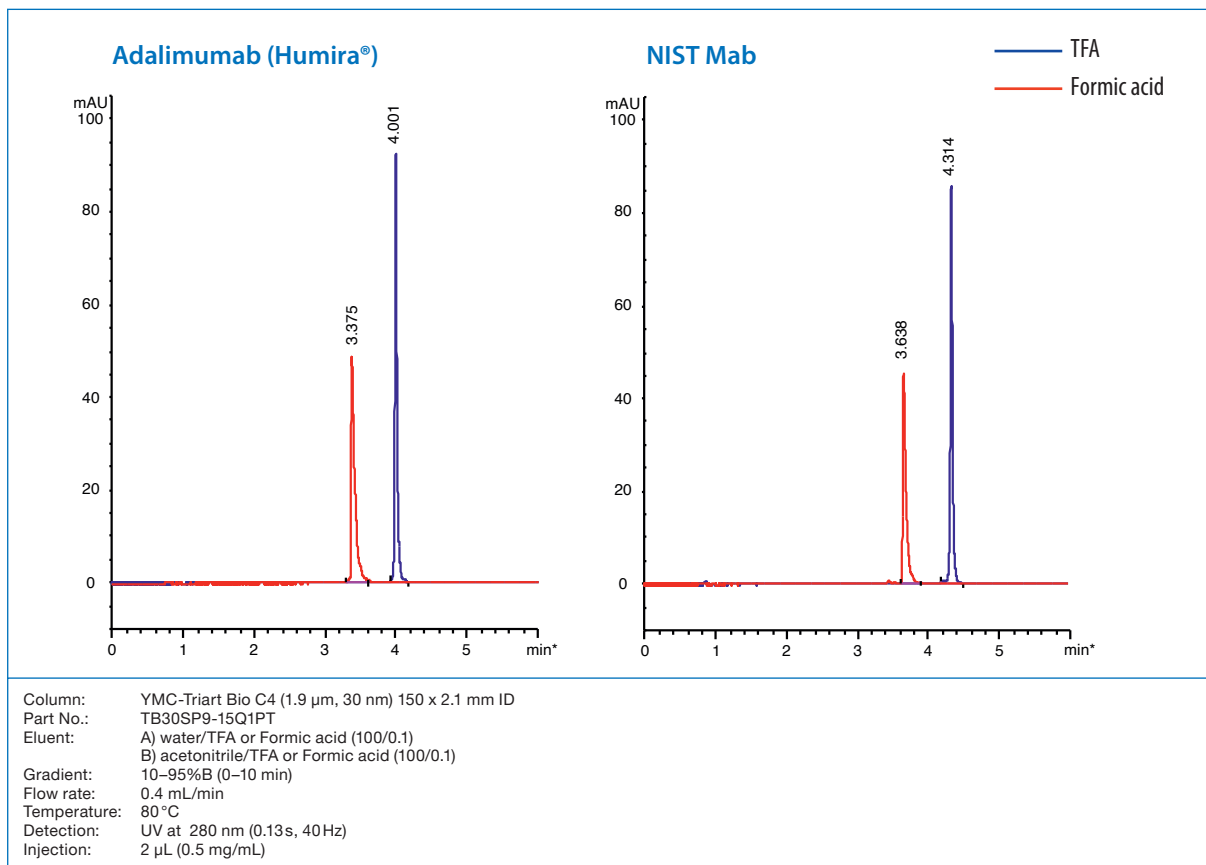


Column: YMC-Triart C18 (3 μm) 50 x 2.1 mm ID
 Part Nos.: TA12S03-05Q1PTP (**metal-free**) or
 TA12S03-05Q1PTH (regular hardware)
 Eluent: 5 mM HCOONH_4
 Flow rate: 0.21 mL/min
 Temperature: 25 $^\circ\text{C}$
 Detection: UV at 265 nm
 Injection: 1 μL (10 mg/mL)
 System: bioinert/"non-metal" HPLC system

Metal coordinating compounds, which have a phosphate group in their structure, tend to show poor peak shape due to interactions with metals, such as the stainless steel in column bodies and frits. By using the **metal-free** column hardware, better peak shapes can be expected. Nucleotides with phosphate groups show better peak shapes when compared to the regular column hardware. The **metal-free** column hardware is very suitable for highly sensitive analyses using LC/MS.

Use of MS compatible conditions for antibody analysis by RP

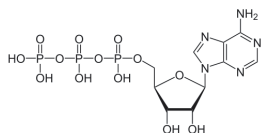
Although the best peak shapes and greater sensitivity can be provided by mobile phases containing TFA, suitable peaks can also be obtained with mobile phases containing formic acid – especially in combination with YMC-Triart Bio C4.



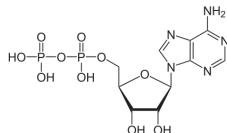
RP – Expert Tips: (Oligo)nucleotides

Influence of system and column hardware on the analysis of nucleotides

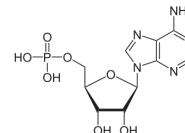
1 ATP



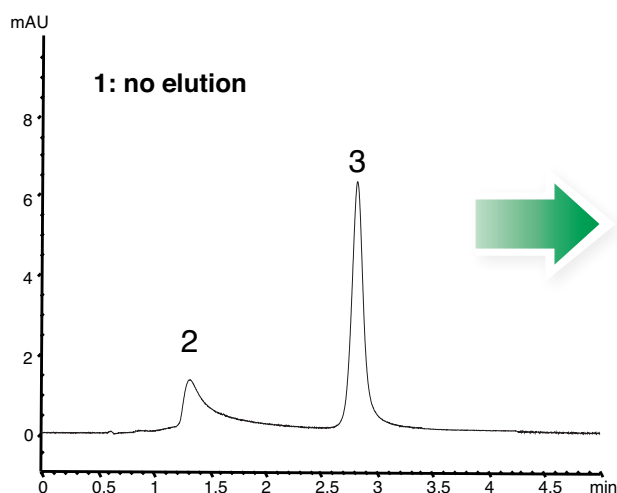
2 ADP



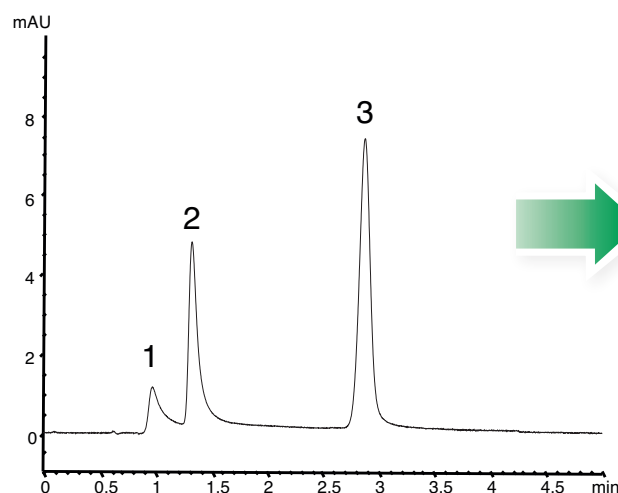
3 AMP



Ordinary HPLC system with standard column

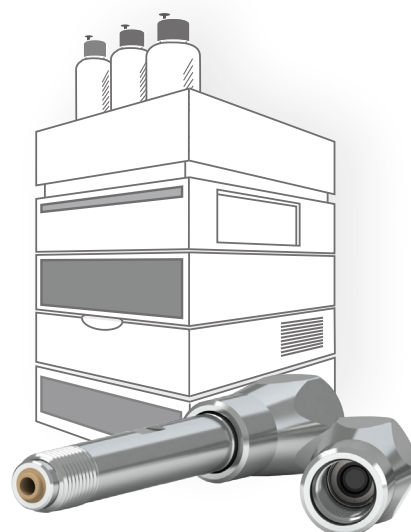
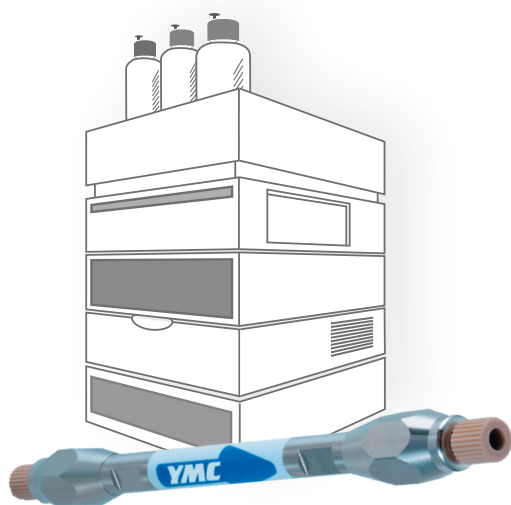


Ordinary HPLC system with metal-free column



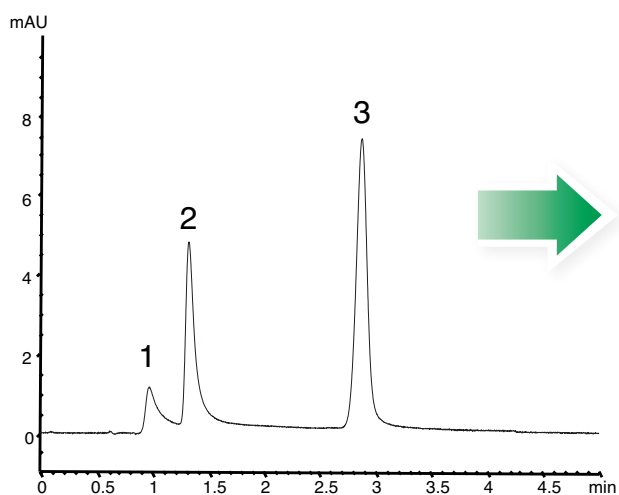
Column: YMC-Triart C18 (3 μ m, 12 nm) 50 x 2.1 mm ID
 Part Nos: TA12S03-05Q1PT (standard hardware)
 TA12S03-05Q1PTP (metal-free hardware)
 Eluent: 5 mM HCOONH₄
 Flow rate: 0.21 mL/min
 Temperature: 25 °C
 Detection: UV at 265 nm
 Injection: 1 μ L (10 μ g/mL)

"Non-metal" HPLC system: PEEK sample loop, PEEK injector port, and PEEK tubing are used.

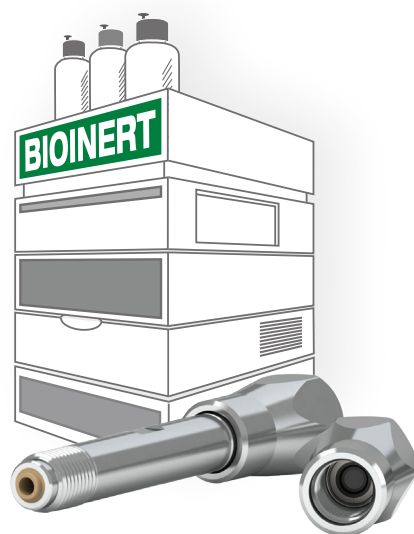
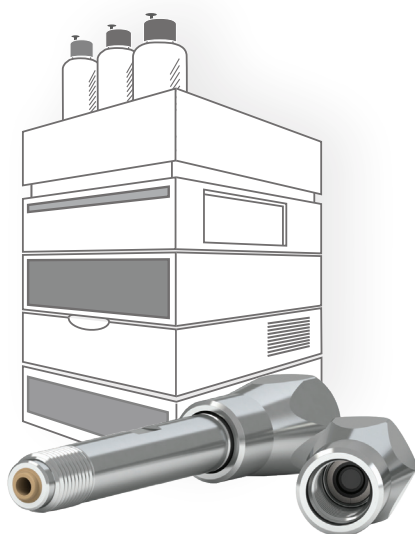
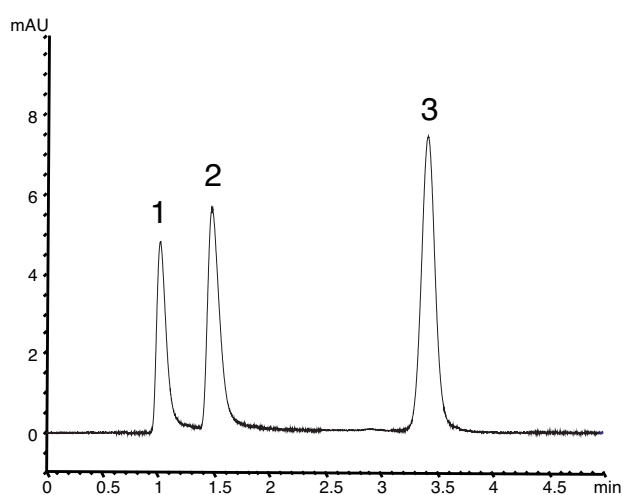


ATP peak is detected, and peak shape of ADP is improved as a result of using the metal-free column.

Ordinary HPLC system
with metal-free column



“Non-metal” HPLC system*
with metal-free column

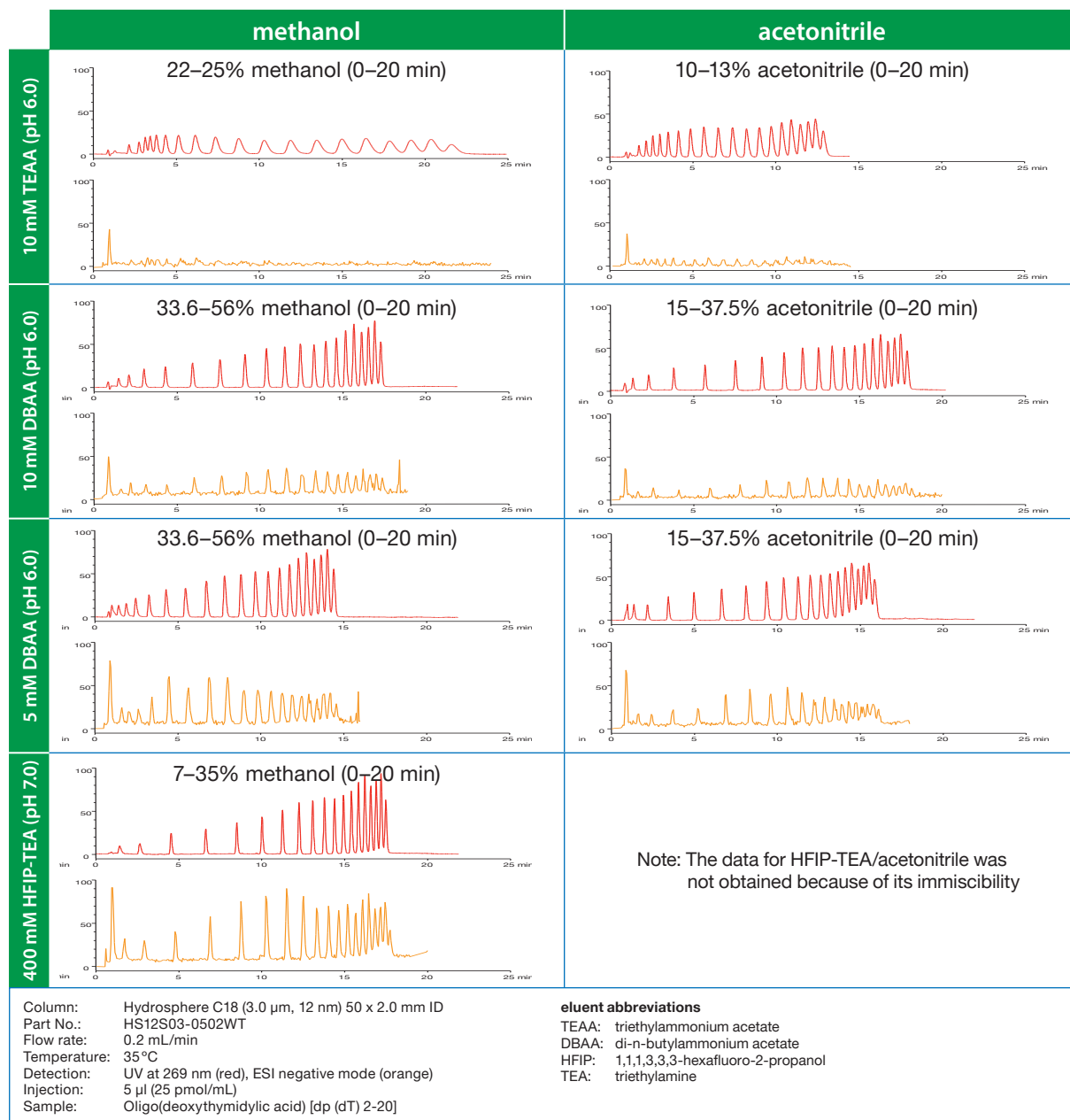


Peak shape is greatly improved as a result of using “non-metal” HPLC system

RP – Expert Tips: Oligonucleotides

Effect of composition and salt concentration of ion-pairing mobile phase on the separation and signal intensity

Comparison of separation and ESI-MS signal intensity using different ion-pairing buffers and organic solvents



The mobile phase composition has different effects on the separation and signal intensity in electrospray ionisation mass spectrometry (ESI-MS) of oligonucleotides. Using different gradient conditions, acceptable retention and resolution can be achieved (upper UV chromatograms; red trace) for each separation by optimising the gradient slope of the organic solvent regardless of the type of mobile phase. The ESI-MS intensity is significantly influenced by

the type and concentration of ion-pairing buffer as shown in the lower MS chromatograms (orange trace). HFIP-TEA buffer/methanol systems provide the maximum MS intensity. Enhanced retention and MS intensity are obtained using 10 mM DBAA buffer compared to 10 mM TEAA buffer, and the lower DBAA concentration results in approximately 1.5–3 times increase in the intensity without any change in the concentration of organic solvent.

1.9 µm UHPLC columns

| Phase | Column ID [mm] | Column length [mm] | | | | | Guard cartridges* with 5 mm length (pack of 3) |
|--------------------|----------------|--------------------|----------------|----------------|----------------|----------------|---|
| | | 30 | 50 | 75 | 100 | 150 | |
| YMC-Triart C18 | 2.0 | TA12SP9-0302PT | TA12SP9-0502PT | TA12SP9-L502PT | TA12SP9-1002PT | TA12SP9-1502PT | TA12SP9-E5Q1CC** |
| | 2.1 | TA12SP9-03Q1PT | TA12SP9-05Q1PT | TA12SP9-L5Q1PT | TA12SP9-10Q1PT | TA12SP9-15Q1PT | TA12SP9-E5Q1CC** |
| | 3.0 | — | TA12SP9-0503PT | TA12SP9-L503PT | TA12SP9-1003PT | TA12SP9-1503PT | TA12SP9-E503CC |
| YMC-Triart Bio C18 | 2.0 | TA30SP9-0302PT | TA30SP9-0502PT | TA30SP9-L502PT | TA30SP9-1002PT | TA30SP9-1502PT | TA30SP9-E5Q1CC** |
| | 2.1 | TA30SP9-03Q1PT | TA30SP9-05Q1PT | TA30SP9-L5Q1PT | TA30SP9-10Q1PT | TA30SP9-15Q1PT | TA30SP9-E5Q1CC** |
| | 3.0 | — | TA30SP9-0503PT | TA30SP9-L503PT | TA30SP9-1003PT | TA30SP9-1503PT | TA30SP9-E503CC |
| YMC-Triart C8 | 2.0 | T012SP9-0302PT | T012SP9-0502PT | T012SP9-L502PT | T012SP9-1002PT | T012SP9-1502PT | T012SP9-E5Q1CC** |
| | 2.1 | T012SP9-03Q1PT | T012SP9-05Q1PT | T012SP9-L5Q1PT | T012SP9-10Q1PT | T012SP9-15Q1PT | T012SP9-E5Q1CC** |
| | 3.0 | — | T012SP9-0503PT | T012SP9-L503PT | T012SP9-1003PT | T012SP9-1503PT | T012SP9-E503CC |
| YMC-Triart Bio C4 | 2.0 | TB30SP9-0302PT | TB30SP9-0502PT | TB30SP9-L502PT | TB30SP9-1002PT | TB30SP9-1502PT | TB30SP9-E5Q1CC** |
| | 2.1 | TB30SP9-03Q1PT | TB30SP9-05Q1PT | TB30SP9-L5Q1PT | TB30SP9-10Q1PT | TB30SP9-15Q1PT | TB30SP9-E5Q1CC** |
| | 3.0 | — | TB30SP9-0503PT | TB30SP9-L503PT | TB30SP9-1003PT | TB30SP9-1503PT | TB30SP9-E503CC |

*Guard cartridge holder required, part no. XPCHUHP





**Guard cartridge: 2.1 mm ID

1.9 µm metal-free UHPLC columns

| Phase | Column ID [mm] | Column length [mm] | | |
|--------------------|----------------|--------------------|-----------------|-----------------|
| | | 50 | 100 | 150 |
| YMC-Triart C18 | 2.1 | TA12SP9-05Q1PTP | TA12SP9-10Q1PTP | TA12SP9-15Q1PTP |
| YMC-Triart Bio C18 | 2.1 | TA30SP9-05Q1PTP | TA30SP9-10Q1PTP | TA30SP9-15Q1PTP |
| YMC-Triart C8 | 2.1 | T012SP9-05Q1PTP | T012SP9-10Q1PTP | T012SP9-15Q1PTP |
| YMC-Triart Bio C4 | 2.1 | TB30SP9-05Q1PTP | TB30SP9-10Q1PTP | TB30SP9-15Q1PTP |

Special column connectors required.

Column connectors for metal-free (U)HPLC columns

| Recommendation | ✓ ✓ | | ✓ | |
|-----------------|---|---|--|---|
| Ferrule | no | | replaceable | |
| Product | MarvelX™ | MarvelXACT™ | Handy connector 2 | Hand-tight EXP® fitting |
| Manufacturer | IDEX Health & Science LLC | IDEX Health & Science LLC | YMC Co., Ltd. | Optimize Technologies, Inc. |
| Image |  |  |  |  |
| Pressure rating | 131 MPa / 1,310 bar | 131 MPa / 1,310 bar | 42 MPa / 420 bar | 137 MPa / 1,370 bar |
| Product code | e.g. UPPF-6050250 | e.g. UPPF-YM7050250 | XRP0204 | XRHTF-01 |

MarvelX (ACT) is a registered trademark of IDEX Health & Science LLC · EXP® is a registered trademark of Optimize Technologies, Inc.

RP – Ordering information

3 µm HPLC columns

| Phase | Column ID [mm] | Column length [mm] | | | | | | Guard cartridges* with 10 mm length |
|--------------------|----------------|--------------------|-----------------|-----------------|-----------------|-----------------|-----------------|-------------------------------------|
| | | 30/33 | 50 | 75 | 100 | 150 | 250 | |
| YMC-Triart C18 | 2.1 | TA12S03-H301PTH | TA12S03-0501PTH | TA12S03-L501PTH | TA12S03-1001PTH | TA12S03-1501PTH | – | TA12S03-0101GC |
| | 3.0 | – | TA12S03-0503PTH | TA12S03-L503PTH | TA12S03-1003PTH | TA12S03-1503PTH | – | TA12S03-0103GC |
| | 4.6 | TA12S03-H346PTH | TA12S03-0546PTH | TA12S03-L546PTH | TA12S03-1046PTH | TA12S03-1546PTH | TA12S03-2546PTH | TA12S03-0104GC |
| YMC-Triart Bio C18 | 2.1 | TA30S03-H301PTH | TA30S03-0501PTH | TA30S03-L501PTH | TA30S03-1001PTH | TA30S03-1501PTH | – | TA30S03-0101GC |
| | 3.0 | – | TA30S03-0503PTH | TA30S03-L503PTH | TA30S03-1003PTH | TA30S03-1503PTH | – | TA30S03-0103GC |
| | 4.6 | TA30S03-H346PTH | TA30S03-0546PTH | TA30S03-L546PTH | TA30S03-1046PTH | TA30S03-1546PTH | TA30S03-2546PTH | TA30S03-0104GC |
| YMC-Triart C8 | 2.1 | T012S03-H301PTH | T012S03-0501PTH | T012S03-L501PTH | T012S03-1001PTH | T012S03-1501PTH | – | T012S03-0101GC |
| | 3.0 | – | T012S03-0503PTH | T012S03-L503PTH | T012S03-1003PTH | T012S03-1503PTH | – | T012S03-0103GC |
| | 4.6 | T012S03-H346PTH | T012S03-0546PTH | T012S03-L546PTH | T012S03-1046PTH | T012S03-1546PTH | T012S03-2546PTH | T012S03-0104GC |
| YMC-Triart Bio C4 | 2.1 | TB30S03-0301PTH | TB30S03-0501PTH | TB30S03-L501PTH | TB30S03-1001PTH | TB30S03-1501PTH | – | TB30S03-0101GC |
| | 3.0 | – | TB30S03-0503PTH | TB30S03-L503PTH | TB30S03-1003PTH | TB30S03-1503PTH | – | TB30S03-0103GC |
| | 4.6 | TB30S03-0346PTH | TB30S03-0546PTH | TB30S03-L546PTH | TB30S03-1046PTH | TB30S03-1546PTH | TB30S03-2546PTH | TB30S03-0104GC |
| Hydrosphere C18 | 2.1 | HS12S03-0301WT | HS12S03-0501WT | HS12S03-L501WT | HS12S03-1001WT | HS12S03-1501WT | HS12S03-2501WT | HS12S03-0101GC |
| | 3.0 | HS12S03-0303WT | HS12S03-0503WT | HS12S03-L503WT | HS12S03-1003WT | HS12S03-1503WT | HS12S03-2503WT | HS12S03-0103GC |
| | 4.6 | HS12S03-0346WT | HS12S03-0546WT | HS12S03-L546WT | HS12S03-1046WT | HS12S03-1546WT | HS12S03-2546WT | HS12S03-0104GC |
| YMCbasic (eq. C8) | 2.1 | BA99S03-0301WT | BA99S03-0501WT | BA99S03-L501WT | BA99S03-1001WT | BA99S03-1501WT | BA99S03-2501WT | BA99S03-0101GC |
| | 3.0 | BA99S03-0303WT | BA99S03-0503WT | BA99S03-L503WT | BA99S03-1003WT | BA99S03-1503WT | BA99S03-2503WT | BA99S03-0103GC |
| | 4.6 | BA99S03-0346WT | BA99S03-0546WT | BA99S03-L546WT | BA99S03-1046WT | BA99S03-1546WT | BA99S03-2546WT | BA99S03-0104GC |

*Guard cartridge holder required, part no. XPGCH-Q1

3 µm metal-free HPLC columns

| Phase | Column ID [mm] | Column length [mm] | | |
|--------------------|----------------|--------------------|-----------------|-----------------|
| | | 50 | 100 | 150 |
| YMC-Triart C18 | 2.1 | TA12S03-0501PTP | TA12S03-1001PTP | TA12S03-1501PTP |
| | 4.6 | TA12S03-0546PTP | TA12S03-1046PTP | TA12S03-1546PTP |
| YMC-Triart Bio C18 | 2.1 | TA30S03-0501PTP | TA30S03-1001PTP | TA30S03-1501PTP |
| | 4.6 | TA30S03-0546PTP | TA30S03-1046PTP | TA30S03-1546PTP |
| YMC-Triart C8 | 2.1 | T012S03-0501PTP | T012S03-1001PTP | T012S03-1501PTP |
| | 4.6 | T012S03-0546PTP | T012S03-1046PTP | T012S03-1546PTP |
| YMC-Triart Bio C4 | 2.1 | TB30S03-0501PTP | TB30S03-1001PTP | TB30S03-1501PTP |
| | 4.6 | TB30S03-0546PTP | TB30S03-1046PTP | TB30S03-1546PTP |

Special column connectors required.

2.7 µm Core-Shell columns

| Phase | Column ID [mm] | Column length [mm] | | | | | Precolumn filter 0.5 µm |
|---------------------------------------|----------------|--------------------|-----------------|-----------------|-----------------|-----------------|-------------------------|
| | | 30 | 50 | 75 | 100 | 150 | |
| | | | | | | | (pack of 3) |
| Meteoritic Core C18 BIO, 16 nm | 2.1 | CAW16SQ7-03Q1PT | CAW16SQ7-05Q1PT | CAW16SQ7-L5Q1PT | CAW16SQ7-10Q1PT | CAW16SQ7-15Q1PT | XRPRCS35 |
| | 3.0 | CAW16SQ7-0303PT | CAW16SQ7-0503PT | CAW16SQ7-L503PT | CAW16SQ7-1003PT | CAW16SQ7-1503PT | |
| | 4.6 | CAW16SQ7-0346PT | CAW16SQ7-0546PT | CAW16SQ7-L546PT | CAW16SQ7-1046PT | CAW16SQ7-1546PT | |

*Holder required, part no. XRPRCS03

5 µm HPLC columns

| Phase | Column ID [mm] | Column length [mm] | | | | | | Guard cartridges* with 10 mm length |
|---------------------------|----------------|--------------------|-----------------|-----------------|-----------------|-----------------|-----------------|-------------------------------------|
| | | 30/33 | 50 | 75 | 100 | 150 | 250 | |
| | | | | | | | | (pack of 5) |
| YMC-Triart C18 | 2.1 | TA12S05-H3Q1PTH | TA12S05-05Q1PTH | TA12S05-L5Q1PTH | TA12S05-10Q1PTH | TA12S05-15Q1PTH | – | TA12S05-01Q1GC |
| | 3.0 | – | TA12S05-0503PTH | TA12S05-L503PTH | TA12S05-1003PTH | TA12S05-1503PTH | – | TA12S05-0103GC |
| | 4.6 | TA12S05-H346PTH | TA12S05-0546PTH | TA12S05-L546PTH | TA12S05-1046PTH | TA12S05-1546PTH | TA12S05-2546PTH | TA12S05-0104GC |
| YMC-Triart Bio C18 | 2.1 | TA30S05-H3Q1PTH | TA30S05-05Q1PTH | TA30S05-L5Q1PTH | TA30S05-10Q1PTH | TA30S05-15Q1PTH | – | TA30S05-01Q1GC |
| | 3.0 | – | TA30S05-0503PTH | TA30S05-L503PTH | TA30S05-1003PTH | TA30S05-1503PTH | – | TA30S05-0103GC |
| | 4.6 | TA30S05-H346PTH | TA30S05-0546PTH | TA30S05-L546PTH | TA30S05-1046PTH | TA30S05-1546PTH | TA30S05-2546PTH | TA30S05-0104GC |
| YMC-Triart C8 | 2.1 | T012S05-H3Q1PTH | T012S05-05Q1PTH | T012S05-L5Q1PTH | T012S05-10Q1PTH | T012S05-15Q1PTH | – | T012S05-01Q1GC |
| | 3.0 | – | T012S05-0503PTH | T012S05-L503PTH | T012S05-1003PTH | T012S05-1503PTH | – | T012S05-0103GC |
| | 4.6 | T012S05-H346PTH | T012S05-0546PTH | T012S05-L546PTH | T012S05-1046PTH | T012S05-1546PTH | T012S05-2546PTH | T012S05-0104GC |
| YMC-Triart Bio C4 | 2.1 | TB30S05-H3Q1PTH | TB30S05-05Q1PTH | TB30S05-L5Q1PTH | TB30S05-10Q1PTH | TB30S05-15Q1PTH | – | TB30S05-01Q1GC |
| | 3.0 | – | TB30S05-0503PTH | TB30S05-L503PTH | TB30S05-1003PTH | TB30S05-1503PTH | – | TB30S05-0103GC |
| | 4.6 | TB30S05-H346PTH | TB30S05-0546PTH | TB30S05-L546PTH | TB30S05-1046PTH | TB30S05-1546PTH | TB30S05-2546PTH | TB30S05-0104GC |
| Hydrosphere C18 | 2.1 | HS12S05-03Q1WT | HS12S05-05Q1WT | HS12S05-L5Q1WT | HS12S05-10Q1WT | HS12S05-15Q1WT | HS12S05-25Q1WT | HS12S05-01Q1GC |
| | 3.0 | HS12S05-0303WT | HS12S05-0503WT | HS12S05-L503WT | HS12S05-1003WT | HS12S05-1503WT | HS12S05-2503WT | HS12S05-0103GC |
| | 4.6 | HS12S05-0346WT | HS12S05-0546WT | HS12S05-L546WT | HS12S05-1046WT | HS12S05-1546WT | HS12S05-2546WT | HS12S05-0104GC |
| YMCbasic (eq. C8) | 2.1 | BA99S05-03Q1WT | BA99S05-05Q1WT | BA99S05-L5Q1WT | BA99S05-10Q1WT | BA99S05-15Q1WT | BA99S05-25Q1WT | BA99S05-01Q1GC |
| | 3.0 | BA99S05-0303WT | BA99S05-0503WT | BA99S05-L503WT | BA99S05-1003WT | BA99S05-1503WT | BA99S05-2503WT | BA99S05-0103GC |
| | 4.6 | BA99S05-0346WT | BA99S05-0546WT | BA99S05-L546WT | BA99S05-1046WT | BA99S05-1546WT | BA99S05-2546WT | BA99S05-0104GC |

*Guard cartridge holder required, part no. XPGCH-Q1

RP – Ordering information

5 µm metal-free HPLC columns

| Phase | Column ID [mm] | Column length [mm] | | |
|---------------------------|----------------|--------------------|-----------------|-----------------|
| | | 50 | 100 | 150 |
| YMC-Triart C18 | 2.1 | TA12S05-05Q1PTP | TA12S05-10Q1PTP | TA12S05-15Q1PTP |
| | 4.6 | TA12S05-0546PTP | TA12S05-1046PTP | TA12S05-1546PTP |
| YMC-Triart Bio C18 | 2.1 | TA30S05-05Q1PTP | TA30S05-10Q1PTP | TA30S05-15Q1PTP |
| | 4.6 | TA30S05-0546PTP | TA30S05-1046PTP | TA30S05-1546PTP |
| YMC-Triart C8 | 2.1 | T012S05-05Q1PTP | T012S05-10Q1PTP | T012S05-15Q1PTP |
| | 4.6 | T012S05-0546PTP | T012S05-1046PTP | T012S05-1546PTP |
| YMC-Triart Bio C4 | 2.1 | TB30S05-05Q1PTP | TB30S05-10Q1PTP | TB30S05-15Q1PTP |
| | 4.6 | TB30S05-0546PTP | TB30S05-1046PTP | TB30S05-1546PTP |

Special column connectors required.

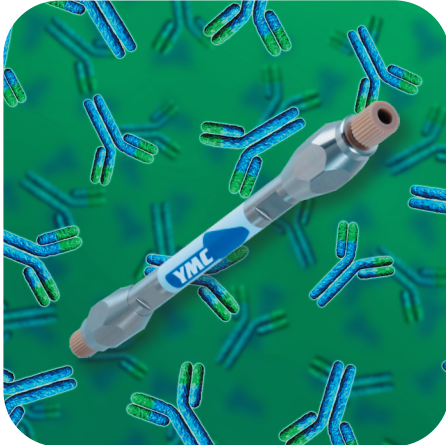
5 µm YMC-Actus high-throughput semipreparative columns

| Phase | Column ID [mm] | Column length [mm] | | | | | Guard cartridges* with 10 mm length |
|---------------------------|----------------|--------------------|----------------|----------------|----------------|----------------|-------------------------------------|
| | | 50 | 75 | 100 | 150 | 250 | (pack of 5) |
| YMC-Triart C18 | 20 | TA12S05-0520WX | TA12S05-L520WX | TA12S05-1020WX | TA12S05-1520WX | TA12S05-2520WX | TA12S05-0120CCN |
| | 30 | TA12S05-0530WX | TA12S05-L530WX | TA12S05-1030WX | TA12S05-1530WX | TA12S05-2530WX | TA12S05-0130CCN |
| | 50 | TA12S05-0553DX | – | TA12S05-1053DX | TA12S05-1553DX | TA12S05-2553DX | TA12S05-0553DXG** |
| YMC-Triart Bio C18 | 20 | TA30S05-0520WX | TA30S05-L520WX | TA30S05-1020WX | TA30S05-1520WX | TA30S05-2520WX | TA30S05-0120CCN |
| | 30 | TA30S05-0530WX | TA30S05-L530WX | TA30S05-1030WX | TA30S05-1530WX | TA30S05-2530WX | TA30S05-0130CCN |
| | 50 | TA30S05-0553DX | – | TA30S05-1053DX | TA30S05-1553DX | TA30S05-2553DX | TA30S05-0553DXG** |
| YMC-Triart C8 | 20 | T012S05-0520WX | T012S05-L520WX | T012S05-1020WX | T012S05-1520WX | T012S05-2520WX | T012S05-0120CCN |
| | 30 | T012S05-0530WX | T012S05-L530WX | T012S05-1030WX | T012S05-1530WX | T012S05-2530WX | T012S05-0130CCN |
| | 50 | T012S05-0553DX | – | T012S05-1053DX | T012S05-1553DX | T012S05-2553DX | T012S05-0553DXG** |
| YMC-Triart Bio C4 | 20 | TB30S05-0520WX | TB30S05-L520WX | TB30S05-1020WX | TB30S05-1520WX | TB30S05-2520WX | TB30S05-0120CCN |
| | 30 | TB30S05-0530WX | TB30S05-L530WX | TB30S05-1030WX | TB30S05-1530WX | TB30S05-2530WX | TB30S05-0130CCN |
| | 50 | TB30S05-0553DX | – | TB30S05-1053DX | TB30S05-1553DX | TB30S05-2553DX | TB30S05-0553DXG** |
| Hydrosphere C18 | 20 | HS12S05-0520WX | HS12S05-L520WX | HS12S05-1020WX | HS12S05-1520WX | HS12S05-2520WX | HS12S05-0120CCN |
| | 30 | HS12S05-0530WX | HS12S05-L530WX | HS12S05-1030WX | HS12S05-1530WX | HS12S05-2530WX | HS12S05-0130CCN |
| | 50 | HS12S05-0553DX | – | HS12S05-1053DX | HS12S05-1553DX | HS12S05-2553DX | – |
| YMCbasic (eq. C8) | 20 | BA99S05-0520WX | BA99S05-L520WX | BA99S05-1020WX | BA99S05-1520WX | BA99S05-2520WX | BA99S05-0120CCN |
| | 30 | BA99S05-0530WX | BA99S05-L530WX | BA99S05-1030WX | BA99S05-1530WX | BA99S05-2530WX | BA99S05-0130CCN |
| | 50 | BA99S05-0553DX | – | BA99S05-1053DX | BA99S05-1553DX | BA99S05-2553DX | – |

*Guard cartridge holder required, part no. XPGHF2P20ID (20 mm ID)
XPGHF2P30ID (30 mm ID)
no holder required for 50 mm



SEC



SEC – UHPLC / HPLC Selectivities

- **Applicable to proteins, antibodies, their fragments and peptides**
- **Also applicable to carbohydrates and nucleic acid components**
- **Excellent reproducibility with minimal secondary interactions**
- **2 µm for UHPLC**
- **Cost effective**

| | YMC-Pack Diol-60 | YMC-Pack Diol-120 | YMC-Pack Diol-200 | YMC-Pack Diol-300 | YMC-SEC MAB |
|---------------------------|---|---------------------------|--------------------|-------------------------|--|
| | For peptides and small proteins | For intermediate proteins | For large proteins | For very large proteins | For antibodies, fragments and aggregates |
| Base particle | Silica | | | | |
| Particle Size / µm | 3, 5 | 3, 5 | 2, 3, 5 | 2, 3, 5 | 3 |
| Pore Size / nm | 6 | 12 | 20 | 30 | 25 |
| Modification | Dihydroxypropyl | | | | |
| Temperature range | 40 °C | | | | |
| Pressure limit | 2 µm: 45 MPa (6,525 psi); 3/5 µm: 20 MPa (3,000 psi) | | | | 14 MPa (2,030 psi) |

“

“The YMC-Pack Diol SEC column has been successfully used for subsequent method validation.”

Rubén Pedrosa Segon, Head of Quality Control Pharmaceutical Department, OFICE S.L. (ES)

”

Column Selection Tool

for MW < 10,000

● YMC-Pack Diol-60

for MW 1,000 to 100,000

● YMC-Pack Diol-120

for MW 5,000 to 300,000

● YMC-Pack Diol-200

for MW 10,000 to 700,000

● YMC-SEC MAB

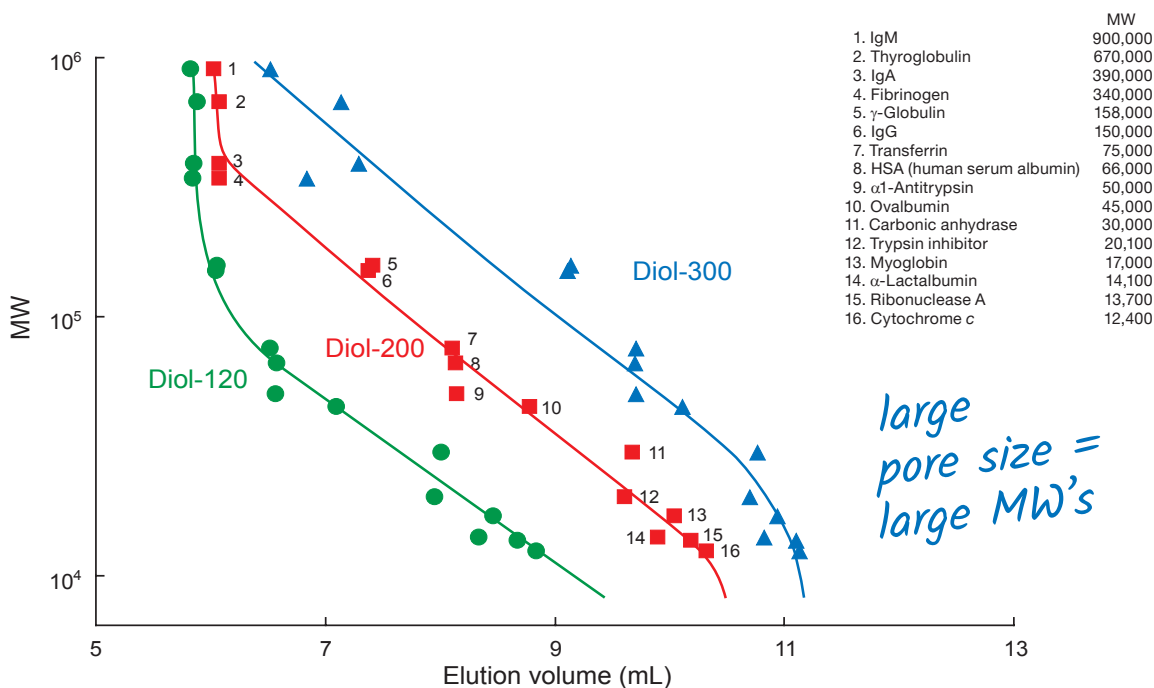
for MW 20,000 to 1,000,000

● YMC-Pack Diol-300

SEC – YMC-Pack Diol: Phase selection for proteins

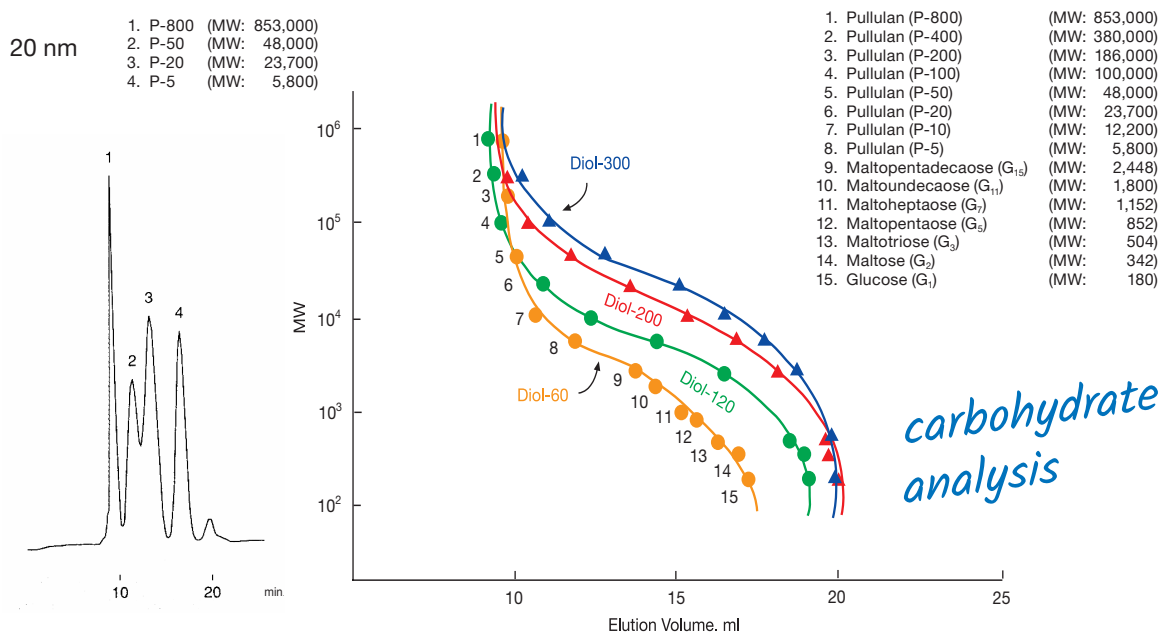
Phases for different MW ranges

For separation of proteins with molecular weights from 10,000 to several 100,000 Da



Column: YMC-Pack Diol, 300 x 8.0 mm ID
 Part Nos.: DL12S05-3008WT, DL20S05-3008WT, DL30S05-5008WT
 Eluent: 0.1 M KH_2PO_4 - K_2HPO_4 (pH 7.0) containing 0.2 M NaCl
 Flow rate: 0.5 mL/min
 Temperature: 25 °C
 Detection: UV at 280 nm

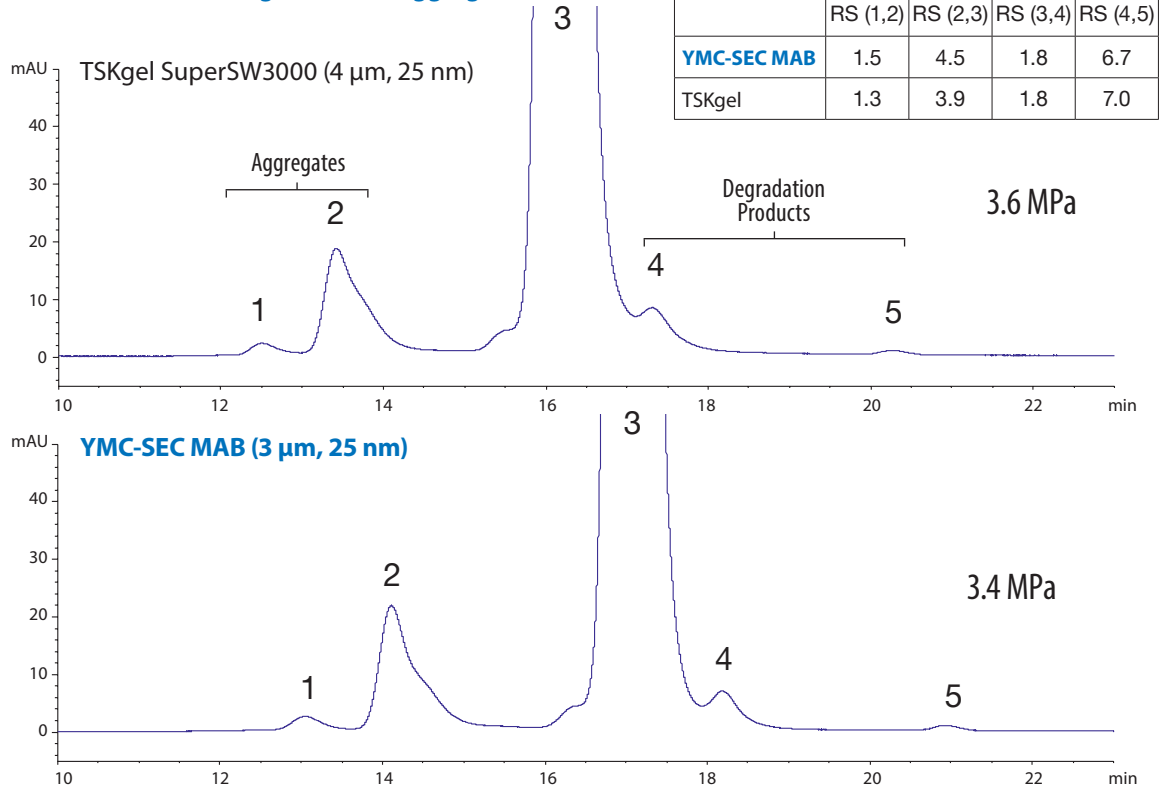
For molecular weight determination of oligosaccharides and polysaccharides



Column: YMC-Pack Diol (20 nm) 500 x 8.0 mm ID
 Part No.: DL20S05-5008WT
 Eluent: water
 Flow rate: 1.0 mL/min
 Temperature: ambient
 Detection: RI

Ideal choice for monoclonal antibodies

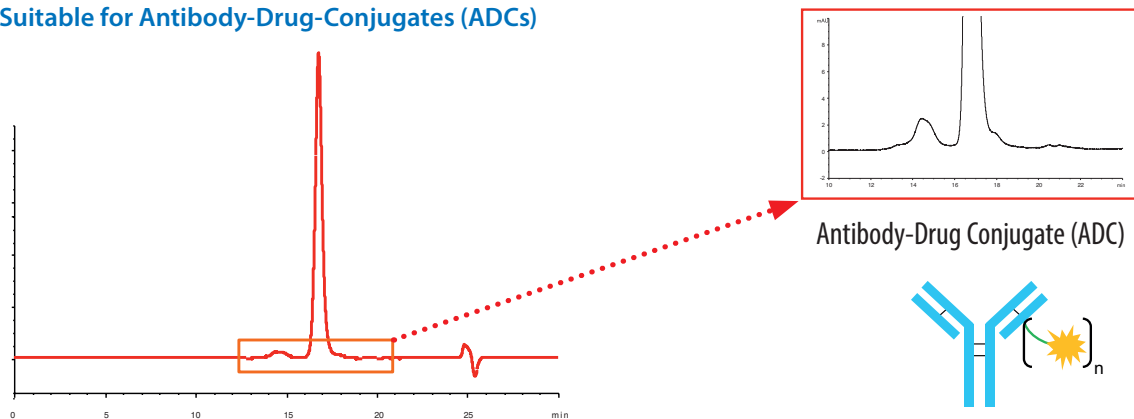
Bevacizumab and its fragments and aggregates



Column: 300 x 4.6 mm ID
 Part No.: DLM25S03-3046WT
 Eluent: 0.1 M phosphate buffer (pH = 7) cont. 0.2 M NaCl
 Flow rate: 0.165 mL/min
 Temperature: 25 °C

Detection: UV at 280 nm
 Cell path: 10 mm
 Injection: 10 µL (5 mg/mL)
 Sample: Bevacizumab (Avastin®)

Suitable for Antibody-Drug-Conjugates (ADCs)



Column: YMC-SEC MAB (3 µm, 25 nm) 300 x 4.6 mm ID
 Part No.: DLM25S03-3046WT
 Eluent: 0.1 M phosphate buffer (pH = 7) cont. 0.2 M NaCl /
 2-propanol (85 / 15)
 Flow rate: 0.165 mL/min

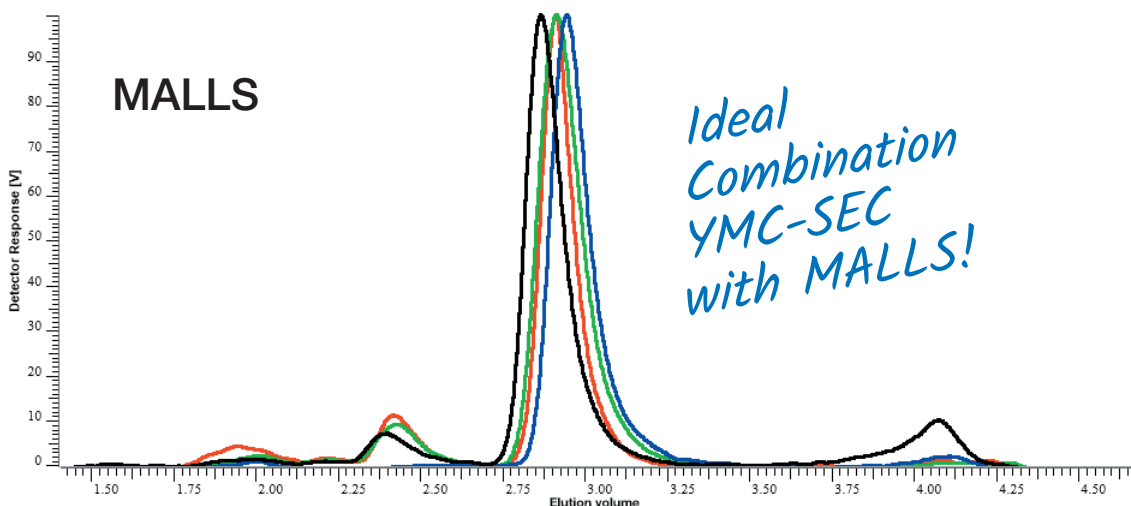
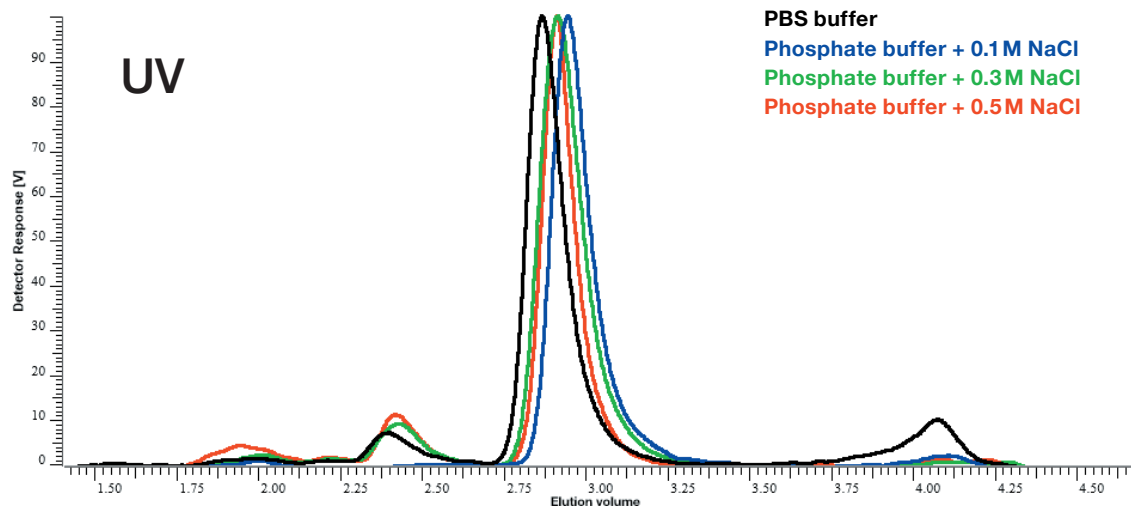
Temperature: 25 °C
 Detection: UV at 280 nm
 Injection: 4 µL (2.5 mg/mL)
 Sample: SigmaMAb Antibody Drug Conjugate Mimic

YMC-SEC MAB is also suitable for the analysis of Antibody-Drug Conjugates (ADC). The addition of an organic solvent to the mobile phase can improve the results obtained for ADC analysis.

SEC – YMC-SEC MAB: MALLS

YMC-SEC columns ideally combined with light scattering detection

Detection of higher molar mass species by MALLS



Column: YMC-SEC MAB (3 μ m, 25 nm) 300 x 4.6 mm ID
 Part No.: DLM25S03-3046WT
 Eluent: Phosphate buffer pH 6.6 containing 0.3 M NaCl
 Flow rate: 0.33 mL/min
 Temperature: 25 °C
 Detection: MALLS at 90° angle (PSS SLD7100), UV at 280 nm
 Injection volume: 10 μ L
 Sample: Bevacizumab (Avastin®) dosage form (10 mg/mL, diluted to 1 mg/mL)
 System: PSS-SECcurity GPC systems, 1260 Infinity II
 Software: WinGPC Unichrom

Courtesy of Thorsten Hofe, PSS Polymer Standards Service GmbH, Mainz, Germany.

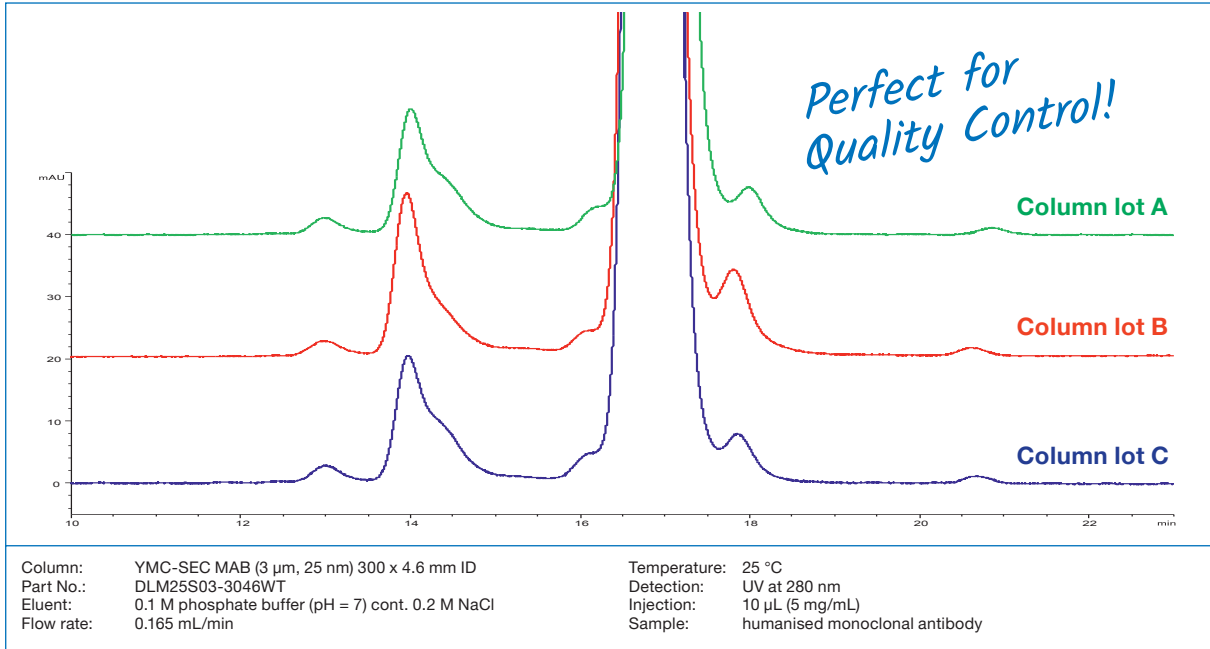
Four different buffers, a phosphate buffered saline (PBS) pH 7.4 and phosphate buffers pH 6.6 with varying concentrations of NaCl, were used to develop a suitable MALLS detection method for MABs.

A defined minimum ionic strength is necessary to achieve a robust method with good resolution. The phosphate buffer with 0.3 M NaCl appeared to be the most suitable eluent.

Compared to UV detection, the MALLS signal shows 2 higher molar mass species, aggregates of Bevacizumab, at about 2.0 mL and 2.3 mL elution volume.

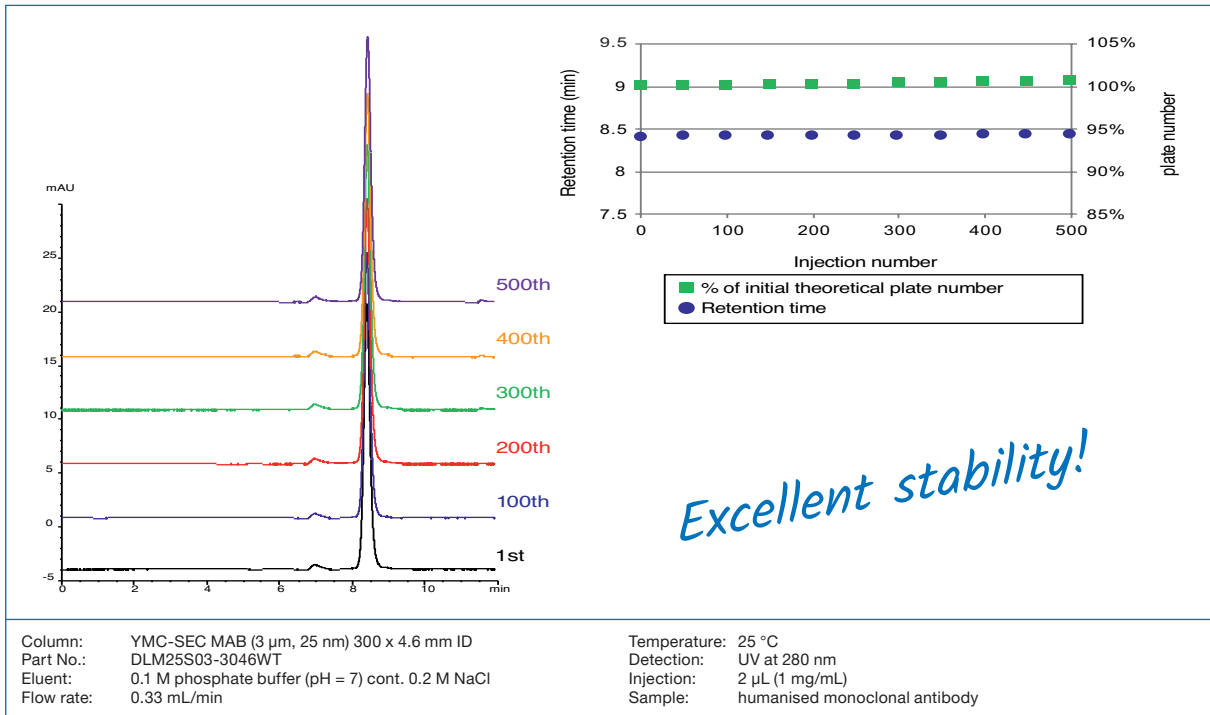
SEC – YMC-SEC MAB: Reproducibility & stability

Excellent lot-to-lot reproducibility



YMC-SEC MAB provides excellent reproducibility for the separation of monomer and aggregates as well as for monomer and their fragments, making it very effective for quality control of antibody drugs.

High column stability



Excellent stability is provided for monoclonal antibody analysis without any changes in theoretical plate number or elution time even after more than 500 injections.

SEC – YMC-Pack Diol: Resolution & throughput

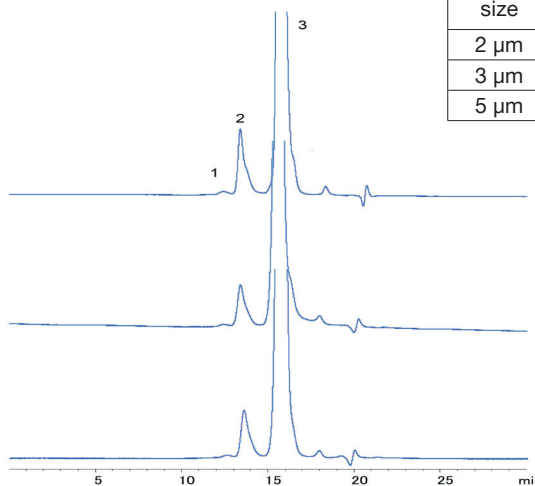
Benefits of using smaller particles

Higher resolution for analysis of monoclonal antibodies

(A) YMC-Pack Diol-300 (2 μm)
300 x 4.6 mm ID

(B) YMC-Pack Diol-300 (3 μm)
300 x 4.6 mm ID

(C) YMC-Pack Diol-300 (5 μm)
300 x 4.6 mm ID



| Particle size | Rs (1,2) | Rs (2,3) | N (3) |
|-----------------|----------|----------|--------------|
| 2 μm | 1.17 | 4.15 | 16,200 |
| 3 μm | 1.03 | 3.18 | 10,400 |
| 5 μm | 0.88 | 2.67 | 8,500 |

Columns: YMC-Pack Diol-300, 300 x 4.6 mm ID
Part Nos.: (A) DL30S02-3046PTH
(B) DL30S03-3046WT
(C) DL30S05-3046WT
Eluent: 0.1 M KH_2PO_4 - K_2HPO_4 (pH 7.0) containing 0.2 M NaCl

Flow rate: 0.2 mL/min
Temperature: ambient
Detection: UV at 280 nm
Sample: Humanised monoclonal antibody (IgG1)

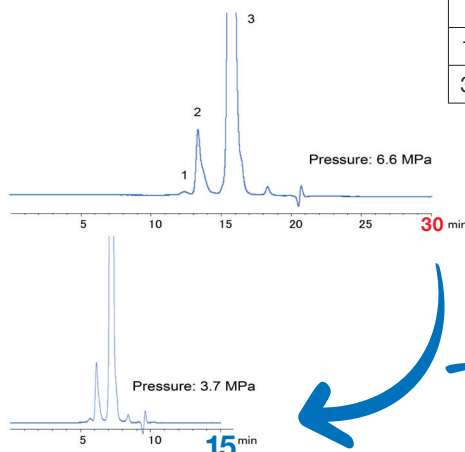
All three particle sizes show identical separation patterns for monoclonal antibody analysis. This allows easy method transfer between HPLC and UHPLC. A method developed using conventional HPLC can be directly transferred to UHPLC using a 2 μm YMC-Pack Diol

column. YMC-Pack Diol UHPLC columns greatly improve the resolution between aggregates and the monomer peak. In addition, a shoulder peak which can be observed after the monomer peak can be partially separated using the 2 μm column.

High throughput analysis of monoclonal antibodies

YMC-Pack Diol-300 (2 μm)
300 x 4.6 mm ID

YMC-Pack Diol-300 (2 μm)
150 x 4.6 mm ID



| Column length | Rs (1,2) | Rs (2,3) | N (3) |
|---------------|----------|----------|--------|
| 150 mm | 0.85 | 2.75 | 8,700 |
| 300 mm | 1.17 | 4.15 | 16,200 |

Columns: YMC-Pack Diol-300, 150 or 300 x 4.6 mm ID
Part Nos.: DL30S02-3046PTH / DL30S02-1546PTH
Eluent: 0.1 M KH_2PO_4 - K_2HPO_4 (pH 7.0) containing 0.2 M NaCl
Flow rate: 0.2 mL/min

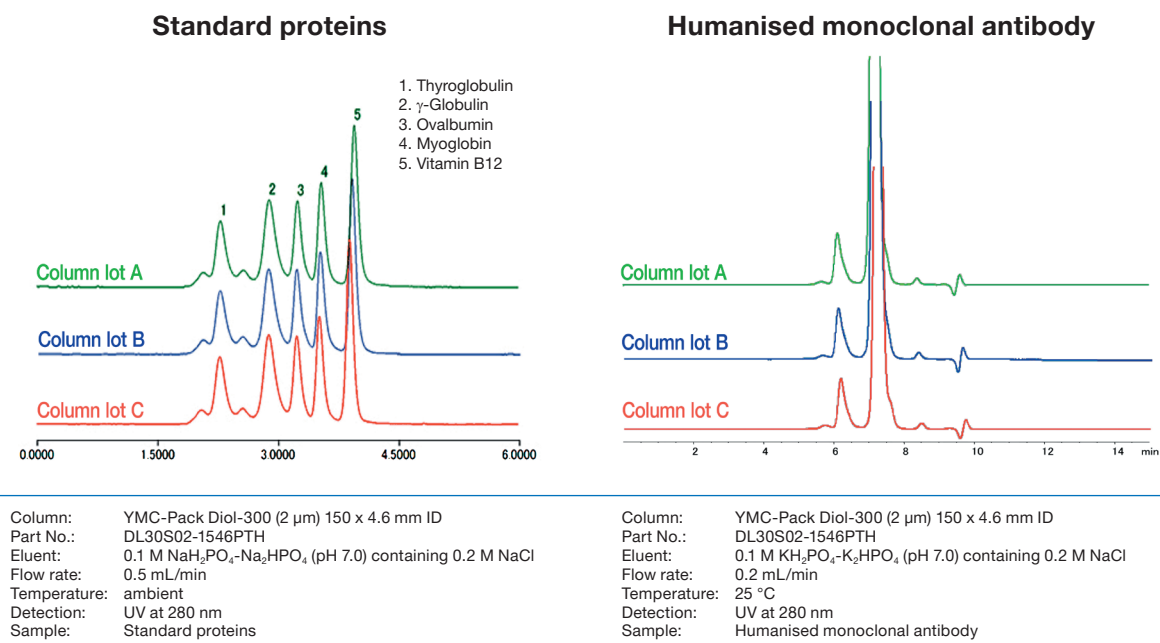
Temperature: ambient
Detection: UV at 280 nm
Sample: Humanised monoclonal antibody (IgG1)

By using a 150 mm length column, 50% shorter run times can be achieved with the same resolution as for a 300 mm length column (compare upper and lower chromatograms). This allows an increase in throughput to be achieved. The backpressure is only 6.6 MPa, even for the 300 mm column. Therefore, YMC-Pack Diol 2 μm columns can be used with both UHPLC and HPLC systems.

SEC – YMC-Pack Diol: Reproducibility & stability

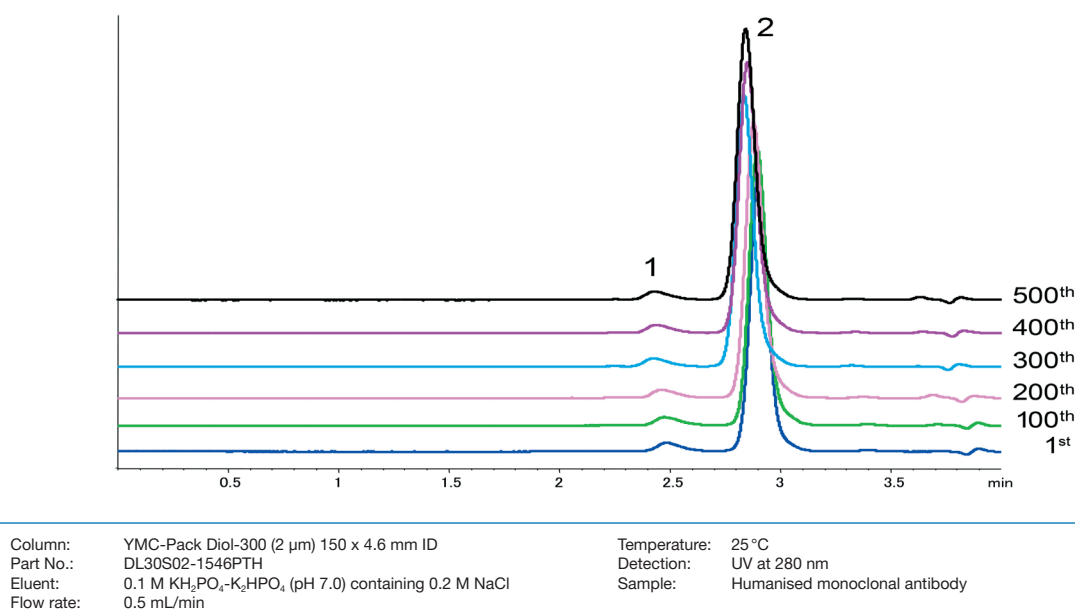
Reproducibility and stability data

Excellent batch-to-batch reproducibility



YMC-Pack Diol UHPLC columns have excellent batch-to-batch reproducibility. This makes YMC-Pack Diol 2 μ m columns the ideal choice for the quality control of bio-based drugs including monoclonal antibodies.

Long-term stability



YMC-Pack Diol UHPLC columns maintain their performance for more than 500 injections of sample during monoclonal antibody analysis. This ensures reproducible and reliable quality control of bio-based drugs including monoclonal antibodies.

SEC – Ordering information

2 µm UHPLC columns

| Phase | Column ID [mm] | Column length [mm] | | Guard cartridges* with 10 mm length (pack of 5) |
|-------------------|----------------|--------------------|-----------------|--|
| | | 150 | 300 | |
| YMC-Pack Diol-200 | 4.6 | DL20S02-1546PTH | DL20S02-3046PTH | DL20S02-0104GC |
| YMC-Pack Diol-300 | 4.6 | DL30S02-1546PTH | DL30S02-3046PTH | DL30S02-0104GC |

*Guard cartridge holder required, part no. XPGCH-Q1

3 µm HPLC columns

| Phase | Column ID [mm] | Column length [mm] | | | Guard cartridges* with 10/30 mm length (pack of 5) |
|-------------------|----------------|--------------------|----------------|-----------------|---|
| | | 150 | 250 | 300 | |
| YMC-SEC MAB | 4.6 | DLM25S03-1546WT | – | DLM25S03-3046WT | DLM25S03-0104GC |
| | 6.0 | – | – | – | – |
| | 8.0 | – | – | DLM25S03-3008WT | – |
| YMC-Pack Diol-60 | 4.6 | DL06S03-1546WT | DL06S03-2546WT | DL06S03-3046WT | DL06S03-0104GC |
| | 6.0 | – | – | DL06S03-3006WT | – |
| | 8.0 | DL06S03-1508WT | – | DL06S03-3008WT | DL06S03-0308WTG** |
| YMC-Pack Diol-120 | 4.6 | DL12S03-1546WT | DL12S03-2546WT | DL12S03-3046WT | DL12S03-0104GC |
| | 6.0 | – | – | DL12S03-3006WT | – |
| | 8.0 | DL12S03-1508WT | – | DL12S03-3008WT | DL12S03-0308WTG** |
| YMC-Pack Diol-200 | 4.6 | DL20S03-1546WT | DL20S03-2546WT | DL20S03-3046WT | DL20S03-0104GC |
| | 6.0 | – | – | DL20S03-3006WT | – |
| | 8.0 | DL20S03-1508WT | – | DL20S03-3008WT | DL20S03-0308WTG** |
| YMC-Pack Diol-300 | 4.6 | DL30S03-1546WT | DL30S03-2546WT | DL30S03-3046WT | DL30S03-0104GC |
| | 6.0 | – | – | DL30S03-3006WT | – |
| | 8.0 | DL30S03-1508WT | – | DL30S03-3008WT | DL30S03-0308WTG** |

*Guard cartridge holder required, part no. XPGCH-Q1

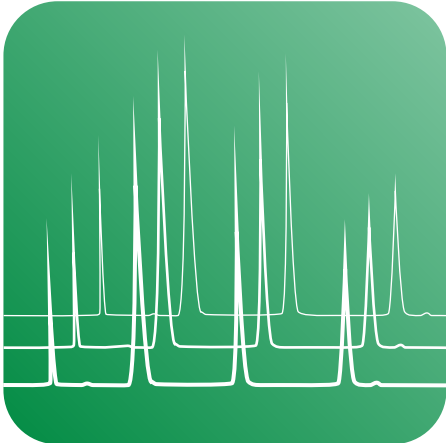
**no holder required for 30 x 8 mm ID guards

5 µm HPLC columns

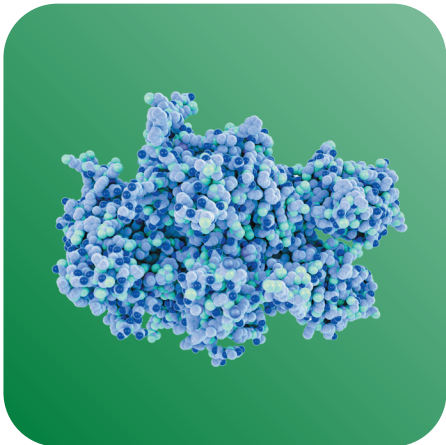
| Phase | Column ID [mm] | Column length [mm] | | | Guard cartridges* with 10/30 mm length (pack of 5) |
|-------------------|----------------|--------------------|----------------|----------------|---|
| | | 250 | 300 | 500 | |
| YMC-Pack Diol-60 | 4.6 | DL06S05-2546WT | DL06S05-3046WT | – | DL06S05-0104GC |
| | 6.0 | DL06S05-2506WT | DL06S05-3006WT | DL06S05-5006WT | – |
| | 8.0 | – | DL06S05-3008WT | DL06S05-5008WT | DL06S05-0308WTG** |
| | 10.0 | DL06S05-2510WT | DL06S05-3010WT | DL06S05-5010WT | DL06S05-0310WTG** |
| YMC-Pack Diol-120 | 4.6 | DL12S05-2546WT | DL12S05-3046WT | – | DL12S05-0104GC |
| | 6.0 | DL12S05-2506WT | DL12S05-3006WT | DL12S05-5006WT | – |
| | 8.0 | – | DL12S05-3008WT | DL12S05-5008WT | DL12S05-0308WTG** |
| | 10.0 | DL12S05-2510WT | DL12S05-3010WT | DL12S05-5010WT | DL12S05-0310WTG** |
| YMC-Pack Diol-200 | 4.6 | DL20S05-2546WT | DL20S05-3046WT | – | DL20S05-0104GC |
| | 6.0 | DL20S05-2506WT | DL20S05-3006WT | DL20S05-5006WT | – |
| | 8.0 | – | DL20S05-3008WT | DL20S05-5008WT | DL20S05-0308WTG** |
| | 10.0 | DL20S05-2510WT | DL20S05-3010WT | DL20S05-5010WT | DL20S05-0310WTG** |
| YMC-Pack Diol-300 | 4.6 | DL30S05-2546WT | DL30S05-3046WT | – | DL30S05-0104GC |
| | 6.0 | DL30S05-2506WT | DL30S05-3006WT | DL30S05-5006WT | – |
| | 8.0 | – | DL30S05-3008WT | DL30S05-5008WT | DL30S05-0308WTG** |
| | 10.0 | DL30S05-2510WT | DL30S05-3010WT | DL30S05-5010WT | DL30S05-0310WTG** |

*Guard cartridge holder required, part no. XPGCH-Q1

**no holder required for 30 x 8/10 mm ID guards

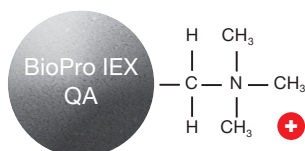


IEX

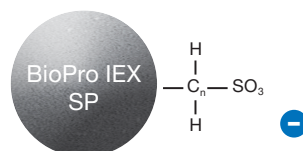


IEX – Bio Pro Series

- porous or non-porous hydrophilic polymers
- high binding capacity and recovery of biomolecules
- very high resolution
- low nonspecific adsorption
- excellent reproducibility

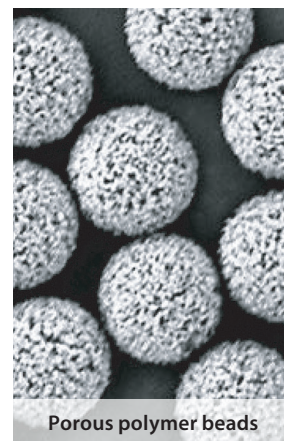


strong anion exchanger



strong cation exchanger

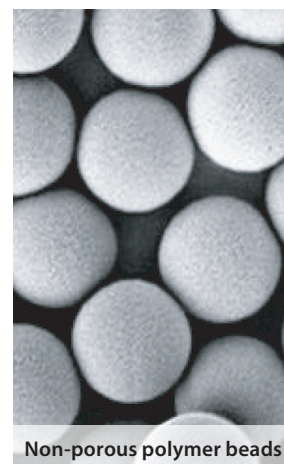
| | BioPro IEX QA | BioPro IEX SP |
|-------------------------------|---|--|
| Matrix | hydrophilic polymer (polymethacrylate) | hydrophilic polymer (polymethacrylate) |
| Particle size / μm | 5 | 5 |
| Pore size / nm | 100 | 100 |
| Charged group | $-\text{CH}_2\text{N}^+(\text{CH}_3)_3$ | $-(\text{CH}_2)_3\text{SO}_3^-$ |
| Counter ion | Cl^- | Na^+ |
| Available pH range | 2.0–12.0 | 2.0–12.0 |
| Temperature range | 4–60 °C | |
| Pressure limit | 2.5–3.5 MPa (360–510 psi) | |
| Column hardware | PEEK | |



Porous polymer beads

Also available in 10, 20, 30 or 75 μm for preparative scale

| | BioPro IEX QF | BioPro IEX SF |
|-------------------------------|--|--|
| Matrix | hydrophilic polymer (polymethacrylate) | hydrophilic polymer (polymethacrylate) |
| Particle size / μm | 3, 5 | 3, 5 |
| Pore size / nm | non-porous | non-porous |
| Charged group | $-\text{CH}_2\text{N}^+(\text{CH}_3)_3$ | $-(\text{CH}_2)_3\text{SO}_3^-$ |
| Counter ion | Cl^- | Na^+ |
| Available pH range | 2.0–12.0 | 2.0–2.0 |
| Temperature range | 4–60 °C | |
| Pressure limit | 3 μm : 25 MPa (3,625 psi) 5 μm : 6–12 MPa (870–1,740 psi) | |
| Column hardware | PEEK | |



Non-porous polymer beads

YMC's BioPro IEX series of ion exchange columns are available in QA and SP chemistries, based on 5 μm porous (QA or SP columns) or on 3 or 5 μm non-porous (QF and SF columns) hydrophilic polymer beads.

The porous materials offer excellent binding capacity with exceptionally high efficiency and low operating pressure, whilst the non-porous particles offer high efficiency, very high resolution and low operating pressures.

IEX – BioPro IEX: Reproducibility & DBC

High binding capacity and high recovery for porous type

The porous versions of YMC's BioPro IEX show high dynamic binding capacity and excellent recovery, making them useful for semi-preparative separations of proteins and antibodies.

Comparison of dynamic binding capacity (DBC) for BSA

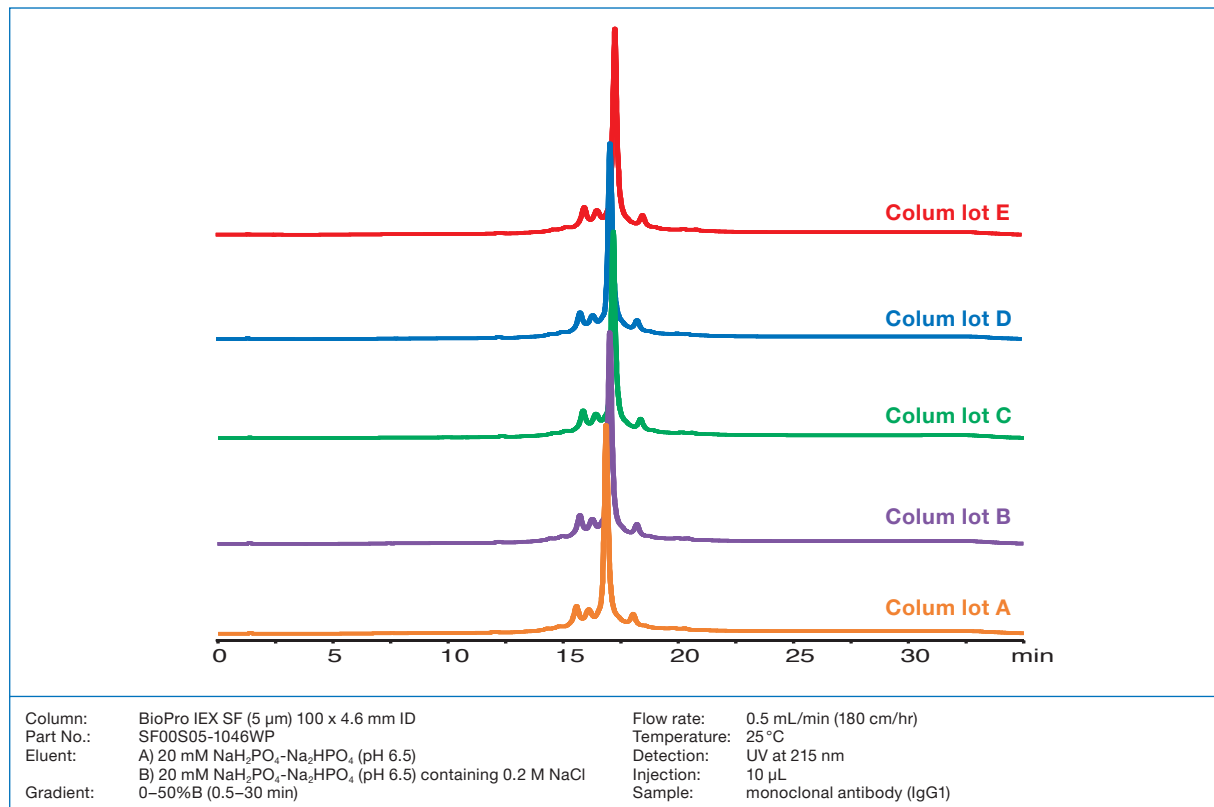
| | Dynamic binding capacity (mg/mL-gel, 10% breakthrough) | Eluted amount (mg/mL-gel) | Recovery* (%) |
|--------------------------------|--|---------------------------|---------------|
| BioPro IEX QA | 126 | 120 | 95 |
| Mono Q (GE Healthcare) | 100 | 35 | 35 |
| BioAssist Q (Tosoh Bioscience) | 73 | 58 | 79 |

High recovery rates for BioPro IEX

* Recovery: (Eluted amount/Dynamic binding capacity) x 100

Compared with conventional porous polymer anion exchange columns, BioPro IEX QA provides higher DBC and recovery rates. This indicates that BioPro IEX has a much lower nonspecific adsorption compared to conventional columns.

Excellent batch-to-batch reproducibility



BioPro IEX SF columns exhibit excellent batch-to-batch reproducibility for MAb analysis with resolution of peaks for small charge variants. All gel batches are inspected by rigorous quality control tests, including HPLC analysis of MAb, and must meet the required criteria before release.

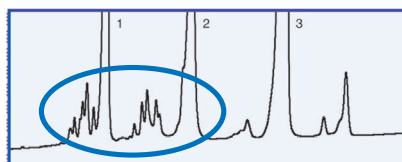
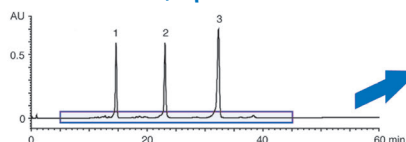
BioPro IEX columns are the best choice for the quality control of MAb and other biopharmaceuticals.

IEX – BioPro IEX: Resolution & throughput

Superior resolution

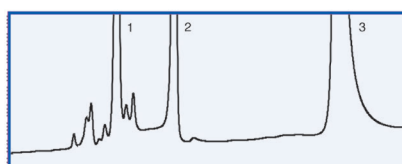
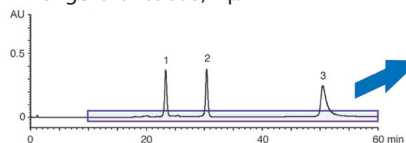
Comparison of standard protein separation on BioPro IEX SP and commercial S type products

BioPro IEX SP, 5 µm

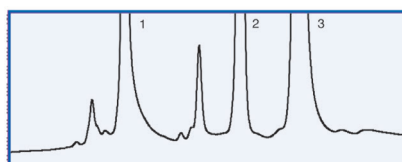
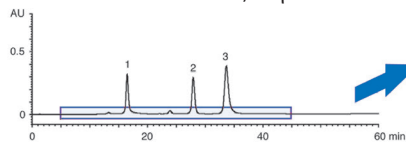


Superior resolution

TSKgel BioAssist S, 7 µm



GE Healthcare Mono S, 10 µm



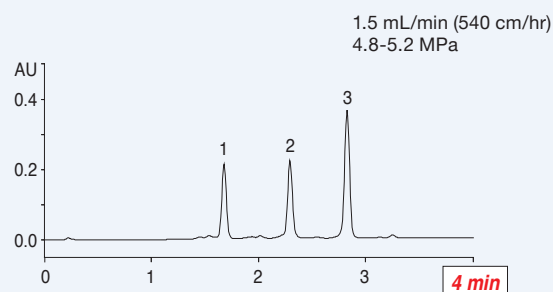
Eluent: A) 20 mM KH_2PO_4 - K_2HPO_4 (pH 6.8)
 B) 20 mM KH_2PO_4 - K_2HPO_4 (pH 6.8) containing 0.5 M NaCl
 Gradient: 0–100%B (0–60 min)
 Flow rate: BioPro IEX SP, TSKgel BioAssist S 0.5 mL/min
 GE Healthcare Mono S 0.59 mL/min
 Temperature: 25 °C

Detection: UV at 220 nm
 Injection: BioPro IEX SP, TSKgel BioAssist S 20 µL
 GE Healthcare Mono S 23.6 µL
 Sample: 1. Ribonuclease A (0.5 mg/mL)
 2. Cytochrome c (0.5 mg/mL)
 3. Lysozyme (0.5 mg/mL)

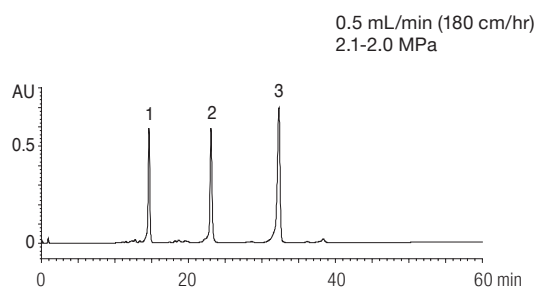
Only BioPro IEX is available in the smaller particle size and is therefore able to provide superior resolution.

Ultra-high-throughput analysis with non-porous BioPro IEX

Non-porous type BioPro IEX SF (5 µm) 30 x 4.6 mm ID



Porous type BioPro IEX SP (5 µm) 50 x 4.6 mm ID



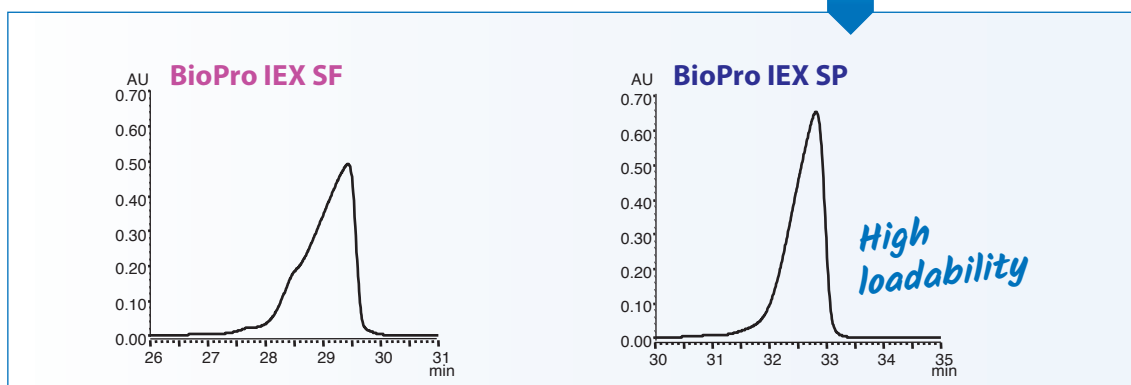
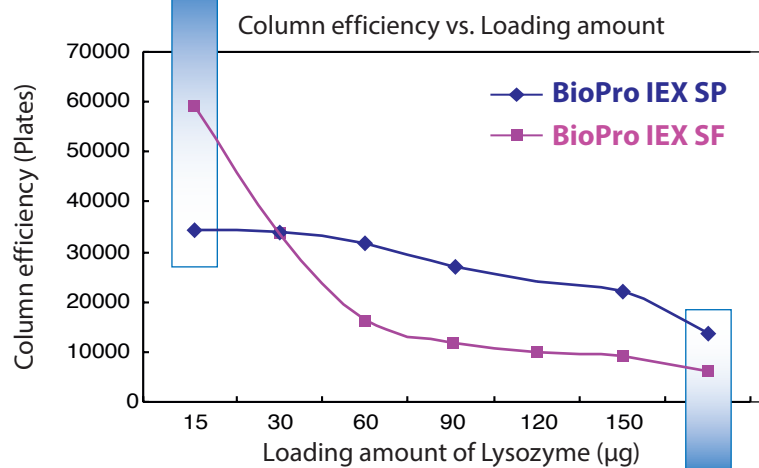
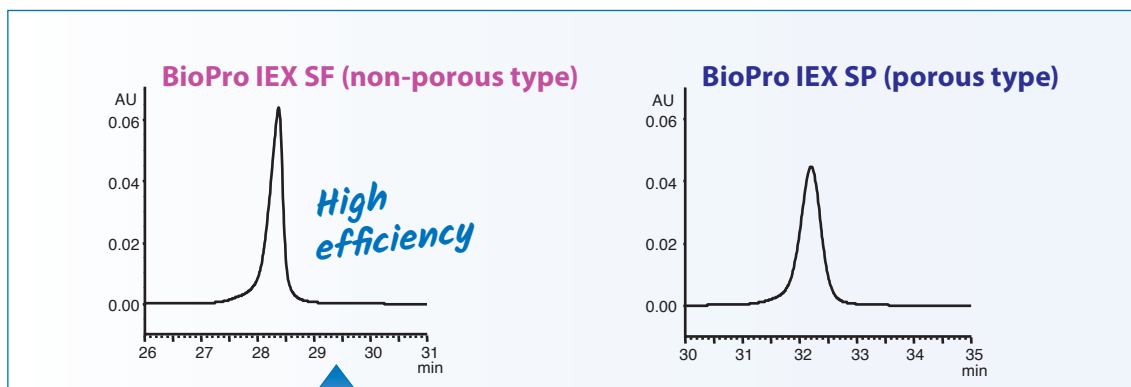
Eluent: A) 20 mM KH_2PO_4 - K_2HPO_4 (pH 6.8)
 B) 20 mM KH_2PO_4 - K_2HPO_4 (pH 6.8) containing 0.5 M NaCl
 Part Nos.: SF00S05-0346WP (non-porous)
 SPA0S05-0546WP (porous)
 Gradient: 0–100%B (0–4 min) for BioPro IEX SF
 0–100%B (0–60 min) for BioPro IEX SP

Temperature: 25 °C
 Detection: UV at 220 nm
 Injection: 20 µL
 Sample: 1. Ribonuclease A (0.5 mg/mL)
 2. Cytochrome c (0.5 mg/mL)
 3. Lysozyme (0.5 mg/mL)

The high mechanical stability of non-porous polymer beads and the short column length allow faster elution of proteins at a higher flow rate without any loss of resolution.

Column efficiency and loadability

When to use porous and non-porous BioPro IEX



Eluent: A) 20 mM NaH₂PO₄-Na₂HPO₄ (pH 6.8)
 B) 20 mM NaH₂PO₄-Na₂HPO₄ (pH 6.8) containing 0.5 M NaCl
 Gradient: 0–100%B (0–60 min)
 Flow rate: 0.5 mL/min
 Temperature: 25 °C

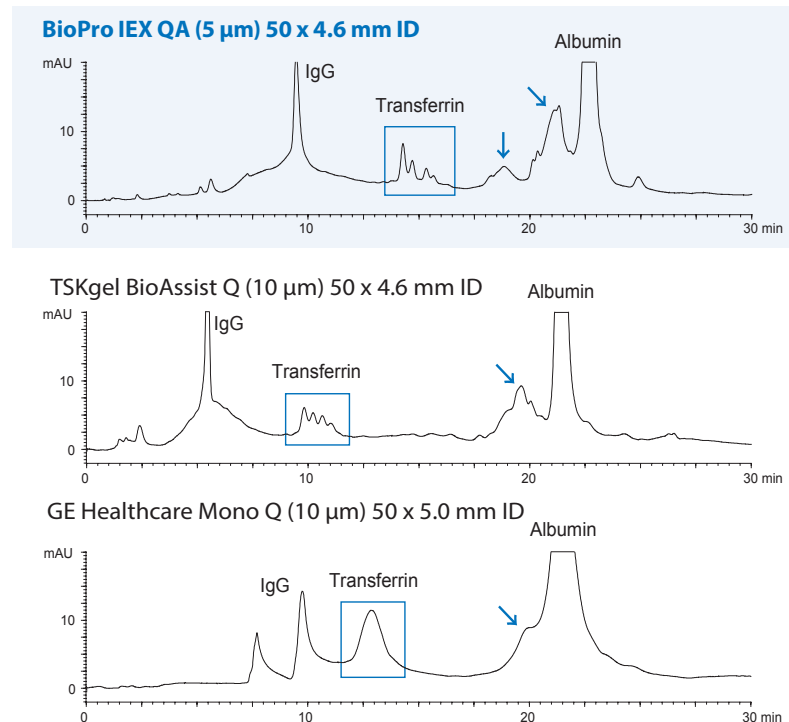
Detection: UV at 280 nm
 Injection: 100 µL
 Sample: 1. Ribonuclease A
 2. Cytochrome c
 3. Lysozyme

BioPro IEX SF offers outstanding column efficiency at small amount of sample loading. Non-porous type of BioPro IEX columns are especially suitable for microscale analysis which requires higher resolution. BioPro IEX SP maintains the good peak shape even when the loading amount increases. Porous type BioPro IEX columns with high capacity are useful for high-load analytical separations and laboratory-scale purification.

IEX – BioPro IEX: Challenging separations

Protein separation in challenging matrices

Separation of proteins in human serum on BioPro IEX QA and commercial Q-type products

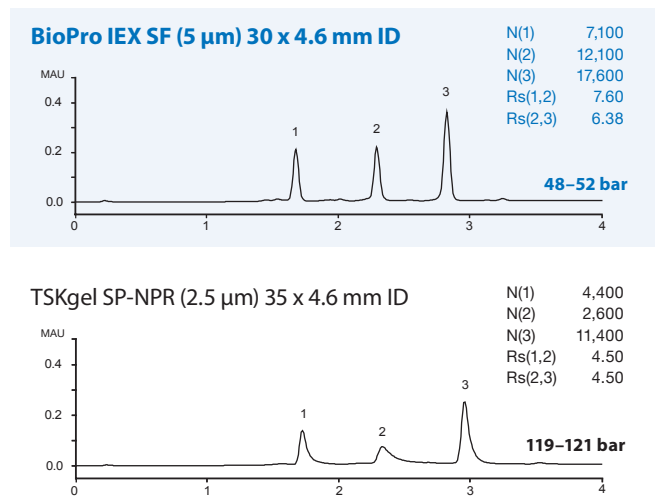


For high resolution porous BioPro IEX QA/SP is recommended!

| | | | |
|-----------|--|--------------|-------------------------|
| Part No.: | QAA0S05-0546WP | Flow rate: | 0.5 mL/min |
| Eluent: | A) 20 mM Tris-HCl (pH 8.6) B) 20 mM Tris-HCl (pH 8.6) containing 0.5 M NaCl | Temperature: | 25 °C |
| Gradient: | 0–30%B (0–15 min), 30–100%B (15–30 min) | Detection: | UV at 280 nm |
| | | Injection: | 20 μL |
| | | Sample: | Human serum (100 μL/mL) |

Better performance at lower backpressure

Comparison of standard protein separation on BioPro IEX SF and a commercial SP-type product



BioPro IEX SF elutes the proteins in sharper peaks without peak-tailing compared to TSKgel SP-NPR. Despite the larger particle size, the theoretical plate count for BioPro IEX SF is higher than that for TSKgel SP-NPR.

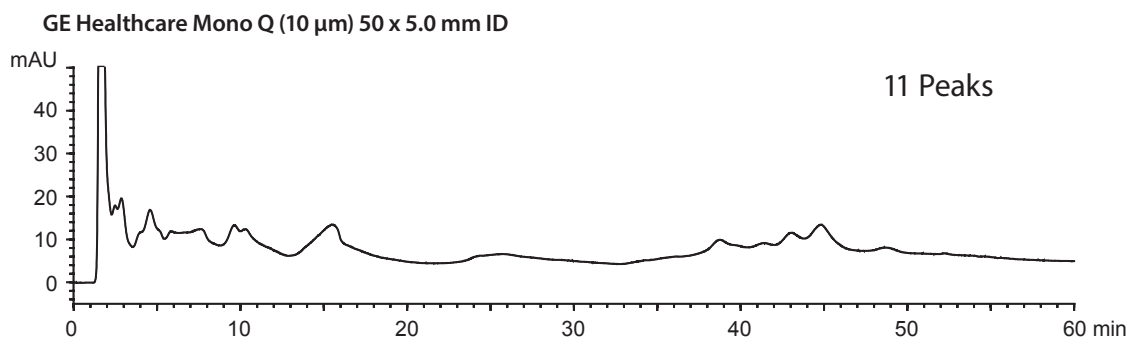
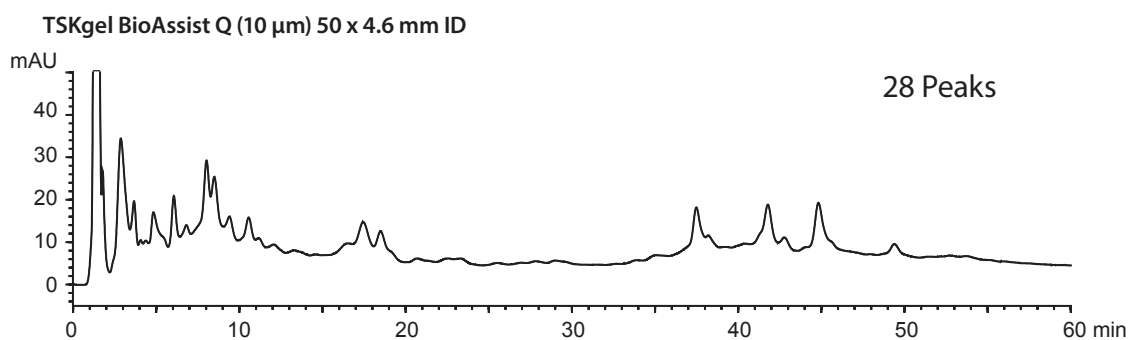
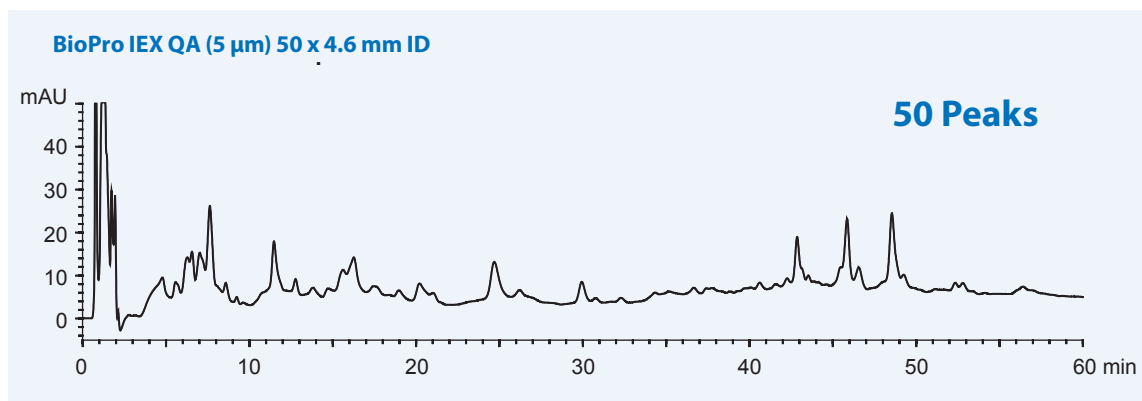
higher plate count

| | |
|--------------|--|
| Part No.: | SF00S05-0346WP |
| Eluent: | A) 20 mM KH ₂ PO ₄ -K ₂ HPO ₄ (pH 6.8) B) 20 mM KH ₂ PO ₄ -K ₂ HPO ₄ (pH 6.8) containing 0.5 M NaCl |
| Gradient: | BioPro IEX SF 0-100%B (0–4 min) TSKgel SP-NPR 0-100%B (0–4.67 min) |
| Flow rate: | 1.5 mL/min |
| Temperature: | 25 °C |
| Detection: | UV at 220 nm |
| Injection: | 20 μL |
| Sample: | 1. Ribonuclease A (0.1 mg/mL) 2. Cytochrome c (0.1 mg/mL) 3. Lysozyme (0.1 mg/mL) |

Compared to the competitor's column, BioPro IEX SF gives higher theoretical plate counts, excellent peak shapes, and lower backpressures. This makes BioPro IEX SF most suitable for high-throughput analysis.

Peptide mapping

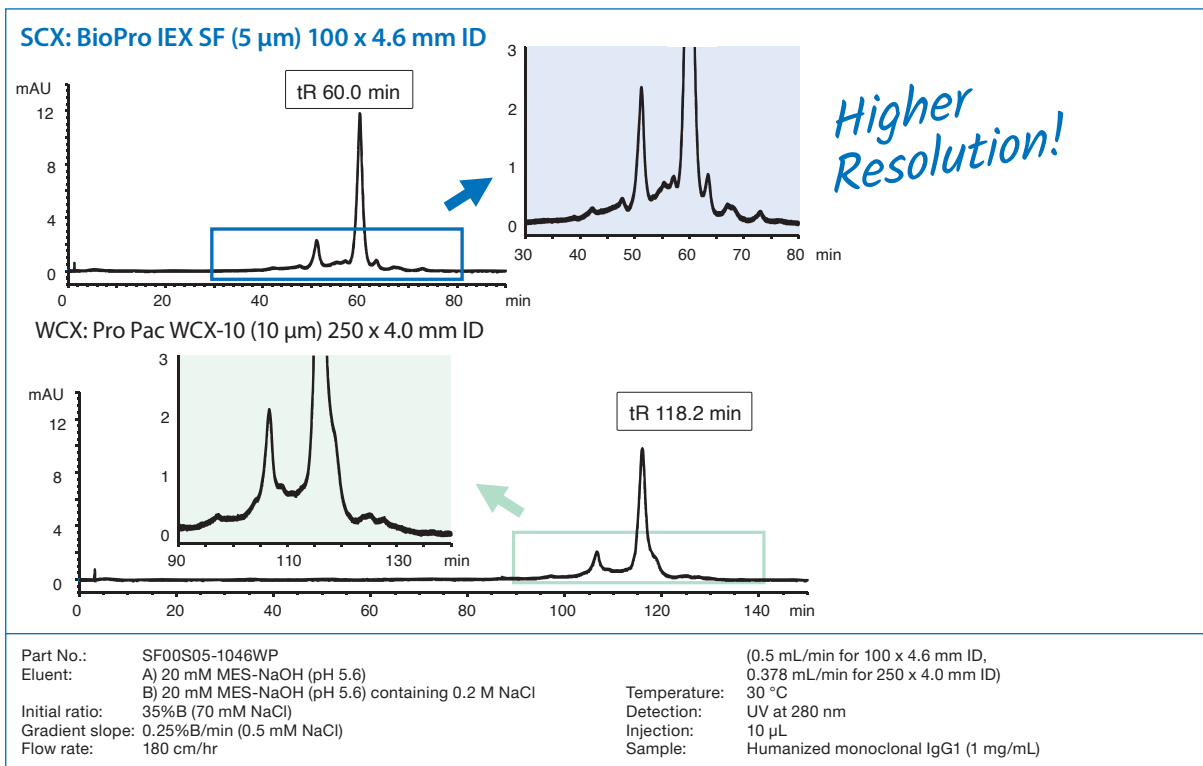
Peptide mapping of tryptic digests of BSA with enhanced sensitivity



Part No.: QAA0S05-0546WP
 Eluent: A) 20 mM Tris-HCl (pH 8.6)
 B) 20 mM Tris-HCl (pH 8.6) containing 0.5 M NaCl
 Gradient: 0–15%B (0–30 min), 15–60%B (30–60 min)
 Flow rate: 0.5 mL/min
 Temperature: 25 °C
 Detection: UV at 220 nm
 Injection: 20 μ L
 Sample: Tryptic digest of BSA

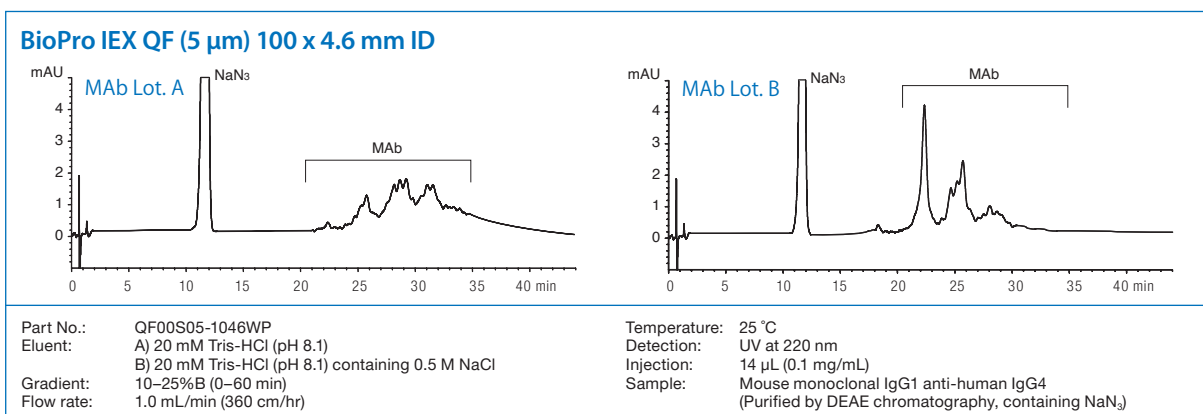
IEX – BioPro IEX: Antibody analysis

Monoclonal antibody analysis with non-porous cation exchange columns



The separation of MAb is compared using a strong cation (BioPro IEX SF) and a weak cation exchange column (ProPac WCX-10) under the same gradient conditions at pH 5.6. BioPro IEX SF can achieve a higher resolution of MAB than the competitor's column in a shorter analysis time.

QC of monoclonal antibodies with non-porous BioPro IEX QF



Two different batches of commercially available MAB purified by DEAE chromatography were analyzed on a BioPro IEX QF column (100 mm length). The MAB was separated into several peaks, and the batch-to-batch variability is observed. The BioPro IEX QF/SF 100 mm length columns, which have high efficiency, are ideal for characterization of glycoproteins, such as monoclonal antibodies, and for quality control assessment of biopharmaceuticals.

Optimisation of oligonucleotide separations on ion exchange chromatography

Non-porous anion exchange column is generally suitable for analysis of oligonucleotides. For optimisation of single-stranded DNA and RNA of about 20 mer some conditions such as type of mobile phase and column temperature can be changed.

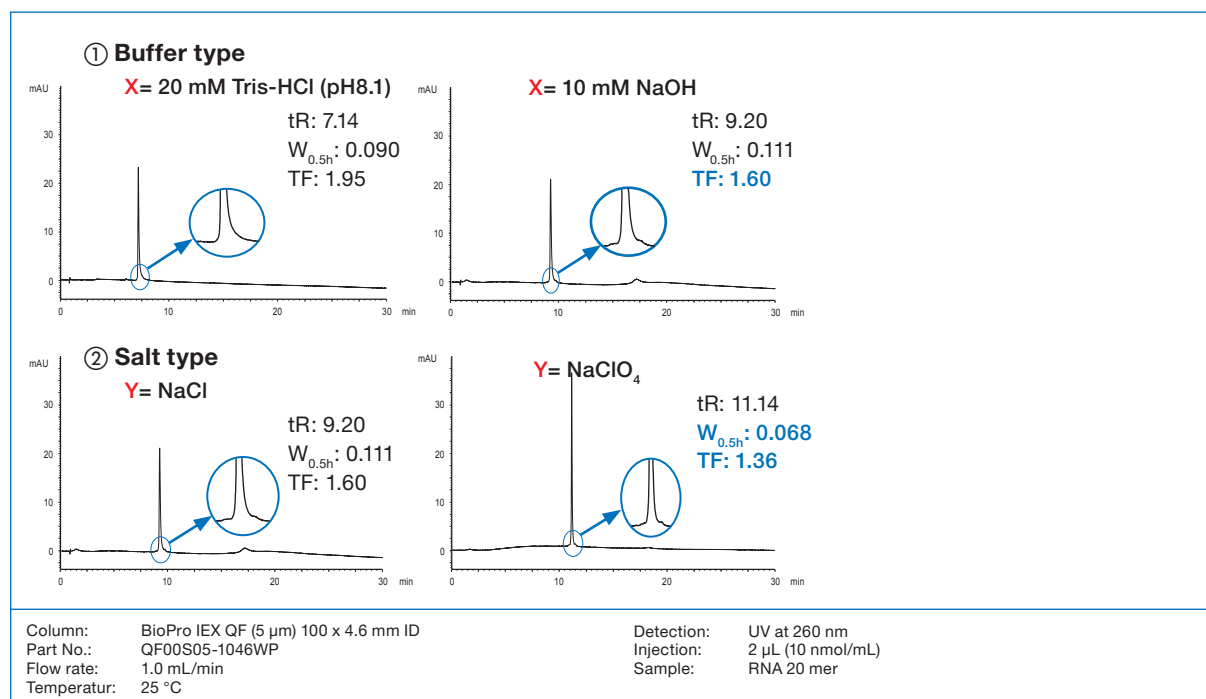
1 Improvement of peak tailing

Sample Group 1 (Phosphodiester oligonucleotides; PO)

| | | |
|---|-----------------------------|--|
| 1 | Single-stranded DNA (ssDNA) | 5'-TCATCACACTGAATACCAAT-3' (DNA 20 mer) |
| 2 | Single-stranded DNA (ssDNA) | 5'-GTCATCACACTGAATACCAAT-3' (DNA 21 mer) |
| 3 | Single-stranded RNA (ssRNA) | 5'-UCAUCACACUGAAUACCAU-3' (RNA 20 mer) |
| 4 | | 5'-GUCAUCACACUGAAUACCAU-3' (RNA 21 mer) |
| 5 | Single-stranded RNA (ssRNA) | 5'-U(M)C(M)A(M)U(M)C(M)A(M)C(M)A(M)C(M)U(M)G(M)A(M)A(M)U(M)A(M)C(M)C(M)A(M)A(M)U(M)-3' (2'-OMe RNA 20 mer) |
| 6 | | 5'-G(M)U(M)C(M)A(M)U(M)C(M)A(M)C(M)A(M)C(M)U(M)G(M)A(M)A(M)U(M)A(M)C(M)C(M)A(M)A(M)U(M)-3' (2'-OMe RNA 21 mer) |

N(M)=2'-OMe RNA

By changing the buffer from 20 mM Tris-HCl (pH 8.1) to 10 mM NaOH, the tailing factor for an oligonucleotide is reduced. Furthermore, the peak tailing is further suppressed when NaClO₄ was added to 10 mM NaOH instead of NaCl.



2 Improvement of carryover

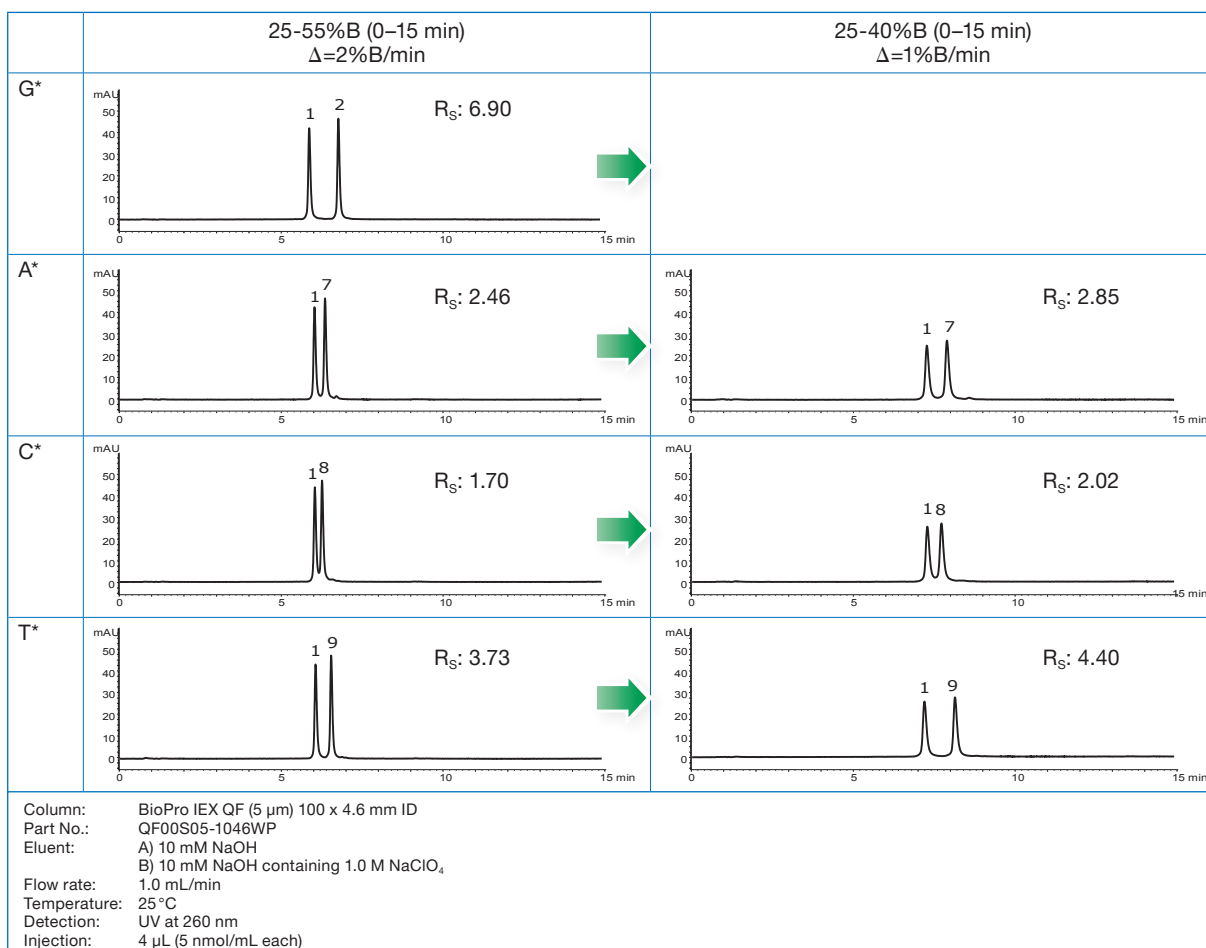
When the initial gradient concentration of NaCl is low (ex. 50 mM), carryover is observed. By increasing the initial gradient concentration of NaCl up to 400 carryover can be avoided with good reproducibility.

IEX – Expert Tips: Oligonucleotides

3 Improvement of ssDNA separation with single-base differences (differing in the type of base of 5'end of DNA 21mer)

When ssDNAs with single-base differences (differing in the type of base of 5'end of DNA 21mer) is analysed under the described condition, all of peak separations got worse. By using a shallower gradient, better separations could be achieved.

| | | |
|---|---------------------|--|
| 1 | Single-stranded DNA | 5'-TCATCACACTGAATACCAAT-3' (DNA 20 MER) |
| 2 | | 5'-GTCATCACACTGAATACCAAT-3' (DNA 21 MER) |
| 7 | | 5'-ATCATCACACTGAATACCAAT-3' (DNA 21 MER) |
| 8 | | 5'-CTCATCACACTGAATACCAAT-3' (DNA 21 MER) |
| 9 | | 5'-TTCATCACACTGAATACCAAT-3' (DNA 21 MER) |



*base of 5'end of DNA 21 mer

4 Improvement of the separation of phosphorothioate oligonucleotides with single-base differences in length

Since acidity of all PS is much higher than all PO, a higher salt concentration is required for elution. The peak of all PS is much broader because it is thought that all PS contains as many as 219 (524,288) stereoisomers. A steeper gradient curve, increasing column temperature and adding organic solvent can improve peak separation. However, increase of organic solvent ratio gave little improvement in peak separation.

IEX – Ordering information

3 µm non-porous analytical columns, PEEK hardware (max. pressure 250 bar)

| Phase | Column ID [mm] | Column length [mm] | | | | Precolumn filter 2 µm* |
|---------------|----------------|--------------------|----------------|----------------|----------------|---------------------------|
| | | 30 (250 bar) | 50 (250 bar) | 100 (250 bar) | 150 (250 bar) | |
| | | | | | | (pack of 5) |
| BioPro IEX QF | 4.6 | QF00S03-0346WP | QF00S03-0546WP | QF00S03-1046WP | QF00S03-1546WP | XRPRCP25 |
| BioPro IEX SF | 4.6 | SF00S03-0346WP | SF00S03-0546WP | SF00S03-1046WP | SF00S03-1546WP | |

5 µm non-porous analytical columns, PEEK hardware (max. pressure 60–120 bar)

| Phase | Column ID [mm] | Column length [mm] | | | | Precolumn filter 2 µm* |
|---------------|----------------|--------------------|----------------|----------------|----------------|---------------------------|
| | | 30 (60 bar) | 50 (100 bar) | 100 (120 bar) | 150 (120 bar) | |
| | | | | | | (pack of 5) |
| BioPro IEX QF | 4.6 | QF00S05-0346WP | QF00S05-0546WP | QF00S05-1046WP | QF00S05-1546WP | XRPRCP25 |
| BioPro IEX SF | 4.6 | SF00S05-0346WP | SF00S05-0546WP | SF00S05-1046WP | SF00S05-1546WP | |

5 µm porous analytical columns, PEEK hardware (max. pressure 25–35 bar)

| Phase | Column ID [mm] | Column length [mm] | | | Precolumn filter 2 µm* |
|---------------|----------------|--------------------|----------------|----------------|---------------------------|
| | | 30 (25 bar) | 50 (30 bar) | 100 (35 bar) | |
| | | | | | (pack of 5) |
| BioPro IEX QA | 4.6 | QAA0S05-0346WP | QAA0S05-0546WP | QAA0S05-1046WP | XRPRCP25 |
| BioPro IEX SP | 4.6 | SPA0S05-0346WP | SPA0S05-0546WP | SPA0S05-1046WP | |

* Holder required, part no. XRPRCP02

6 µm non-porous semiprep. columns, stainless steel hardware (max. pressure 30–90 bar)

| Phase | Column ID [mm] | Column length [mm] | |
|---------------|----------------|--------------------|--|
| | | 100 | |
| BioPro IEX QF | 10 | QF00S06-1010WT | |
| | 20 | QF00S06-1020WT | |
| | 30 | QF00S06-1030WT | |
| BioPro IEX SF | 10 | SF00S06-1010WT | |
| | 20 | SF00S06-1020WT | |
| | 30 | SF00S06-1030WT | |

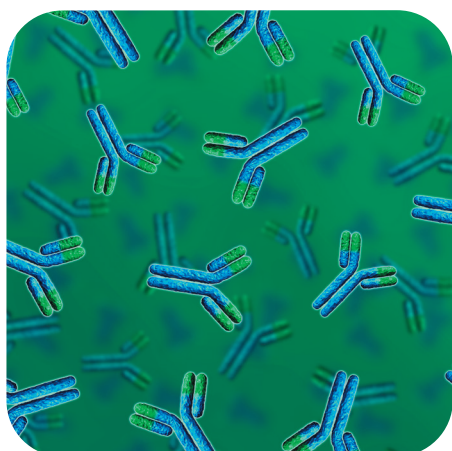
6 µm porous semiprep. columns, stainless steel hardware (max. pressure 40 bar)

| Phase | Column ID [mm] | Column length [mm] | |
|---------------|----------------|--------------------|--|
| | | 100 | |
| BioPro IEX QA | 10 | QAA0S06-1010WT | |
| | 20 | QAA0S06-1020WT | |
| BioPro IEX SP | 10 | SPA0S06-1010WT | |
| | 20 | SPA0S06-1020WT | |

Other dimensions on demand



HIC



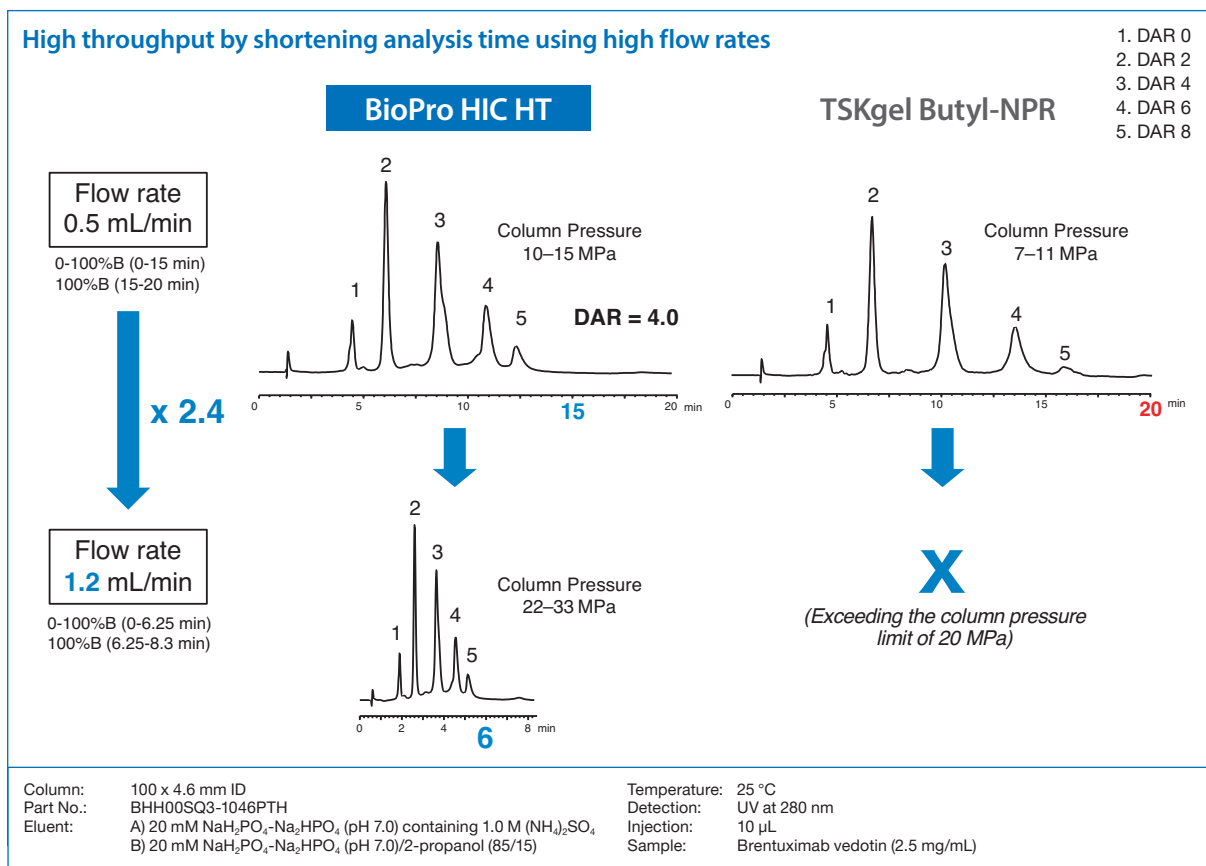
HIC – BioPro Series

- Specifically designed for drug-to-antibody conjugates (ADCs) and antibodies
- Ideal drug-to-antibody ratio (DAR) analysis
- High throughput by reducing analysis time
- Excellent batch-to-batch reproducibility
- Long term stability

| | BioPro HIC HT | BioPro HIC BF |
|-------------------------------|--|--|
| Base particle | hydrophilic polymer (polymethacrylate) | hydrophilic polymer (polymethacrylate) |
| Particle size / μm | 2.3 | 4 |
| Pore | non-porous | non-porous |
| Functional group | butyl | butyl |
| pH range | 2–12 | 2–12 |
| Pressure limit | 40 MPa (5,800 psi) | 20 MPa (2,900 psi) |
| Temperature range | 10–60°C | 10–60°C |

High column stability

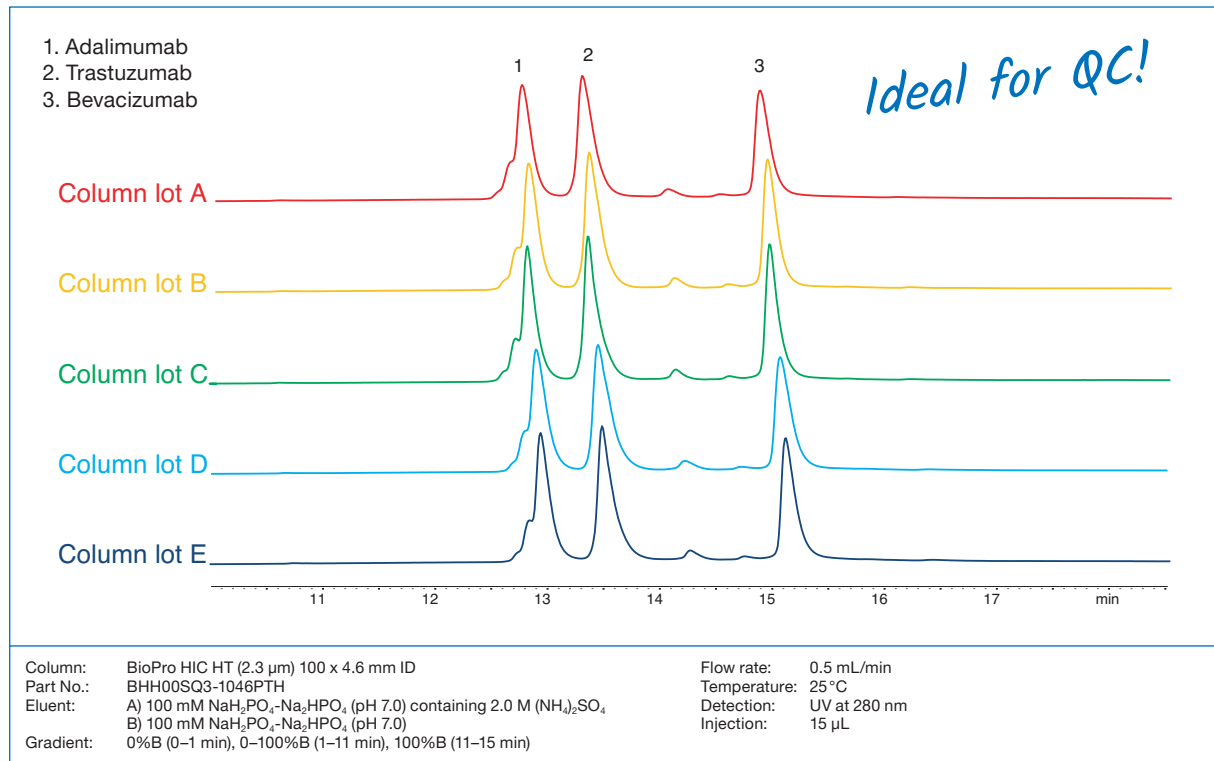
High throughput by shortening analysis time using high flow rates



BioPro HIC HT improves analysis throughput of ADCs by 2–3 times with an excellent Drug-to-Antibody Ratio (DAR). The rapid analysis is possible without loss of resolution. Competitor HIC columns fail under these conditions.

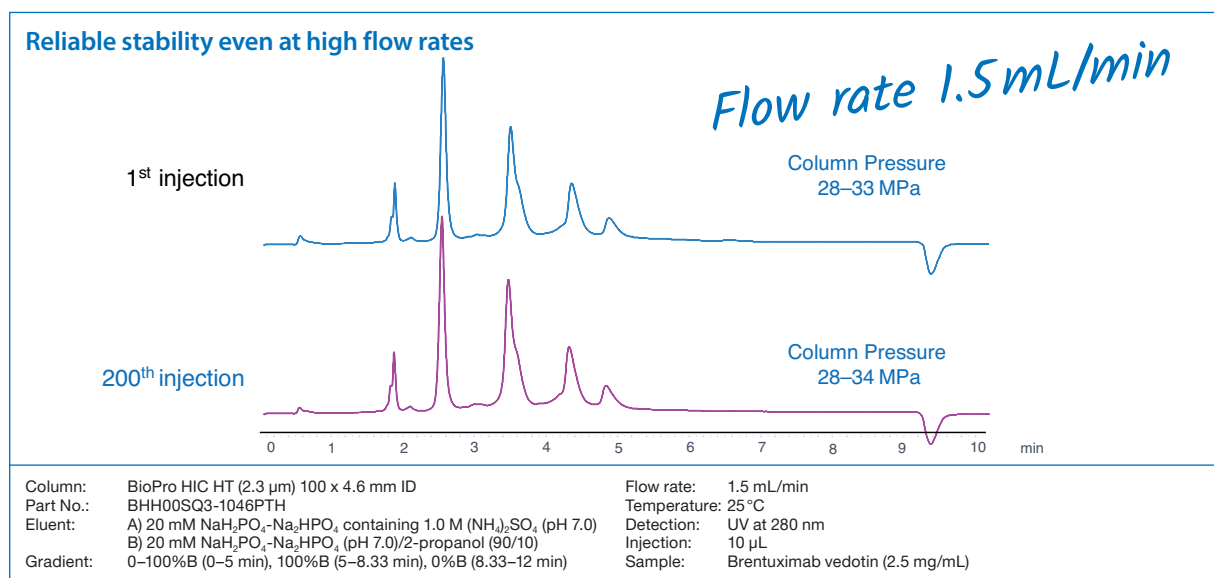
HIC – BioPro HIC: Reproducibility & stability

Excellent batch-to-batch reproducibility



BioPro HIC HT exhibits an excellent batch-to-batch reproducibility making it the ideal choice for quality control analysis of biopharmaceuticals such as MAbs.

Exceptional stability



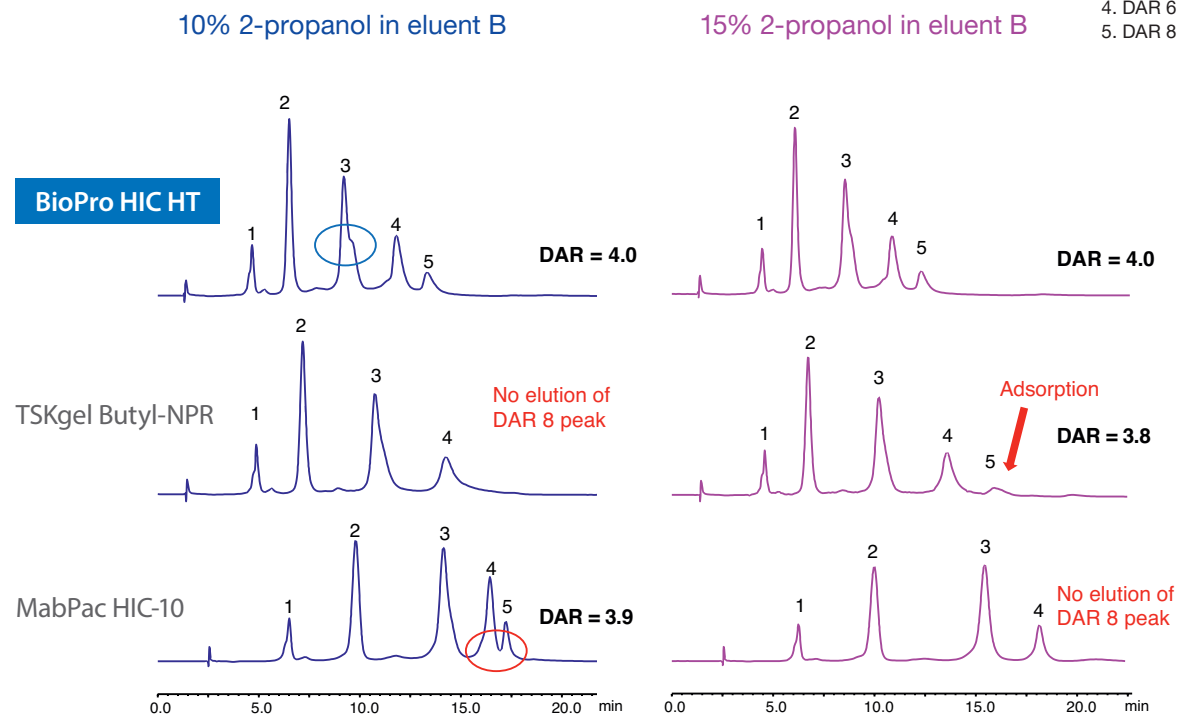
BioPro HIC HT offers excellent stability under high flow rates/high pressure conditions due to its unique rigid particle and optimised column packing technology.

HIC – BioPro HIC: ADC analysis

Designed for analysis of ADCs

Novel surface chemistry for drug-to-antibody ratio (DAR) analysis

1. DAR 0
2. DAR 2
3. DAR 4
4. DAR 6
5. DAR 8



Column: 100 x 4.6 mm ID
 Part No.: BHH00SQ3-1046PTH
 Eluent: A) 20 mM NaH₂PO₄-Na₂HPO₄ (pH 7.0) containing 1.0 M (NH₄)₂SO₄
 B) 20 mM NaH₂PO₄-Na₂HPO₄ (pH 7.0)/2-propanol (90/10) or (85/15)
 Gradient: 0–100%B (0–15 min), 100%B (15–20 min), 0%B (20–35 min)
 Flow rate: 0.5 mL/min
 Temperature: 25 °C
 Detection: UV at 280 nm
 Injection: 10 µL
 Sample: Brentuximab vedotin (2.5 mg/mL)

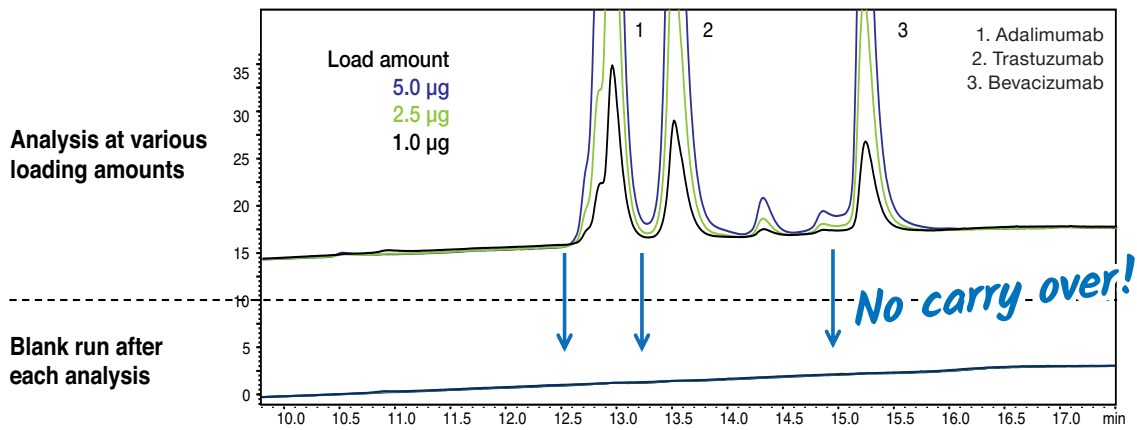
BioPro HIC HT offers higher resolution than conventional HIC columns. Its surface modification suppresses excessive or too strong adsorption of ADCs and results in highly reliable quantification. With varying 2-propanol content, all peaks are completely eluted from the BioPro HIC HT column with high resolution. Another peak is partially separated from peak 3. Additionally, the same DAR values are observed at any content of 2-propanol.

BioPro HIC HT offers:

- Higher resolution than conventional HIC columns
- Highly reliable quantification
- Flexible method development

Excellent recovery and virtually no carry over

Highly accurate quantification of ADCs and antibodies



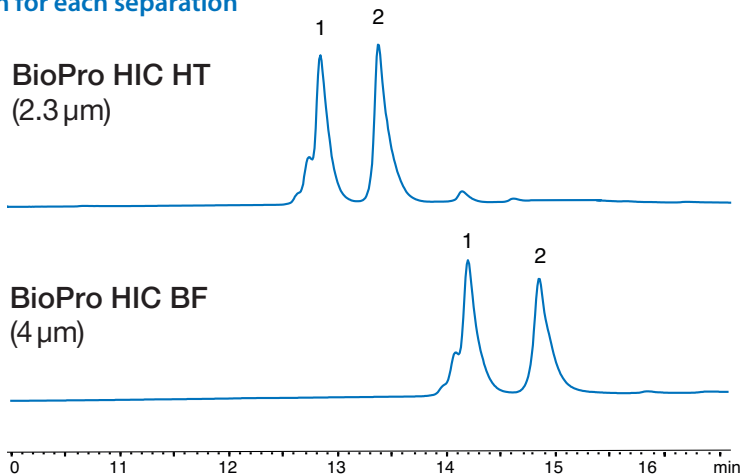
Column: BioPro HIC HT (2.3 µm) 100 x 4.6 mm ID
 Part No.: BHH00SQ3-1046PTH
 Eluent: A) 100 mM NaH₂PO₄-Na₂HPO₄ (pH 7.0) containing 2.0 M (NH₄)₂SO₄
 B) 100 mM NaH₂PO₄-Na₂HPO₄ (pH 7.0)

Gradient: 0%B (0–1 min), 0–100%B (1–11 min), 100%B (11–15 min)
 Flow rate: 0.5 mL/min
 Temperature: 25 °C
 Detection: UV at 280 nm

BioPro HIC HT offers higher linearity over wide loading and virtually no carryover. This contributes to highly accurate quantitation of ADCs and antibodies.

Different hydrophobicity

The right column for each separation



Column: 100 x 4.6 mm ID
 Part Nos.: BHH00SQ3-1046PTH
 BHB00S04-1046WT
 Eluent: A) 100 mM NaH₂PO₄-Na₂HPO₄ (pH 7.0) containing 2.0 M (NH₄)₂SO₄
 B) 100mM NaH₂PO₄-Na₂HPO₄ (pH 7.0)
 Gradient: 0%B (0–1 min), 0–100%B (1–11 min), 100%B (11–15 min)

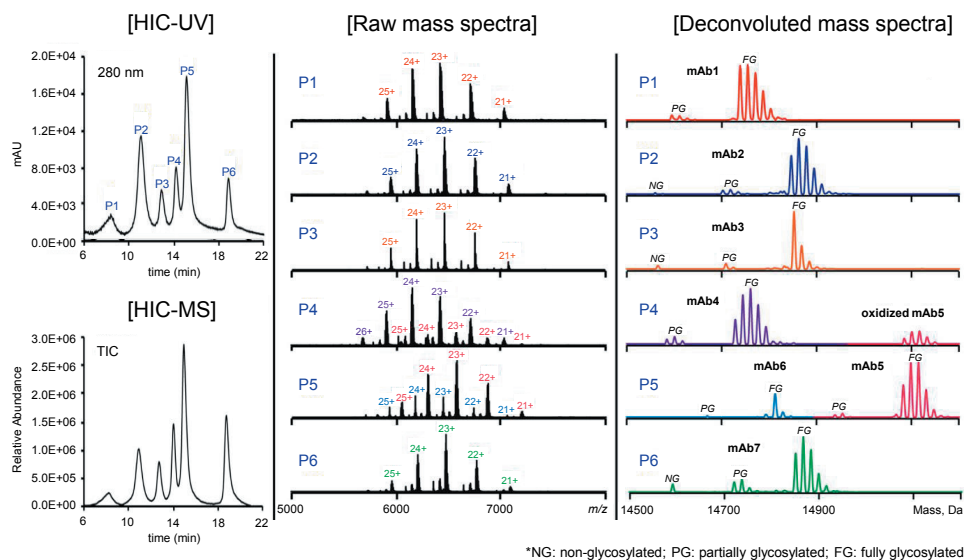
Flow rate: 0.5 mL/min
 Temperature: 25 °C
 Detection: UV at 280 nm
 Injection: 15 µL
 Sample: 1. Adalimumab (Humira®; 0.5 mg/mL)
 2. Trastuzumab (Herceptin®; 0.5 mg/mL)

BioPro HIC HT is the first choice for ADCs or MABs. BioPro HIC BF columns show a stronger retention and can therefore be used for the separation of low hydrophobic proteins or especially for the analysis of oxidised MABs.

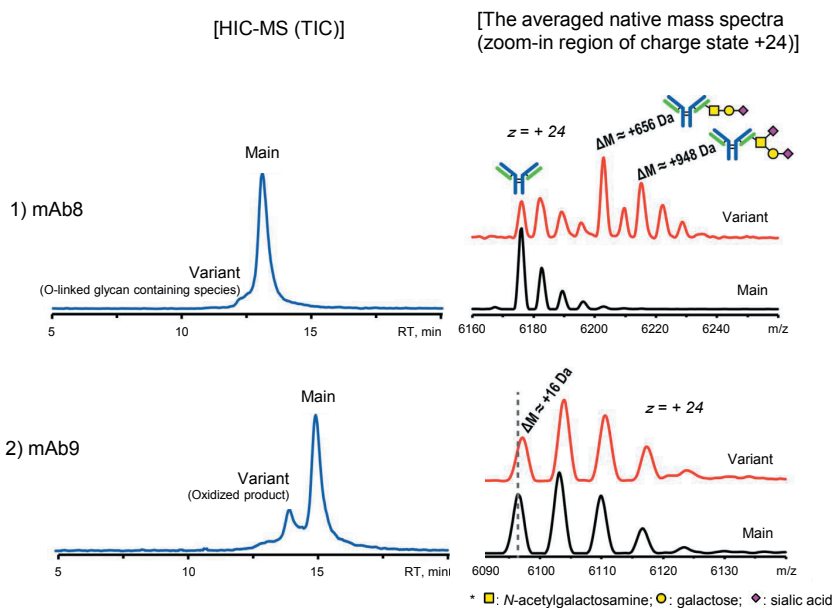
HIC – BioPro HIC: Direct HIC-MS coupling

Online native HIC-MS analysis of MABs and their molecular variants

Separation of an antibody mixture of seven different MABs



Separation of two MABs from their molecular variants



Column: BioPro HIC BF (4 μm) 100 x 4.6 mm ID
 Part No.: BHB00S04-1046WT
 Eluent: A) 3 M ammonium acetate in water
 B) 100 % water
 Gradient: 0% B (0–2 min), 0–90% B (2–18 min), 90% B (18–22 min)
 Flow rate: 0.3 mL/min
 Temperature: ambient
 Detection: UV at 280 nm, NSI-MS

Injection: MAB mixture: 3 μL (3–6 μg)
 MAB 8 and MAB 9: 10 μg each
 Sample: Mixture of 7 in-house MABs at 1–2 mg/mL each
 2 in-house MABs with molecular variants
 Setup: Post-column makeup flow:
 100 % water at 1.5 mL/min (reducing salt conc. 6-fold)
 Splitter to reduce the flow rate to 1–5 μL/min

Courtesy by S. Wang, Regeneron Pharmaceuticals Inc.

To enable simultaneous UV and MS detection a post-column makeup flow and a splitter were used. The makeup flow decreases the salt concentration while the splitter reduces the flow rate to enable the coupling to MS. A nanospray ionisation (NSI) was chosen because of its high sensitivity and salt tolerance.

Reference: Y. Yan, T. Xing, S. Wang, T. J. Daly, N. Li, Online coupling of analytical hydrophobic interaction chromatography with native mass spectrometry for the characterization of monoclonal antibodies and related products, *J. Pharm. Biomed. Anal.* 186 (2020) 113313.

The influence of salts in HIC separations

The technique known as hydrophobic interaction chromatography is a mode of chromatography that separates proteins by differences in surface hydrophobicity. [1] This method utilises reversible interactions that occur between protein molecules and hydrophobic stationary phase ligands attached to the particle surface.

Certain non-denaturing salts are used to improve the hydrophobic interactions between proteins and the stationary phase. The mobile phase is typically an aqueous solution of salts such as ammonium sulfate or sodium chloride and a buffer to control pH (usually phosphate

buffer between pH 6 and 7). The Hofmeister series of lyotropic and chaotropic ions shown below in Fig. 1 provides a template for salt selection. High concentrations of salt, particularly ammonium sulfate, may precipitate proteins; therefore, solubility should be checked under the initial gradient (binding) conditions. The strength of the interaction between the protein and stationary phase decreases with decreasing salt gradient (see Fig. 2). Another option is a change of pH which results in an increase in the charge on the protein due to the ionisation of acidic groups.

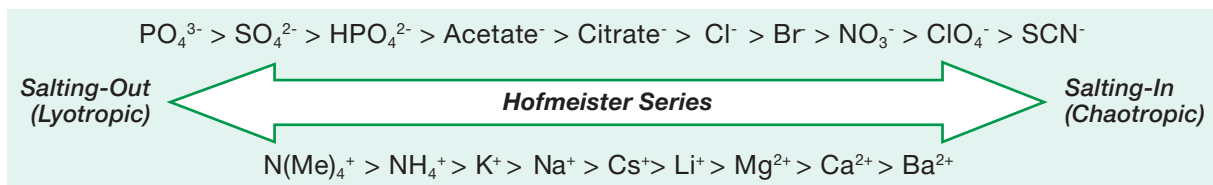


Fig. 1: The Hofmeister Series of lyotropic and chaotropic ions.

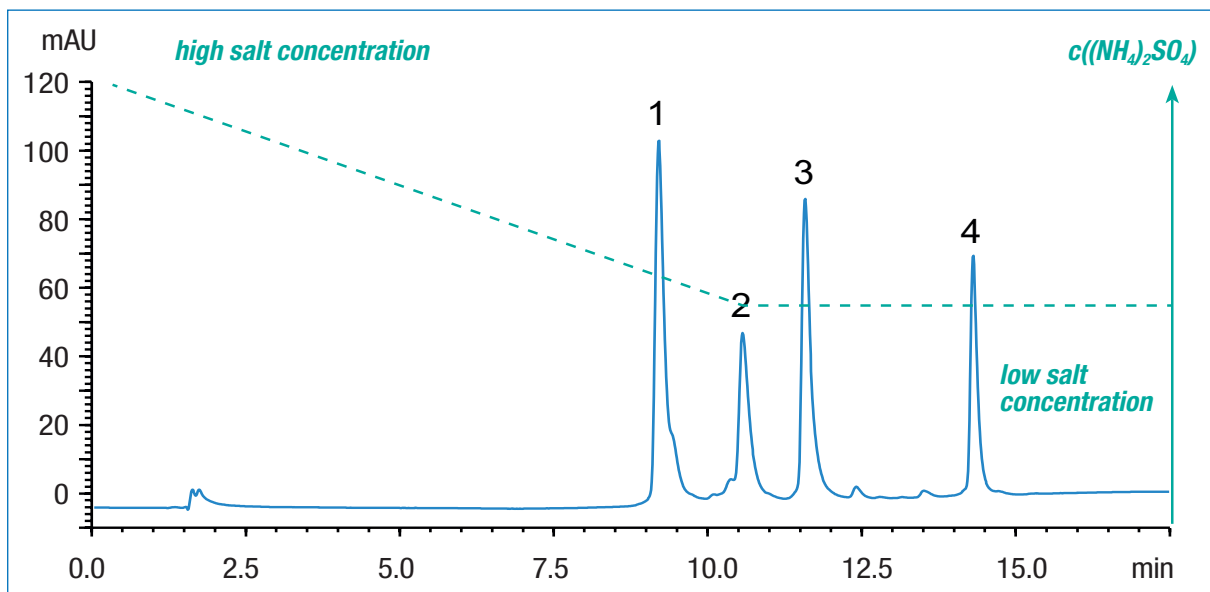


Fig. 2: Method with decreasing salt gradient.

Column: BioPro HIC BF (100 x 4.6 mm ID)
 Part No.: BHB00S04-1046WT
 Eluent: A) 100 mM NaH₂PO₄-Na₂HPO₄ (pH 7.0) containing 2.0 M (NH₄)₂SO₄
 B) 100 mM NaH₂PO₄-Na₂HPO₄ (pH 7.0)
 Flow rate: 0.5 mL/min
 Gradient: 0–100%B (0–11 min), 100%B (11–15 min)
 Temperature: 25°C
 Detection: UV at 280 nm
 Injection: 15 µL

Samples: 1. Myoglobin (0.73 mg/mL)
 2. Ribonuclease A (0.75 mg/mL)
 3. Lysozyme (0.25 mg/mL)
 4. α-Chymotrypsinogen A (0.25 gm/mL)

HIC is particularly effective when used to separate proteins and monoclonal antibodies. The separation of monoclonal antibodies (MAb), MAb aggregates and glycosylated MAbs can be achieved due to their specific hy-

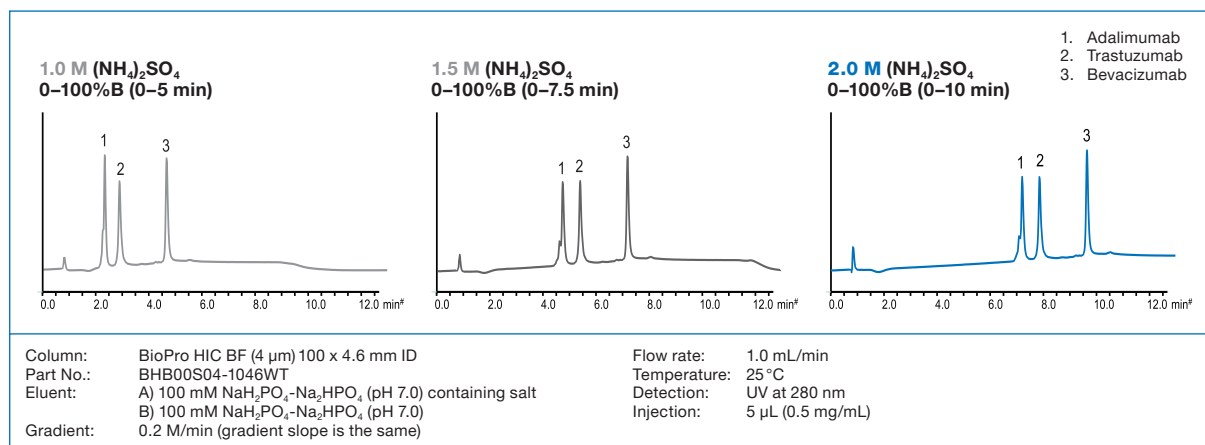
drophobic properties. It also provides an excellent method for determination of drug-to-antibody ratios (DAR) in antibody-drug conjugates (ADCs).

[1] Queiroza, J.A.; Tomaza, C.T.; Cabral, J.M.: Hydrophobic interaction chromatography of proteins, J Biotechnol. 2001, 87, 143-159.

HIC – Expert Tips: Separation factors

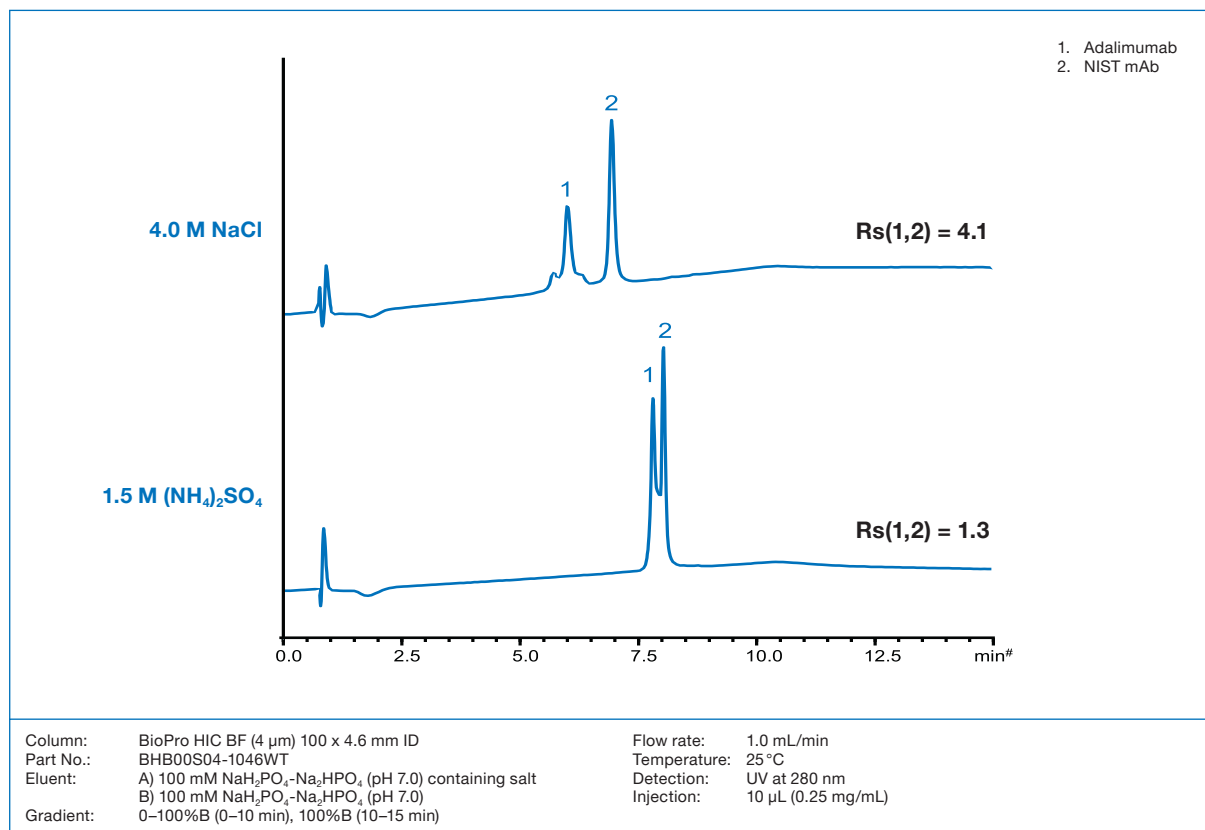
Effect of initial salt concentration

Buffers containing $(\text{NH}_4)_2\text{SO}_4$ are often used as a mobile phase in HIC mode because $(\text{NH}_4)_2\text{SO}_4$ has a strong salt-ing-out effect. The higher the initial concentration of $(\text{NH}_4)_2\text{SO}_4$, the stronger will be the retention of proteins. Therefore, a buffer with a high salt concentration is more suitable for the separation of low hydrophobic proteins with weak retention.



Influence of the type of salt

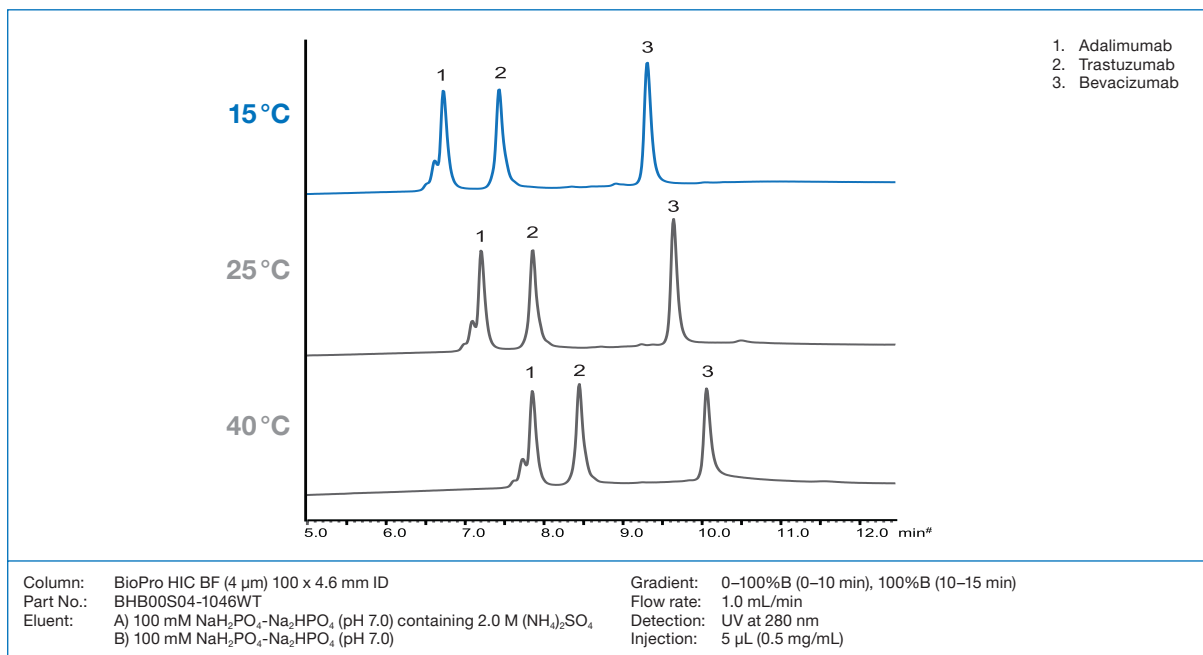
NaCl and $\text{CH}_3\text{COONH}_4$ are also used as buffer salts. The separation selectivity varies with the type of salt used in some cases, so changing the type of salt can also be effective when the separation is not sufficient. However, these salts have to be used at very high concentrations to gain retentions comparable to $(\text{NH}_4)_2\text{SO}_4$. Attention needs to be paid to the prevention of precipitation of salts in the buffer and damage of the LC system.



HIC – Expert Tips: Separation factors / Ordering information

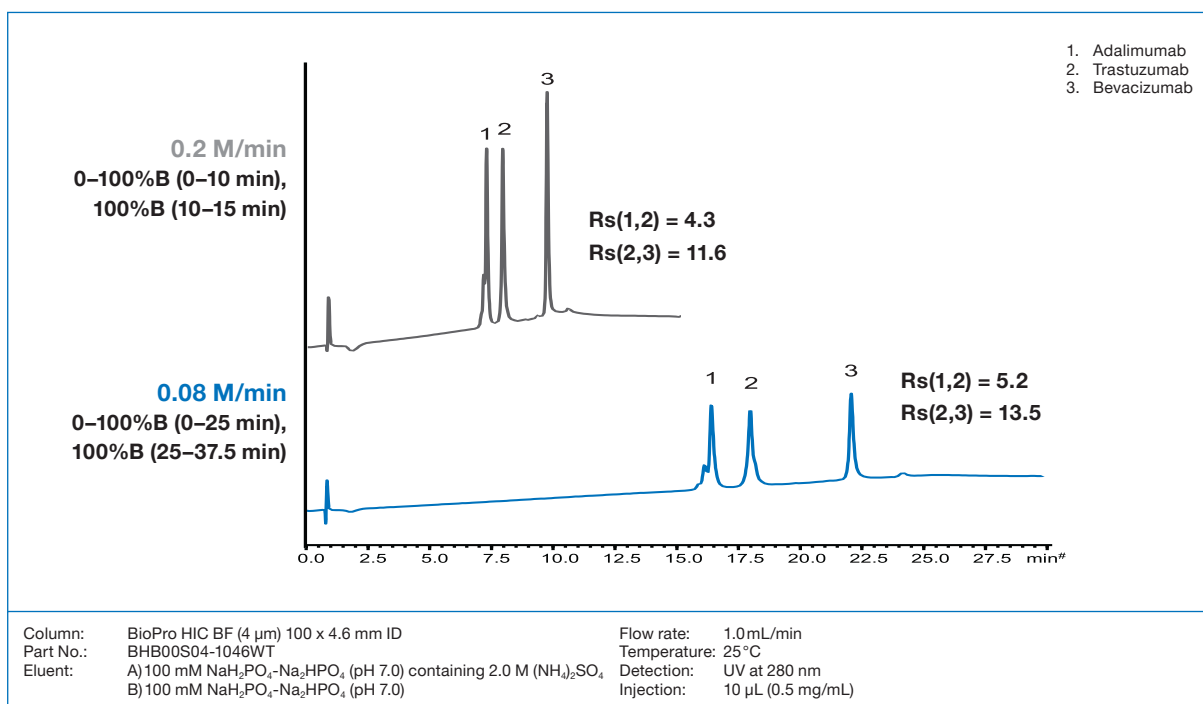
Temperature influence

In HIC mode, higher temperatures result in longer retention times of proteins. This assumes that the hydrophobic area interacting with the stationary phase becomes larger due to a change in the structure of proteins with increasing temperature, so that the hydrophobic interactions become stronger.



Variation of gradient slope

In general, shallower gradients improve the separation and the resulting resolution.

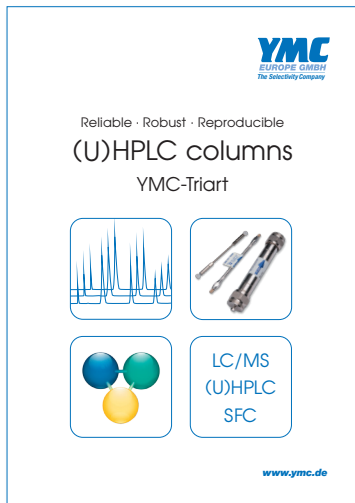


Ordering information

Ordering information

| Phase | Particle size [µm] | Column ID [mm] | Column length [mm] | Part number | Precolumn filter 2 µm* (pack of 5) |
|---------------|--------------------|----------------|--------------------|------------------|---------------------------------------|
| BioPro HIC HT | 2.3 | 4.6 | 100 | BHH00SQ3-1046PTH | XRPRCS35 |
| | 2.3 | 4.6 | 33 | BHH00SQ3-H346PTH | XRPRCS35 |
| BioPro HIC BF | 4 | 4.6 | 100 | BHB00S04-1046WT | XRPRCS35 |

*Holder required, part no XRPRCS03
Other dimensions on demand



YMC-Triart



YMC-Triart metal-free



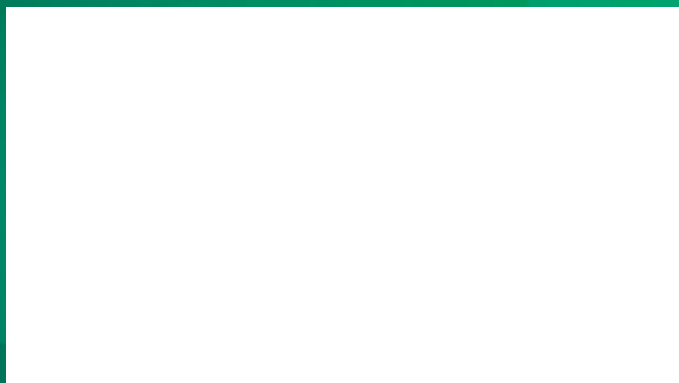
BioPro IEX Resins

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